



Mechanism and Consequences of Extracellular Adenosine Accumulation in the Hypoxic Hippocampal Slice

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Thesis submitted for the degree of Doctor of Philosophy
in
The University of Adelaide
(Faculty of Science)

submitted through the Department of Physiology, July 1995

David Doolittle 1995

David Doolittle 1995

TABLE OF CONTENTS

Figures and tables	viii
Abbreviations and chemical names	x
Abstract	xii
1. Introduction and overview	1
1.1. General overview of cerebral hypoxia.....	1
1.2. Aims.....	3
2. Involvement of adenosine in hypoxia in the hippocampal slice: background literature review.....	5
2.1. Introduction.....	5
2.2. Hippocampal formation	5
2.2.1. Anatomy and connections of the hippocampal formation	5
2.2.1.1. General cortical organisation	5
2.2.1.2. Architectonics of the hippocampal formation	6
2.2.1.2.1. Entorhinal area	7
2.2.1.2.2. Parasubiculum and presubiculum	8
2.2.1.2.3. Subiculum.....	8
2.2.1.2.4. Hippocampus.....	8
2.2.1.2.5. Pyramidal neurons	9
2.2.1.2.6. Dentate gyrus.....	9
2.2.1.3. Major pathways of the hippocampal formation	10
2.2.1.3.1. Alveus.....	10
2.2.1.3.2. Schaffer collaterals	10
2.2.1.3.3. Mossy fibres.....	10
2.2.1.3.4. Perforant path.....	11
2.2.1.3.5. Commissural connections.....	11
2.2.1.3.6. Associative pathways.....	12
2.2.1.3.7. Lamellar organisation	12
2.2.1.4. External connections of the hippocampal formation	13
2.2.1.4.1. Afferents.....	13
2.2.1.4.2. Efferents	14
2.2.1.5. Local circuits neurons in the CA ₁ area.....	14
2.2.1.5.1. Recurrent inhibition	16
2.2.1.5.2. Feed-forward inhibition	17
2.2.2. Electrophysiology	18
2.2.2.1. Hippocampal slices and whole animal preparations.....	18
2.2.2.2. Anatomy of the recording site in the CA ₁	19
2.2.2.3. Extracellular recordings	20
2.2.2.4. Intracellular recording	21
2.2.2.4.1. Spikes	22
2.2.2.4.2. IPSP	22
2.2.2.4.3. EPSP	23
2.2.2.4.4. Pyramidal neuron voltage activated calcium and potassium currents	23
2.2.2.4.5. Interneuron electrophysiology	24
2.2.2.5. Inhibitory synapses	26
2.2.2.5.1. Postsynaptic inhibition	26
2.2.2.5.2. Presynaptic inhibition.....	27
2.2.2.6. Excitatory synapses	27

2.2.2.6.1. AMPA receptors.....	28
2.2.2.6.2. NMDA receptor-ionophore	28
2.2.2.6.3. Glutamate inactivation.....	29
2.2.2.7. Other neurotransmitters and neuromodulators.....	29
2.3. Adenosine.....	30
2.3.1. Adenosine receptor classification	30
2.3.1.1. A ₁ receptors	32
2.3.1.2. A ₃ receptors	33
2.3.1.3. A ₂ receptors.....	33
2.3.2. Adenosine A ₁ receptors mediate depression of synaptic transmission and of neuronal excitability in the CA ₁	34
2.3.2.1. Adenosine inhibition in the hippocampus is mediated by A ₁ receptors	35
2.3.2.2. Presynaptic actions of adenosine A ₁ receptors	35
2.3.2.2.1. Adenosine inhibits neurotransmitter release	35
2.3.2.2.2. Adenosine A ₁ receptor inhibition of N-type calcium channels	37
2.3.2.2.3. Adenosine modulation of presynaptic potassium currents.....	37
2.3.2.3. Postsynaptic actions of adenosine A ₁ receptors.....	38
2.3.2.3.1. Membrane hyperpolarisation	38
2.3.2.3.2. Slow after-hyperpolarisation.....	39
2.3.2.3.3. Other postsynaptic A ₁ receptor actions.....	40
2.3.3. Adenosine A ₂ receptors	40
2.3.3.1. Excitatory actions of adenosine receptor agonists.....	40
2.3.3.2. Post-inhibitory hyperexcitability	41
2.3.3.3. Adenosine A _{2a} receptor-mediated excitation.....	42
2.3.3.4. Adenosine A _{2b} receptor enhancement of P-type calcium channels	43
2.3.3.5. Presynaptic and postsynaptic mechanisms of adenosine hyperexcitability	43
2.3.3.5.1. Presynaptic mechanisms	43
2.3.3.5.2. Postsynaptic mechanisms.....	44
2.3.4. Adenosine receptor second messenger systems	44
2.3.4.1. G proteins.....	44
2.3.4.2. G protein linked adenosine receptors modulate adenylate cyclase.	45
2.3.4.3. Adenosine A ₁ receptor inhibitory actions are G protein linked.....	45
2.3.4.3.1. G _o proteins mediate A ₁ receptor neuronal inhibition	46
2.3.4.4. Adenosine A _{2a} receptor is G protein linked	47
2.3.4.5. Phospholipase C	47
2.3.5. Adenosine metabolism and release.....	47
2.3.5.1. Adenosine formation by 5'-nucleotidase.....	47
2.3.5.1.1. Heart and liver.....	48
2.3.5.1.2. Brain	49
2.3.5.2. Adenosine release.....	50
2.3.5.2.1. Basal adenosine release	50
2.3.5.2.1.1. Basal adenosine levels.....	51
2.3.5.2.2. Adenosine accumulation secondary to presynaptic nucleotide release	51
2.3.5.2.3. Adenosine release with depolarisation	52
2.3.5.2.4. Adenosine release during hypoxia	53
2.3.5.3. Adenosine action is terminated by uptake and catabolism	54
2.4. Hypoxia.....	55

