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An unusual cause of severe metabolic acidosis

John V Peter, Natasha Rogers, Shailesh Murty, Rosemarie Gerace, Richard Mackay and Sandra L Peake

A 50-year-old man was transferred to the intensive care unit with high anion gap metabolic acidosis. Investigations suggested a diagnosis of pyroglutamic acidemia. Factors contributing to the acidosis were medications (paracetamol and flucloxacillin), sepsis and renal failure. The acidosis resolved with supportive therapy and withdrawal of the drugs. It is important to recognise this treatable aetiology of metabolic acidosis. (MJA 2006; 185: 223-225)

Clinical record

A 50-year-old man with cerebral palsy, intellectual impairment and epilepsy was referred to hospital with a 1-week history of fever, chills, rigors, haematuria and loin pain. His usual medications included phenytoin 300 mg/day, phenobarbitone 30 mg/day and carbamazepine 1200 mg/day. There was no history of prior renal disease.

At hospital admission (Day 1), he was conscious, in no obvious distress, and afebrile. He was dehydrated and tachypnoeic (respiratory rate, 22 breaths/minute), but haemodynamically stable. There was no pallor, jaundice or cyanosis. Cardiorespiratory examination was unremarkable. Abdominal examination revealed tenderness in the left renal angle, left loin and right upper quadrant with no features of peritonism.

Key initial (and subsequent) screening investigations are summarised in Box 1. Of note was neutrophilia and significant renal impairment (glomerular filtration rate, 28 mL/min by MDRD 4-variable equation). Urinalysis showed sterile pyuria, haematuria (dysmorphic red blood cells on microscopy) and proteinuria (3.46 g following 24-hour collection). A chest x-ray (CXR) showed mild generalised bronchial wall thickening, with no focal parenchymal opacity. Treatment with empirical broad-spectrum intravenous antibiotics (ceftriaxone and gentamicin) was initiated for suspected urinary tract infection following blood and urine cultures.

A computed tomography (CT) scan on Day 1 revealed a 2.3 cm simple cyst in the upper pole of the right kidney, multiple nodules in the lung bases and a small left pleural effusion. A vasculitic work-up, including assessment of antinuclear antibody, extractable nuclear antigen, antibodies to double-stranded DNA, antineutrophil cytoplasmic antibodies and complement levels, did not assist in diagnosis. Urine culture was negative, so a renal biopsy was performed to establish the aetiology of the acute nephritic syndrome, consistent with IgA nephropathy with mild activity (segmental crescents/necrotising lesions in two of 14 glomeruli), but without scarring. No treatment was indicated.

In view of the persistent fever on Day 3, a repeat CXR was performed. A small left-sided effusion was noted and pleural fluid aspirate was consistent with an exudate (pleural fluid white cell count [WCC], $16.6 \times 10^9/L$; 85% neutrophils; total protein, 37 g/L; lactate dehydrogenase [LDH], 420 U/L), which was presumed secondary to an underlying pneumonic process. Intravenous flucloxacillin was commenced at 2 g/day to broaden the gram-positive antibiotic cover and continued for 11 days. Blood, urine and pleural fluid cultures were negative.

Despite initial clinical and laboratory improvement, the fever recurred on Day 7 with a radiographically visible increase in the

