

09 PH
K139



IMMUNE RESPONSE TO ENZYME REPLACEMENT THERAPY IN MPS I AND GSD II PATIENTS

By

Revecca Kakavanos

B.Sc. (Hons)

Faculty of Health Sciences
Department of Paediatrics



This thesis is submitted for the degree of Doctor of Philosophy

February, 2006

TABLE OF CONTENTS

Thesis Summary	i
Declaration	iv
Acknowledgements	v
List of Abbreviations	vi
List of Figures	viii
List of Tables	xi
Publications arising from this study	xii
Chapter 1: Introduction	1
<i>1.1 The lysosome and the endocytic network</i>	<i>2</i>
1.1.1 Synthesis of lysosomal hydrolases and delivery to the lysosome	4
<i>1.2 LSD</i>	<i>5</i>
1.2.1 Prevalence and genetics of LSD	6
1.2.2 Clinical presentation	10
1.2.3 Diagnosis of LSD.....	12
<i>1.3 GSD II</i>	<i>14</i>
1.3.1 History of GSD II	14
1.3.2 GSD II clinical phenotype	16
1.3.2.1 Infantile-onset GSD II	17
1.3.2.2 Juvenile-onset GSD II.....	19
1.3.2.3 Adult-onset GSD II.....	20
1.3.3 Diagnosis of GSD II	20
1.3.4 Genetics of GSD II	22
1.3.5 Protein synthesis and post translational processing of α-D-glucosidase....	23
<i>1.4 MPS I</i>	<i>24</i>
1.4.1 Clinical phenotype of MPS I	26
1.4.1.1 Hurler syndrome	26
1.4.1.2 Hurler-Scheie syndrome	28
1.4.1.3 Scheie syndrome.....	28
1.4.2 Genetics of MPS I.....	28
1.4.3 Diagnosis of MPS I.....	30
1.4.4 Protein synthesis and post-translational processing of α-L-iduronidase	31
<i>1.5 Therapy for LSD</i>	<i>31</i>
1.5.1 <i>In vitro</i> studies of ERT in MPS I and GSD II.....	37
1.5.2 Preclinical and clinical trials of ERT in GSD II	38
1.5.3 Preclinical and clinical trials of ERT in MPS I	41
<i>1.6 Immune response to ERT</i>	<i>42</i>
1.6.1 Measures of an immune response	42
1.6.2 Frequency of immune response in ERT-treated LSD animals and patients	43
1.6.3 Adverse effects of antibody production.....	46
1.6.4 Antibody effects on ERT efficacy	48
1.6.5 Significance of immune response to ERT	51
<i>1.7 Hypothesis and aims</i>	<i>53</i>