2.4.1. Hypoxia and ischaemia.....	55
2.4.2. Energy metabolism during hypoxia, ischaemia and hypoglycaemia	56
2.4.2.1. Mitochondrial respiration	56
2.4.2.2. Glycolysis	56
2.4.2.3. Enzymes catalysing nucleotide metabolism.....	58
2.4.2.4. Hypoxic nucleotide metabolism	59
2.4.3. Hypoxic release of amino acid transmitters	61
2.4.4. Excitotoxicity.....	62
2.4.4.1. Acute neuronal damage	62
2.4.4.2. Delayed neuronal damage	63
2.4.4.3. Glutamate receptors mediate excitotoxicity.....	64
2.4.5. Hypoxic and ischaemic neuronal damage	64
2.4.5.1. Selective vulnerability of the hippocampus	64
2.4.5.2. Synaptic release of amino acids mediates hypoxic neuronal death.	65
2.4.5.3. Delayed neuronal death	66
2.4.5.4. Irreversible loss of synaptic transmission in hippocampal slices....	66
2.4.6. Hypoxic inhibition of synaptic transmission	67
2.4.7. Hypoxic actions on postsynaptic neurons.....	69
2.4.7.1. Hypoxic hyperpolarisation.....	69
2.4.7.1.1. ATP-sensitive potassium channels and hypoxic neuronal inhibition.....	70
2.4.7.2. hypoxic spreading depression.....	71
2.4.8. Relationship of neuronal depression and energy metabolism.....	72
2.4.9. Adenosine and hypoxic neuronal inhibition.....	73
2.4.10. Adenosine protects against hypoxia, ischaemia and excitotoxicity	74
2.4.10.1. Neuronal survival	74
2.4.10.2. Adenosine modulation of ischaemic excitatory amino acid release	75
2.4.10.2.1. A ₁ receptor inhibition of excitatory amino acid release	75
2.4.10.2.2. A ₂ receptor enhancement of excitatory amino acid release...	76
2.4.10.2.3. A ₃ receptors	76
2.4.11. Post-hypoxic hyperexcitability	76
2.5. Summary and conclusions.....	79
3. Methods	80
3.1. Hippocampal slice preparation.....	80
3.1.1. Tissue preparation	80
3.1.2. Recording chamber.....	80
3.1.3. ACSF	81
3.1.4. Diffusion	82
3.1.4.1. Oxygen	83
3.1.4.2. Membrane impermeant substances	84
3.2. Electrophysiological recordings	84
3.2.1. Rationale for use of field potential recordings.....	84
3.2.2. Instrumentation of the slice	86
3.2.3. Data acquisition and analysis.....	87
3.2.3.1. Recording of evoked field potentials	87
3.2.3.2. Field potential measurements	87
3.2.3.3. Input/output curves.....	89
3.2.3.4. Time course studies.....	91
3.3. Presentation and statistical analysis of data	92
3.4. Ethical considerations	92

