

NOVEL INDUCIBLE PHYTOCHEMICAL DEFENCES AGAINST PLANT PARASITIC NEMATODES

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Table of Contents

SUMMARY	i
DECLARATION	jv
PUBLICATION ARISING FROM THIS THESIS	v
ACKNOWLEDGEMENTS	vi
CHAPTER 1. INTRODUCTION	1
CHAPTER 2. REVIEW OF LITERATURE	5
2.1 INTRODUCTION	5
2.2 NEMATODES AS PESTS	5
2.3 NEMATODE CONTROL 2.4 PLANT DEFENCES TO NEMATODES	6 6
2.4.1 Hypersensitive response	7
2.4.2 Preformed chemical defenses	8
2.4.3 Inducible chemical defenses	8
Phytoalexins	9
Proteinase inhibitors	9
Pathogenesis related proteins	10
Phytohormones	10
2.5 ECDYSTEROIDS 2.5.1 History	11 12
2.5.1 Phistory 2.5.2 Chemistry and structure	14
2.6 ECDYSTEROIDS IN PLANTS	17
2.6.1 Distribution	17
2.6.2 Biosynthesis	18
2.6.3 Functions	19
2.6.4 Induction	21
2.7 ECDYSTEROIDS IN NEMATODES	22
2.7.1 Occurrence	22 23
2.7.2 Biosynthesis 2.7.3 Biological activities	23
2.8 FLAVONOIDS IN PLANTS	26
2.8.1 Biosynthesis	27
2.8.2 Defence against nematodes	27
2.9 SUMMARY	30
CHAPTER 3. GENERAL TECHNIQUES	32
3.1 PLANT MATERIALS AND MAINTENANCE	32
3.1.1 Spinacia oleracea	32
3.1.2 Avena sativa	32
3.1.3 Briza spp.	33
. 3.1.4 Medicago sativa	33
3.1.5 Triticum aestivum	34
3.2 NEMATODE 3.2.1 Culture	34 34
Pratylenchus neglectus	34
Heterodera schachtii	34
Heterodera avenae	35
Meloidogyne javanica	36
Ditylenchus dipsaci	36
3.2.2 Extraction	36
Misting Electrical and circuits method	36 37
Flotation and sieving method Whitehead tray	37
Extraction of Meloidogyne egg masses	37
3.2.3 Assessment	38
Direct counting	38
Root staining	39
Vitality test	39

3.3 EXTRACTION OF METHANOL-INDUCIBLE COMPOUNDS	39
3.4 DETECTION OF COMPOUNDS	40
	41
CHAPTER 4. INDUCTION OF COMPOUNDS	41
4.1 INTRODUCTION	43
4.2 MATERIALS AND METHODS	43
4.2.1 Spinach	43
20E induction in different varieties of spinach	44
20E Induction by nematode infestation of spinach	44
Effects of nematode inoculum levels on 20E induction in spinach	45
4.2.2 Oats	45
4.2.3 <i>Briza</i> spp.	46
4.2.4 Lucerne	46
4.2.5 Experimental Design and Statistical Analysis	47
4.3 RESULTS	47
4.3.1 Spinach	47
20E induction in different varieties of spinach	47
20E Induction by nematode infestation of spinach	50
Effects of nematode inoculum levels on 20E induction in spinach	52
4.3.2 Oats	55
4.3.3 <i>Briza</i> spp.	56
4.3.4 Lucerne	59
4.4 DISCUSSION	66
4.4 DISCUSSION	66
CHAPTER & DIOLOGICAL AND CHEMICAL CHAPACTERICATION OF	
CHAPTER 5. BIOLOGICAL AND CHEMICAL CHARACTERISATION OF	66
INDUCIBLE COMPOUNDS IN CRUDE EXTRACTSCOMPOUNDS	
5.1 INTRODUCTION	66
5.2 MATERIALS AND METHODS	67
5.2.1 Preparation of Plant Extracts	67
5.2.2 Microplate-based bioassay for Ecdysteroid-like or Anti-ecdysteroid Activities	67
Culture of BII tumorous blood cell line	67
Microplate assay	67
5.2.3 Gradient Elution RP-HPLC	68
5.2.4 Mass Spectrometry	69
5.2.5 Experimental Design and Statistical Analysis	70
5.3 RESULTS	70
5.3.1 Microplate-based Bioassay	70
20E response curve	70
Plant extracts	72
5.3.2 RP-HPLC	72
5.3.3 UV-VIS spectra of compounds	76
5.3.4 Mass Spectrometry	79
5.4 DISCUSSION	79
3.4 DISCUSSION	19
CHAPTED & ECONOTEDATE IN DIAMEDADA CITICATEMA PONDO	0.2
CHAPTER 6. ECDYSTEROIDS IN PLANT PARASITIC NEMATODES 6.1 INTRODUCTION	83
	83
6.2 MATERIALS AND METHODS	84
6.2.1 Preparation of Nematodes	84
6.2.2 Extraction of Ecdysteroids	84
6.2.3 Detection and Quantification of Ecdysteroids	86
6.3 RESULTS	86
6.4 DISCUSSION	86
CHAPTER 7. PROTECTIVE ROLE OF PHYTOECDYSTEROIDS AGAINST PLANT PARASITIC NEMATODES	90
7.1 INTRODUCTION	90
7.2 MATERIALS AND METHODS	92
7.2.1 Exogenous Ecdysteroid	92
7.2.2 Endogenous Phytoecdysteroid	93
7.2.3 Phytoecdysteroid Quantification	94
7.2.4 Experimental Design and Statistical Analysis	94
7.3 RESULTS	94
THE NEW LID	74

