

# Intrauterine programming of leptin

Jason Elliot Ekert, B.Sc. (Hons)

A thesis submitted to Adelaide University, South Australia for the degree of Doctor of Philosophy

Department of Obstetrics and Gynaecology Adelaide University South Australia

October 2000

# TABLE OF CONTENTS

Table of contents	ii
Abstract	xi
Statement	xiv
Acknowledgements	xv
List of figures	xvii
List of tables	xx
List of Abbreviations	xxi
Publications arising from this thesis	xxii
CHAPTER 1	
INTRODUCTION	
1.1 Introduction	1-2
1.2 Regulation of adipocyte development	
1.2.1 Preadipocyte development	1-3
1.2.2 Clonal expansion	1-5
1.2.3 Expression of adipocyte genes	1-5
1.2.4 Terminal differentiation	1-7
1.2.5 Adipocyte development in the fetal pig	1-8
1.3 Discovery of a satiety signal	1-9
1.4 Leptin gene and protein	1-11

1.5 Regulation of leptin action	
1.5.1 Discovery of the leptin receptor	1-15
1.5.2 Structure and activity of the leptin receptor	1-16
1.5.3 Expression of leptin receptor isoforms	1-18
1.5.4 The movement of leptin across the blood-brain barrier	1-18
1.6 Actions of leptin	1-20
1.6.1 Leptin and regulation of body weight	1-20
1.6.2 Leptin, neuropeptides and appetite regulation	
1.6.2.1 Neuropeptide Y	1-21
1.6.2.2 Melanocortins	1-24
1.6.3 Leptin, metabolic rate and thermogenesis	1-26
1.6.4 Leptin and the adrenal system	1-27
1.6.5 Leptin and skeletal muscle	1-29
1.6.6 Leptin and hematopoiesis	1-30
1.6.7 Leptin and reproduction	1-30
1.6.7.1 Pregnancy	1-31
1.6.7.2 Leptin at birth	1-32
1.6.7.3 Leptin and the placenta	1-32
1.6.7.4 Leptin and postnatal growth and development	1-34
1.6.7.5 Leptin and puberty	1-35
1.7 Regulation of leptin expression	1-36
1.7.1 Nutrition	1-36
1.7.2 Sympathetic nervous system	1-37
1.7.3 Glucocorticoids	1-37

٠	
i	۲.

1.7.4 Insulin	1-37	
1.7.5 Peroxisome proliferator activated receptor-γ	1-40	
1.7.6 Growth hormone and Insulin-like growth factor-I	1-41	
1.7.7 Thyroid axis	1-41	
1.7.8 Gonadal steroids	1-42	
1.8 Intrauterine programming		
1.8.1 Definition	1-42	
1.8.2 Intrauterine programming of adiposity	1-43	
1.8.3 Intrauterine programming of muscle development	1-45	
1.8.4 Intrauterine programming of endocrine systems	1-45	
1.8.5 Factors influencing fetal programming- programming mechanisms	1-46	
1.9 Aims of this study	1-47	
CHAPTER 2		
EXPERIMENTAL ANIMAL MODELS OF LEPTIN PROGRAMMING IN HUMANS		
2.1 Introduction	2-2	
2.2 Materials and Methods		
2.2.1 Identification of guinea pig leptin mRNA		
2.2.1.1 Extraction of total RNA from guinea pig adipose tissue	2-3	
2.2.1.2 Analysis of total RNA content	2-4	
2.2.1.3 Reverse Transcription of guinea pig adipose RNA	2-5	
2.2.1.4 DNA extraction	2-6	

2.2.1.5 PCR amplification of guinea pig leptin cDNA	2-7
2.2.1.6 Gel electrophoresis of PCR products	2-9
2.2.2 Sequencing of the guinea pig leptin PCR product	2-9
2.2.3 PCR amplification of guinea pig leptin cDNA using	
guinea pig specific primers	2-10
2.2.4 Northern analysis of guinea pig and pig leptin mRNA	2-10
2.2.5 Identification of pig leptin mRNA	
2.2.5.1 Extraction of RNA from porcine subcutaneous adipose tissue	2-13
2.2.5.2 Integrity and concentration of porcine adipose RNA	2-14
2.2.5.3 Reverse Transcription of porcine adipose RNA	2-14
2.2.5.4 Polymerase chain reaction amplification of	
porcine adipose cDNA	2-15
2.2.5.5 Gel electrophoresis of porcine RT PCR products	2-15
2.2.5.6 Sequencing of porcine leptin RT PCR products	2-15
2.3 Results and Discussion	
2.3.1 Detection of guinea pig leptin mRNA by RT PCR	2-16
2.3.2 Partial sequencing of the guinea pig leptin cDNA	2-21
2.3.3 Detection of leptin mRNA in adipose tissue from	
adult and fetal guinea pigs	2-25
2.3.4 Detection of guinea pig leptin mRNA by Northern analysis	2-25
2.3.5 Detection of leptin mRNA in porcine adipose tissue by RT PCR	2-26
2.4 Conclusion	2-29