1 Investigations*

	Day 1 Hospital admission	Day 18 ICU admission	Day 63 On recovery	Reference range
Haematology				
Haemoglobin (g/L)	125	115	120	135–175
White cell count ($\times 10^9/L$)	13.6	24.7	7.82	4–11
Neutrophil count ($\times 10^9/L$)	11.1	22.75	6.09	1.8–7.5
International normalised ratio	1.2	7.6	1.0	0.8–1.2
APTT (s)	27	65	26	24–37
Fibrinogen level	—	5.7	—	1.5–4.0
D-dimer FDP	—	2.15	—	< 2.0
Biochemistry				
Sodium (mmol/L)	134	138	137	137–145
Potassium (mmol/L)	3.9	2.8	4.1	3.5–4.9
Chloride (mmol/L)	97	93	99	100–109
Bicarbonate (mmol/L)	23	6	29	22–32
Urea (mmol/L)	9.1	18.6	2.7	2.7–8.0
Creatinine (mmol/L)	0.210	0.541	0.100	0.05–0.12
Gamma GT (U/L)	152	92	116	0–60
Albumin (g/L)	25	17	23	34–48
Lactate dehydrogenase (U/L)	252	293	210	110–230
Serum amylase (U/L)	—	119	—	20–100
C-reactive protein (mg/L)	290	160	47	< 10
Anion gap (mmol/L)	18	42	13	7–17
Lactate level (mmol/L)	—	0.7	—	0.2–2.0
Serum pyroglutamic acid level† ($\mu\text{mol/L}$)	—	11 010	—	15–215
Urine pyroglutamic acid level† ($\mu\text{mol/mmol creatinine}$)	—	20 495	13 103	< 100

* Platelet count, bilirubin, alanine aminotransferase, aspartate aminotransferase, alkaline phosphatase and creatinine kinase were normal when measured.

† Initial serum and urine pyroglutamic acid levels were measured 36 hours after admission to the intensive care unit, and repeat urine test was performed on Day 10. Serum and urine pyroglutamic acid levels not measured at discharge.

— = Not measured. Bold indicates highly abnormal results.

APTT = activated partial thromboplastin time.

FDP = fibrinogen degradation products.

GT = glutamyl transpeptidase. ICU = intensive care unit.

received a cumulative dose of 8 g of paracetamol over 4 days, and it is likely that PGA was precipitated by a combination of factors, including sepsis, renal dysfunction, and co-administration of flucloxacillin.

PGA also occurs with genetic deficiency of either glutathione synthetase or 5-oxoprolinase.⁵ However, not all causes of 5-oxoprolinuria are necessarily associated with acidaemia.

The mechanism of non-hereditary PGA is probably multifactorial. Suppression of glutathione levels because of sepsis, as seen in animal models of polymicrobial sepsis,⁹ may have contributed to the development of PGA in our patient. Flucloxacillin could have further compounded this acidosis by inhibiting the breakdown of pyroglutamic acid by 5-oxoprolinase.² The role of paracetamol in PGA is more complex.

The metabolite of paracetamol, *N*-acetyl benzoquinoneimine, reacts irreversibly with glutathione. Under normal circumstances, glutathione depletion leads to increased γ -glutamyl cysteine synthetase activity and excessive production of γ -glutamyl cysteine (Box 2). However, under altered conditions, glutathione synthetase activity becomes rate-limiting, leading to the conversion of γ -glutamyl cysteine to 5-oxoprolinone by γ -glutamyl cyclotransferase.⁵ Our patient's antiepileptic medications, which are known hepatic enzyme inducers and metabolised via CYP2E1, may have further compromised glutathione availability by decreasing glutathione stores and competing with paracetamol for metabolism.

Renal impairment may also be important. Renal tubular epithelial dysfunction may impair intracellular glutathione re-formation (a high ATP-requiring state), leading to accumulation of 5-oxoprolinone and prompt excretion (because of its small molecular size) into the urine, peritubular capillaries and systemic circulation.

The use of *N*-acetyl cysteine to treat PGA has been advocated to replenish glutathione stores by supplying cysteine for glutathione synthesis. Another theoretical treatment option is the use of cysteamine, which increases cytosolic cysteine and restores substrate availability for the glutamate pathway, normalising pyroglutamic acid levels.

As PGA can be easily missed in a critically ill patient, where several factors may contribute to a metabolic acidosis, a high index of suspicion is required to diagnose this condition. PGA should be considered in the differential diagnosis of high anion gap acidosis, especially when the levels of organic acids do not sufficiently account for the degree of anion gap and when there is co-

administration of drugs such as paracetamol and flucloxacillin. Where PGA is suspected, the offending drugs should be withdrawn and treatment with *N*-acetyl cysteine considered.

Competing interests

None identified.

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