7.3.1 Effect of Exogenous Ecdysteroid	94
7.3.2 Effect of Endogenous Ecdysteroid	97
Spinach	98
Quaking grass	102
7.4 DISCUSSION	105
CHAPTER 8. GREEN MANURE CONTAINING 20E FOR NEMATODE	110
CONTROL	
8.1 INTRODUCTION	110
8.2 MATERIALS AND METHODS	110
8.3 RESULTS	111
8.4 DISCUSSION	114
CHAPTER 9. PROTECTIVE EFFECTS OF FLAVONE-C-GLYCOSIDE IN	116
OATS	
9.1 INTRODUCTION	118
9.2 MATERIALS AND METHODS	118
9.2.1 Plant and Nematode Culture	118 118
9.2.2 Induction of Compounds in Oats9.2.3 Extraction of Inducible compounds	118
9.2.4 Chemical Analysis	118
9.2.5 Fractionation of Shoot and Root Extracts of Oats	119
9.2.6 Test for Biological Activity of Inducible Compounds	120
9.2.7 Protective Effects of Methanol-extractable Compounds	120
9.2.8 Experimental Design and Statistical Analysis	121
9.3 RESULTS	121
9.3.1 Inducibility of Compounds	122
9.3.2 Bioassay of Crude Extract	122
9.3.3 Biological Activity of the Inducible Compounds	124
9.3.4 Protective Effect of Inducible Compounds	124
9.3.5 Identification of the Inducible Compounds	126
9.4 DISCUSSION	128
	131
CHAPTER 10. GENERAL CONCLUSIONS AND DISCUSSION	137
APPENDICES	143
A.1 SPECIFICATIONS FOR U. C. MIX	143
A.2 ANALYSIS OF TAILEM BEND SAND	144
A.3 HOAGLAND SOLUTION	144
A.4 FORMULATION OF OSMOCOTE®	145
REFERENCES	146

The insect moulting hormone, 20-hydroxyecdysone (20E), is inducible in spinach and has been demonstrated to provide defence against insect herbivory. It is not known if such phytoecdysteroids are inducible by and defensive against plant parasitic nematodes. However, given that insects and nematodes belong to the same clade, the ecdysozoa, this is possible. Therefore, plants were tested for the presence of inducible phytoecdysteroids and effects on the nematodes tested.

Induction of possible defence compounds in common cultivars of spinach, two Briza spp., B. maxima and B. minor, oats and lucerne cultivars with varying degree of resistance to stem nematode, Ditylenchus dipsaci, was undertaken by treatment with methyl jasmonate and by challenging plants with nematodes. The influence of nematode inoculum density on the induction of phytoecdysteroid was also assessed in spinach. In addition, the relationship between the levels of inducible compounds and resistance response of lucerne cultivars to the stem nematode were evaluated.