Chapter 2: Materials and Methods	54
2.1 <i>Materials</i>	55
2.2 <i>Tissue culture</i>	56
2.2.1 Culture of CHO cells	56
2.2.1.1 rh- α -D-Glucosidase expression with and without additional D-glucose	56
2.2.1.2 Expression of rh- α -D-glucosidase, rh- α -L-iduronidase and rh-4- sulphatase	57
2.2.2 Culture of GSD II patient skin fibroblasts in the presence of D-glucose.....	57
2.2.3 Cell harvesting	58
2.2.4 Cell lysate preparation	58
2.2.5 Culture of hybridoma cell lines	59
2.2.6 Cryopreservation of hybridoma, fibroblast and CHO-K1 cells.....	59
2.2.7 Thawing of cryopreserved hybridoma, skin fibroblast or CHO-K1 cells....	59
2.3 <i>Immune assays</i>	60
2.3.1 Evaluation of sera samples for antibody titre	60
2.3.2 Epitope mapping	61
2.3.3 Temperature denaturation of α -D-glucosidase	62
2.3.4 Immune quantification assay for lysosomal proteins	64
2.3.5 Multiplex analysis of lysosomal proteins	65
2.4 <i>Infusion of rh-α-D-glucosidase into animal models</i>	66
2.4.1 rh- α -D-Glucosidase immunised rats	66
2.4.2 Subcutaneous infusion of rh- α -D-glucosidase in mice	67
2.5 <i>Blood collection and sera sample preparation</i>	67
2.5.1 Heart puncture.....	67
2.5.2 Tail vein blood sampling from rats and mice	68
2.5.3 Preparation of ovalbumin/BSA-bound Affi-Gel 10.....	68
2.5.4 Albumin absorption of sera samples.....	69
2.6 <i>rh-α-D-Glucosidase purification from CHO-K1 expression cells</i>	69
2.6.1 rh- α -D-Glucosidase purification	69
2.6.2 Modified rh- α -D-glucosidase purification	70
2.7 <i>Enzyme activity assays</i>	71
2.7.1 α -D-Glucosidase activity	71
2.7.2 α -L-Iduronidase activity	71
2.7.3 4-Sulphatase activity.....	72
2.7.4 Enzyme inhibition studies.....	72
2.8 <i>Protein estimation</i>	73
2.8.1 BCA method for protein determination	73
2.8.2 Lowry method for protein determination.....	73
2.9 <i>Western blot analysis</i>	74
Chapter 3: Immune reactivity in MPS I.....	76
3 <i>Introduction</i>	77
3.1 <i>Results</i>	77
3.1.1 Antibody response in rh- α -L-iduronidase-treated mice.....	77
3.1.2 Antibody titres to rh- α -L-iduronidase in ERT treated MPS I patients	79
3.1.3 Epitope reactivity of ERT treated MPS I patient sera antibodies	81

3.2 Discussion.....	89
Chapter 4: Glycosidase cross reactivity.....	94
4 Introduction	95
4.1 Results.....	97
4.1.1 Epitope reactivity of α-D-glucosidase and α-L-iduronidase polyclonal antibodies.....	97
4.1.1.1 Antibody reactivity to native and denatured forms of rh-α-D-glucosidase	99
4.1.1.2 Epitope reactivity of monoclonal antibodies to α-D-glucosidase.....	99
4.1.1.3 Effect of temperature on monoclonal antibody reactivity to α-D-glucosidase	102
4.1.1.4 Antibody titres to rh-α-D-glucosidase in four immunised rats	105
4.1.1.4.1 Epitope reactivity of immunised rat sera antibodies.....	105
4.1.1.5 Epitope cross-reactivity of α-D-glucosidase and α-L-iduronidase antibodies.....	112
4.1.1.6 Analysis of antigenic motifs that had sequence identity.....	114
4.2 Discussion.....	117
Chapter 5: Effect of D-glucose on α-D-glucosidase expression, purification and antibody reactivity	123
5 Introduction	124
5.1 Results.....	125
5.1.1 Differential rh-α-D-glucosidase expression in media.....	125
5.1.1.1 The affect of increasing D-glucose concentration on rh-α-D-glucosidase expression	128
5.1.1.2 Effect of sugar and butyric acid on rh-α-D-glucosidase, rh-α-L-iduronidase and rh-4-sulphatase expression	128
5.1.2 Inhibition studies for two glycosidase enzymes	133
5.1.2.1 Effect of monosaccharide on purified rh-α-D-glucosidase.....	133
5.1.2.2 Comparison of rh-α-D-glucosidase inhibition by D-glucose	135
5.1.2.3 Inhibition of rh-α-L-iduronidase by D-glucose	135
5.1.3 Improved yield of rh-α-D-glucosidase from CHO-K1 cell cultures treated with D-glucose	139
5.1.4 Effect of D-glucose on α-D-glucosidase activity in GSD II patient skin fibroblasts.....	142
5.1.5 Differential antigenicity of rh-α-D-glucosidase from transgenic rabbit milk and a CHO-K1 cell line	142
5.1.5.1 Antigenicity of rh-α-D-glucosidase in enzyme-treated animals	142
5.1.5.2 Epitope reactivity of serum antibodies from mice treated with rh-α-D-glucosidase.....	145
5.1.6 Reduction of antibody reactivity to rh-α-D-glucosidase in the presence of D-glucose	148
5.2 Discussion.....	152
Chapter 6: Concluding discussion.....	158
References	172