4. Acute actions of hypoxia	93
4.1. Introduction	93
4.2. Methods.....	94
4.3. Results	94
4.3.1. Hypoxia.....	94
4.3.1.1. Acute effects of hypoxia	94
4.3.1.2. Input/output curves	95
4.3.1.3. Miscellaneous hypoxic exposures	96
4.3.2. Carbon monoxide	96
4.3.3. 2,4-Dinitrophenol	100
4.3.4. Adenosine A ₁ receptor antagonism and hypoxia	101
4.3.4.1. Acute effects of hypoxia during A ₁ receptor antagonism	101
4.3.4.2. Input/output curves.....	103
4.3.5. Adenosine A ₁ receptor activation with exogenous agonists	104
4.3.6. Adenosine A ₁ receptors, but not ATP-sensitive potassium channels, contribute to 2,4 dinitrophenol-induced depression of synaptic transmission	104
4.3.7. Calculation of adenosine concentration during hypoxia.....	105
4.4. Discussion	109
4.4.1. Hypoxia is well tolerated in the hippocampal slice	109
4.4.2. Carbon monoxide	111
4.4.3. Dinitrophenol	113
4.4.4. Hypoxic depression of synaptic transmission is due to A ₁ receptor activation.....	113
4.4.5. Postsynaptic depression of pyramidal neurons during hypoxia.....	115
4.4.6. Adenosine is not necessary for post-hypoxic neuronal survival in this model.....	116
4.4.7. Summary and conclusions.....	117
5. Mechanism of hypoxic adenosine accumulation	119
5.1. Introduction	119
5.2. methods.....	120
5.3. results	121
5.3.1. Mitochondrial uncoupling.....	121
5.3.2. Electron carrier inhibition.....	122
5.3.3. Inhibition of oxidative phosphorylation	125
5.3.3.1. ATP synthetase inhibition.....	125
5.3.3.2. Mitochondrial nucleotide transporter inhibition.....	127
5.3.4. Reducing ATP depletion.....	128
5.3.4.1. Inhibition of ATPase activity.....	128
5.3.4.2. ATP buffering with phosphocreatine.....	130
5.3.5. Inhibitors of nucleotide metabolism	130
5.3.6. Intracellular calcium stores.....	132
5.3.7. Nucleoside transporter inhibition blocks adenosine uptake but not release.....	134
5.3.8. Adenosine uptake and catabolism	134
5.4. Discussion	135
5.4.1. Respiration.....	136
5.4.2. Energy deprivation	137
5.4.2.1. Electron transport inhibitors	137
5.4.2.2. Uncoupling	138
5.4.2.3. Mitochondrial poisons selectively inhibit synaptic transmission ..	139

5.4.3. Signal for adenosine release.....	139
5.4.3.1. ATP depletion is not responsible for initial inhibition of synaptic transmission	140
5.4.3.1.1. Oxidative phosphorylation	140
5.4.3.1.2. ATP depletion	141
5.4.3.1.3. ATP buffering with phosphocreatine	141
5.4.3.1.4. Hypoglycaemia	142
5.4.4. Adenosine metabolic pathways.....	143
5.4.4.1. S-adenosylhomocysteine hydrolase	143
5.4.4.2. 5'-nucleotidase	144
5.4.4.3. Nucleoside transporter.....	145
5.4.4.4. Adenosine kinase	145
5.4.4.5. Adenosine deaminase.....	146
5.4.5. Activation of enzymes	147
5.4.5.1. Mitochondrial signals	147
5.4.5.2. Increase in AMP	147
5.4.5.3. Enzyme activation	148
5.4.5.4. pH	149
5.4.5.5. Calcium	149
5.4.5.5.1. Mitochondrial calcium homeostasis.....	149
5.4.5.5.2. Calcium evoked hypoxic current	151
5.5. Summary and conclusions.....	152
6. Persistent actions of hypoxia and adenosine: post-inhibitory and post-hypoxic hyperexcitability	153
6.1. Introduction	153
6.2. Methods	154
6.3. Results.....	155
6.3.1. Post-hypoxic hyperexcitability	155
6.3.1.1. Prolonged hypoxia resulted in hyperexcitability upon reoxygenation	155
6.3.1.2. 2,4-Dinitrophenol induced hyperexcitability	156
6.3.1.3. Adenosine A ₁ receptor antagonism	158
6.3.1.4. Input/output curve changes after hypoxia.....	160
6.3.2. Adenosine A ₁ receptor inhibition is not reduced following hypoxia....	163
6.3.3. Adenosine receptor agonists during normoxia	163
6.3.3.1. Adenosine caused rebound hyperexcitability	163
6.3.3.2. Low dose adenosine did not produce multiple population spikes	163
6.3.3.3. A ₁ and A ₂ receptor agonists did not produce multiple population spikes	164
6.3.3.4. Input/output curve changes after adenosine receptor agonists.....	166
6.3.4. Glutamate receptors in post-hypoxic hyperexcitability.....	167
6.3.4.1. NMDA receptor antagonism	168
6.3.4.2. trans-ACDP.....	169
6.3.5. Protein kinase	170
6.4. Discussion	172
6.4.1. Post-hypoxic hyperexcitability is an increase in postsynaptic neuronal excitability	172
6.4.2. Subset of neurons become hyperexcitable.....	176
6.4.3. Glutamate receptors and long-term potentiation	177
6.4.4. Hyperexcitability during onset of hypoxia	178
6.4.5. Adenosine mediated hyperexcitability.....	178