Treatment with methyl jasmonate induced methanol extractable compounds in all plants tested. *Pratylenchus neglectus* induced the same compounds at levels equivalent to methyl jasmonate induction in all plants except lucerne, which was not tested. An inoculum rate of 500 to 10,000 *P. neglectus* induced similar levels of phytoecdysteroids in spinach. *Heterodera schachtii* induced phytoecdysteroids in both roots and shoots of spinach. *H. avenae* induced methanol extractable compounds in the roots of *B. minor* and shoot and roots of oats. *Meloidogyne javanica* was only found to increase levels of phytoecdysteroids in the shoots of spinach. Among the plants inoculated with the stem nematode, induced compounds were detected only in some lucerne cultivars resistant to the nematode.

The methanol extracts from the induced plants were further tested for biological activity using *Drosophila melanogaster* B₁₁ cell microplate-based bioassay to screen and

detect biologically active ecdysteroids. The extracts were subjected to mass spectrometry to confirm the presence of ecdysteroids. The biological and chemical characterisation of the inducible compounds in the plants tested provided evidence that spinach, *Briza* spp. and lucerne contained the ecdysteroids 20E and polypodine B, which were biologically active based on the B_{II} cell bioassay, except for lucerne. Lucerne shoots appeared to contain compounds or conjugate groups that inhibit ecdysteroids. In addition to the ecdysteroids above, *B. maxima* also contained ecdysone. On the other hand, inducible flavonoids were observed in the shoots and roots of oats.

Two plant parasitic nematodes, *P. neglectus* and *Anguina tritici*, were examined for the presence of similar ecdysteroids induced in the plants tested. This information will corroborate the involvement of these compounds in plant defence against nematodes. Based on HPLC and mass spectrometry data, both nematodes did not contain the ecdysteroids induced in the plants. However, compounds with masses similar to 20,26-dihydroxyecdysone, 20,26-dihydroxyecdysone 22-acetate, makisterone A, and possibly an unreported ecdysteroid were observed in *P. neglectus*. No ecdysteroid was observed in *A. tritici*, which consisted only of second stage juveniles in the anhydrobiotic survival state as opposed to the presence of all stages of an actively developing population of *P. neglectus*.

In order to establish that ecdysteroids are potential defence compounds against parasitic nematodes, the effects of direct application of 20E on nematodes was assessed by treating cereal cyst nematode, *H. avenae*, juveniles with concentrations of 20E from 8.2x10⁻⁸ to 5.2x10⁻⁵ M before applying to wheat. *H. avenae*, *H. schachtii*, *M. javanica* and *P. neglectus* were treated with 5.2x10⁻⁵ 20E and incubated in moist sand. To test the protective effects of 20E in plants, *H. schachtii* and *H. avenae* were applied to spinach and quaking grass, respectively, and the latter two nematodes in both plants, in which elevated concentrations of 20E had been induced by methyl jasmonate. Abnormal moulting, immobility, reduced invasion, impaired development and death occurred in nematodes

exposed to 20E either directly at concentration above 4.2x10⁻⁷ M or in plants. Phytoecdysteroid induction apparently protected spinach and *B. maxima* from plant parasitic nematodes and may confer a mechanism for nematode resistance.

Green manure is an alternative option to deliver the defence compound, as high constitutive production in a crop plant might impose unacceptable metabolic cost. Induced spinach when applied as green manure suppressed invasion of *H. avenae* in wheat but the direct involvement of 20E was not established because of the highly toxic effects of the treatment on the nematode.

Three inducible compounds, isolated in methanolic root and shoot extracts of oats were identified as flavone-*C*-glycosides by mass spectrometry. The effect of the flavone-*C*-glycosides on the invasion by and development of cereal cyst nematode, *H. avenae*, was assessed using methanolic extracts of shoots and roots from methyl jasmonate treated plants. Both extracts impaired nematode invasion and development. When the extracts were fractionated by high voltage paper electrophoresis, only one flavone-*C*-glycoside, *O*-methyl-apigenin-*C*-deoxyhexoside-*O*-hexoside, inhibited nematode invasion. The protective effect of the induction of flavone-*C*-glycosides in oats by methyl jasmonate was evaluated against *H. avenae* and *P. neglectus*. Treatment with methyl jasmonate reduced invasion of both nematodes and increased plant mass, compensating for damage caused by the nematodes, and is attributed to the active flavone-*C*-glycoside. The active compound, *O*-methyl-apigenin-*C*-deoxyhexoside-*O*-hexoside, has not been implicated previously in plant defense against any pest or pathogen, and appears to provide protection against the major cereal nematodes *Heterodera* and *Pratylenchus*.