THESIS SUMMARY

Enzyme replacement therapy (ERT) has been shown to be an effective treatment strategy for a number of lysosomal storage diseases (LSD's) in both animal models and in human clinical trials. Immune response to replacement proteins however has been reported for a number of LSD and can be a potential complication for therapy. Immune responses to ERT are variable and dependent on both the inherent antigenicity of the protein being infused and the individual patient. I have investigated the immune reactivity of two lysosomal glycosidases, α -D-glucosidase and α -L-iduronidase. Functional deficiencies of these lysosomal enzymes are the cause of the LSD Glycogen storage disease type II (GSD II) and mucopolysaccharidosis type I (MPS I) respectively.

In a phase I/II clinical trial of ERT in MPS I patients, an immune response to α -L-iduronidase was observed in 50% of patients. The MPS I patients that initially had an immune response were shown in this thesis, to develop natural immune-tolerance to α -L-iduronidase after a year of ERT. This finding has positive implications for MPS I patients who are undergoing long term ERT. A specific peptide that mapped to the catalytic element of α -L-iduronidase also appeared to be important in the development and maintenance of the immune response to α -L-iduronidase. In defining sites on the replacement protein, to which antibodies react, ways to improve ERT by minimising or eliminating antibody reactivity may be engineered.

It was postulated that the conservation of structural features in the active sites of glycosidases using a retaining catalytic mechanism, might result in common antigenicity. This study demonstrated that despite limited sequence identity between α -L-iduronidase and α -D-glucosidase, conserved micro-structural features and regions of short sequence

identity could make a major contribution to common glycosidase antigenicity. This common antigenicity in related glycosidases may contribute some background immune reactivity, representative of the major antigenic sites on the missing glycosidase and may account for why immune response to ERT has proven to be less of an issue than initially postulated.

A high incidence of antibody production to the rh- α -D-glucosidase replacement protein has been reported in the literature for GSD II patients treated by ERT. Furthermore, relative to other LSD types, large doses of rh- α -D-glucosidase are required for effective ERT in GSD II patients. In this thesis, the α -D-glucosidase catalytic site was identified as one of the main antigenic regions of this lysosomal protein. It was postulated that stabilising the catalytic site of α -D-glucosidase could increase the amount of protein recovered during purification and partially mask antibody reactivity against this region. This study demonstrated that D-glucose stabilised α -D-glucosidase allowing increased expression and improved purification of the recombinant protein from an over-expressing CHO-K1 cell line. This simple strategy could accommodate the high demand of enzyme required for ERT studies. D-Glucose also enhanced the residual α -D-glucosidase protein/activity in adult-onset GSD II patient cells suggesting the development of D-glucose analogues may be beneficial as an enzyme enhancement therapy for adult-onset GSD II patients. Furthermore, D-glucose-treated rh- α -D-glucosidase had reduced antibody reactivity *in vitro* suggesting that it may also be useful as a modifier of the immune response to ERT in GSD II patients. These findings have positive implications for ERT in GSD II patients.

When considering the safety and efficacy of ERT in LSD patients, it is important to know when an immune response develops, and whether it is maintained over the course of

treatment. It is clear that some LSD patients will develop antibodies against ERT and now there is also some evidence to suggest that these could compromise the health of the patient and therapy efficacy. Therefore, characterising these immune responses becomes crucial for the management of patients receiving long term ERT. An increased understanding of antibody development in enzyme treated LSD patients will aid in engineering ways to avert the potential affects of antibody production and will assist in the delivery of a more effective and safe therapy.