#### CHAPTER 3

# DEVELOPMENT OF A QUANTITATIVE RT PCR ELISA TO MEASURE PORCINE ADIPOSE TISSUE LEPTIN MRNA

3.1 Introduction	3-2
3.2 Materials and Methods	
3.2.1 RT PCR Digoxigenin Enzyme Linked Immunosorbant Assay (ELISA)	
3.2.1.1 DIG-labelling of RT PCR products from pig adipose tissue	3-3
3.2.1.2 General procedure for ELISA	3-3
3.2.1.3 Immobilisation of DIG-labelled PCR product by	
biotinylated oligonucleotide "capture probe"	3-4
3.2.2 Conditions for amplification of DIG-labelled leptin and $\beta$ -actin	
cDNA fragments by PCR and detection by DIG-ELISA	3-6
3.2.3 Assay specificity	3-7
3.2.4 Anti-DIG-polymerized horse-radish peroxidase antibody	
in the ELISA	3-7
3.2.5 DIG-UTP in the PCR	3-7
3.2.6 Effect of DIG-labelled PCR product in the ELISA	3-8
3.2.7 Amplification efficiency of the leptin and $\beta$ -actin RT PCR DIG-ELISA	3-8
3.2.8 Routine RT PCR DIG-ELISA quantitation	3-10
3.2.8.1 Calibration of ELISA	3-10
3.2.9 Assay Precision	3-11

3.3 Results and Discussion	
3.3.1 Immobilisation of DIG-labelled PCR product by	
biotinylated oligonucleotide	3-11
$3.3.2$ PCR cycle profile of leptin and $\beta$ -actin RT PCR DIG-ELISA	3-12
3.3.3 Assay specificity	3-15
3.3.4 Anti-DIG-polymerized horse-radish peroxidase antibody	
in the ELISA	3-15
3.3.5 Effect of concentration of DIG-labelled leptin PCR product in the ELISA	3-19
3.3.6 Effect of concentration of dNTP mixture containing DIG-UTP	
in the leptin PCR ELISA	3-19
3.3.7 Amplification efficiency of the leptin and $\beta$ -actin RT PCR DIG-ELISA	3-19
3.3.8 Calibration of ELISA	3-22
3.3.9 Assay precision	3-22
3.4 Conclusion	3-22
CHAPTER 4	
LEPTIN EXPRESSION IN OFFSPRING IS PROGRAMMED BY NUTRITION	N IN
PREGNANCY	
4.1 Introduction	4-2
4.2 Materials and Methods	
4.2.1 Animals	4-3
4.2.2 Extraction of total RNA from porcine adipose tissue	4-3

	4.2.3 Integrity and concentration of porcine adipose RNA	4-4
	4.2.4 Reverse Transcription of RNA from porcine adipose tissue	4-4
	4.2.5 Polymerase chain reaction amplification of porcine adipose cDNA	4-4
	4.2.6 Measurement of DNA content of progeny adipose tissue	4-5
	4.2.7 Measurement of RNA content of progeny adipose tissue	4-6
	4.2.8 Measurement of protein content of progeny adipose tissue	4-7
	4.2.9 Measurement of water and lipid of progeny adipose tissue	4-8
	4.2.10 Plasma leptin radioimmunoassay	4-8
	4.2.11 Statistics	4-10
4.3 Re	esults	
	4.3.1 Effect of maternal nutrition during pregnancy on progeny	4-10
	4.3.2 Effect of maternal nutrition during pregnancy on progeny	
	adipocyte characteristics	4-10
	4.3.3 Effect of maternal nutrition during pregnancy on	
	leptin expression in progeny	4-14
	4.3.4 Body weight at birth and postnatal leptin expression	4-15
4.4 Di	iscussion	4-15

#### CHAPTER 5

ENVIRONMENT DURING PREGNANCY PROGRAMS LEPTIN, INSULIN-LIKE GROWTH FACTOR-II, TRIIODOTHYRONINE AND ESTRADIOL IN PROGENY

5.1 Introduction 5-2

5.2 M	aterials and methods	
	5.2.1 Animals	5-4
	5.2.2 Analysis of metabolites and hormones	5-5
	5.2.3 Measurement of DNA content of progeny adipose tissue	5-5
	5.2.4 Measurement of RNA content of progeny adipose tissue	5-6
	5.2.5 Measurement of protein content of progeny adipose tissue	5-6
	5.2.6 Measurement of water and lipid of progeny adipose tissue	5-6
	5.2.7 Statistical analysis	5-6
5.3 R	esults	
	5.3.1 Effects on pregnant dams	5-7
	5.3.2 Effects on progeny	
	5.3.2.1 Size at birth and growth of progeny	5-11
	5.3.2.2 Adipose tissue content of progeny	5-12
	5.3.2.3 Circulating metabolites and hormones of progeny	5-12
	5.3.2.4 Relationships between postnatal growth and maternal measures	5-16
	5.3.2.5 Relationships between progeny circulating metabolites/	
	hormones and maternal measures	5-17
5.4 D	iscussion	5-19