6.4.6. Post-hypoxic hyperexcitability is not mediated by A ₁ receptors.....	180
6.4.7. Second messengers	180
6.4.7.1. Second messenger regulation of potassium channels.....	180
6.4.7.2. Receptor mediated hyperexcitability	181
6.4.7.2.1. β -adrenoceptor and cAMP mediated hyperexcitability.....	182
6.4.7.2.2. Cholinergic receptor mediated hyperexcitability	183
6.4.7.3. Second messengers in post-hypoxic hyperexcitability.....	184
6.5. Summary and conclusions.....	185
7. Concluding remarks	186
7.1. General summary of experimental work	186
7.1.1. Chapter 4.....	186
7.1.2. Chapter 5.....	187
7.1.3. Chapter 6.....	187
7.2. Future directions	188
7.3. framework of early hypoxic events including the findings of the present study	189
7.4. Relevance of the present findings	190
7.4.1. Neuronal death and hypoxia	190
7.4.1.1. Hypoxia and ischaemia.....	190
7.4.1.2. Is post-hypoxic hyperexcitability required for delayed neuronal death?	192
7.4.1.3. Therapeutics and drug design.....	192
7.4.2. Adenosine release	193
7.4.3. Application of field potential recordings and pharmacological methods to the study of hypoxia in the hippocampal slice.....	194
7.4.3.1. Success of field potential studies	194
7.4.3.2. Advantages and limitations of field potential recordings	194
Appendix A: Publications during candidature	196
Bibliography.....	197

ABSTRACT

This thesis examines the alterations in electrophysiological function during hypoxia in the rat hippocampal slice, in particular those alterations induced by extracellular accumulation of adenosine. Evaluation of electrophysiological responses in the *in vitro* rat hippocampal slice is a standard model for neurophysiological investigations, and has been used extensively in the study of hypoxia. Rat hippocampal CA₁ pyramidal neurons are considered to be selectively vulnerable to hypoxic damage; however, adenosine A₁ receptor-mediated depression of excitatory synaptic transmission during hypoxia may protect neurons. Enhanced excitability of neurons is reported following hypoxia, and is proposed to contribute to neuronal damage.

In the present studies, the post-hypoxic recovery of synaptically evoked field potentials was used to measure the survival of CA₁ pyramidal neurons, and to assess any functional alterations precipitated by hypoxia. Excitatory synaptic transmission in the hippocampal CA₁ area was depressed during 30 minutes of hypoxia due to activation of adenosine A₁ receptors, this depression being sensitive to A₁ receptor antagonism. Synaptic transmission always recovered upon reoxygenation, indicating that CA₁ neurons survived prolonged inhibition of respiration, and that adenosine A₁ receptor activation during hypoxia was not necessary for this neuronal recovery. Following prolonged hypoxia, recovered postsynaptic potentials manifested a permanent hyperexcitability, obvious as multiple population spikes. A method for quantifying this hyperexcitability was developed, which established that it results from a postsynaptic alteration in CA₁ neuronal excitability. Similar hyperexcitability also followed normoxic adenosine exposure, but was not produced by agonists for the A₁ or A₂ adenosine receptor subtypes. Apparently, adenosine activation of an unidentified receptor subtype produces postsynaptic hyperexcitability, and this may be the mechanism of hyperexcitability following hypoxic adenosine accumulation.

This work also examined the mechanism of adenosine accumulation during hypoxia, by assessing A₁ receptor-mediated depression of synaptically evoked field potentials. Hypoxia, respiratory inhibitors and mitochondrial uncoupling agents all caused an adenosine induced depression of synaptic transmission, whereas inhibitors of oxidative phosphorylation did not. Furthermore, alleviation of ATP depletion failed to increase the latency of synaptic depression during mitochondrial uncoupling. It seems likely, therefore, that the massive adenosine accumulation during energy deprivation is not a direct consequence of ATP depletion, but is most likely a result of stimulation of cytosolic 5'-nucleotidase and inhibition of adenosine kinase, possibly by changes in free AMP, pH or cytosolic calcium.