## CHAPTER 6

## GENERAL DISCUSSION

6.2 Intrauterine programming of leptin by maternal nutrition	6-4
6.3 Maternal metabolism is associated with intrauterine programming of leptin in pigs	6-7
6.4 Potential mechanisms of intrauterine programming of leptin	6-8
6.5 Implications of high postnatal plasma leptin levels in progeny	6-10
6.6 Clinical significance	6-12
6.6 Animal production implications	6-13
6.7 Future directions	6-14

# Bibliography

#### ABSTRACT

Many epidemiological studies published over the last ten years have indicated that environment during pregnancy affects adult phenotype and health of offspring. The permanent postnatal effects caused by environmental factors during human pregnancy have been termed *in utero* programming. Babies which are shorter or lighter at birth show catchup growth during early postnatal life, develop reduced insulin sensitivity and increased risks of obesity. As adults, they also have higher incidence of diabetes and cardiovascular disease and increased concentrations of leptin in their blood. Leptin is a polypeptide produced by adipose tissue and secreted into blood, that acts to suppress appetite and increase energy expenditure. The guinea pig and pig were evaluated as experimental animal models in which to investigate mechanisms of *in utero* leptin programming in humans. Adipocyte development is more advanced at birth in guinea pigs, pigs and humans than in rodents.

The first aim was to determine whether leptin is expressed in adipose tissue of pigs and guinea pigs as is the case in humans. A leptin cDNA fragment was produced from guinea pig adipose RNA and found to have a nucleotide sequence with greater than 80% identity to leptin genes of human, rat, mouse, pig and cow. Leptin mRNA was detected in several adipose sites in the guinea pig and expression was higher in the adult than the fetus. A partial leptin cDNA was also produced from pig adipose tissue RNA and found to have a nucleotide sequence identical to that concurrently published for porcine leptin cDNA. Leptin mRNA was also detected in subcutaneous adipose tissue of pigs. The pig was chosen to investigate leptin programming because an assay for measuring leptin protein in plasma from this species was available whereas an equivalent assay in the guinea pig was not available.

Increased leptin production in humans is associated with obesity and larger adipocytes. Obesity associated with larger adipocytes in adult rats can be programmed by undernutrition during the first two-thirds of pregnancy. A study of long term outcome from the Dutch famine in the winter of 1944-1945 also found that nutritional restriction during the first half of pregnancy was associated with increased adult obesity in offspring. This lead to the concept that the fetus is susceptible to programming of tissues and endocrine systems during certain phases that affects the subsequent adult phenotype. The relationship between birth weight and adult leptin levels could be hypothetically due to altered adipocyte development in growth-retarded fetuses.

The second quarter of gestation is a critical period for adipocyte developent in the pig, a period of adipocyte commitment and development. A change in adipocyte numbers or characteristics during this period could lead to permanent changes in leptin production postnatally. I therefore hypothesised that the leptin axis in offspring is programmed by maternal nutrition during pregnancy and investigated whether leptin production in offspring is altered by maternal nutrition during the second quarter of pregnancy in pigs. I found that body weight at birth and at ~8.5 weeks of age was unaffected by the level of feed during this period of pregnancy. However, leptin mRNA abundance in adipose tissue (p=0.015) and plasma leptin concentration (p=0.01) were higher in progeny from mothers provided with more feed in the second quarter of pregnancy. Leptin protein concentration in plasma was correlated with leptin mRNA abundance in adipose tissue in these animals.

Growth hormone treatment during pregnancy alters maternal metabolism, especially increasing maternal glucose. This mimics diabetic pregnancy in humans which increases the transfer of glucose to the fetus. The increased glucose delivery to the fetus at a critical stage of adipocyte development might affect adipocyte development or endocrine systems that regulate leptin production. I hypothesised that maternal hyperglycemia would alter leptin programming. To determine whether maternal glucose or other ciculating metabolites are involved in intrauterine leptin programming, pigs were treated with growth hormone in the second quarter of pregnancy and the effects on maternal metabolites and progeny levels of leptin and leptin-regulating hormones were measured. Treatment with GH increased maternal plasma insulin, IGF-I and glucose concentrations. Weight of offspring at birth was not affected. GH treatment (p<0.005) during the second quarter of pregnancy increased plasma leptin concentrations in 61 day old progeny. Treatment with GH in pregnancy also increased triiodothyronine (p=0.002) and estradiol (p=0.002) and decreased IGF-II concentrations (p=0.009) in plasma from 61 day progeny.

Programming of postnatal leptin production by maternal environment in pregnancy is likely to be due to an increase in the availability of glucose to the fetus. A direct mechanism of programming leptin expression could be through glucose altering the UDP-N-acetylglucosamine pathway in the preadipocytes in the fetus or indirectly through the actions of fetal insulin or insulin-like growth factor-I on fetal adipocyte maturation. Also leptin may be programmed indirectly through the actions of fetal or placental leptin, insulin or insulin-like growth factor-I on fetal hypothalamic maturation.