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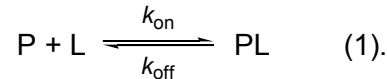
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# Appendix 1

## Derivation of the Equilibrium Binding Dissociation constant, $K_d$

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In order to define  $K_d$ , it is necessary to consider the equations below. Consider a ligand binding event between a protein P, and a ligand L, to form the 1:1 complex PL as described by the following equilibrium:



Thus, the equilibrium binding association constant  $K_a$  is then defined as:

$$K_a = [PL]/[P].[L] \approx k_{on}/k_{off} \quad (2).$$

Then, the equilibrium binding dissociation constant,  $K_d$  is defined as:

$$K_d = [P].[L]/[PL] \approx k_{off}/k_{on} = 1/K_a \quad (3).$$

Now let

$$P_0 = \text{the total concentration of protein} = [P] + [PL] \quad (4),$$

$$\text{and } L_0 = \text{the total concentration of ligand} = [L] + [PL] \quad (5),$$

then  $K_d$  may also be expressed as:

$$K_d = (P_0 - [PL]).(L_0 - [PL])/[PL] \quad (6),$$

and [PL] is given by,

$$[PL] = \frac{1}{2} \cdot \{L_0 + P_0 + K_d - [(L_0 + P_0 + K_d)^2 - 4L_0P_0]^{1/2}\} \quad (7).$$

In practice, the  $K_d$  is often referred to as [L] corresponding to 50% occupancy of the protein P with the ligand L. Thus, according to equation (3), if there is 50% occupancy of P, then  $[PL] = [P]$ , and therefore:

$$K_d = [L] \quad (8).$$

However, it is important to note that this [L] here refers to the 'residual', unbound ligand L, according to the equation

$$[L] = L_0 - [PL] \quad (9).$$

## Appendix 2

### Data analysis process for NMR chemical shift perturbation assay

**Table A-1:** NMR [<sup>1</sup>H, <sup>15</sup>N] HSQC chemical shift data for binding of **2** to Tec SH3 domain.

d <sup>1</sup> H (H-N) (ppm)											
[2] (mM)	D196	E193	G226	L197	Q190	S230	T192	W215	W215e1	W216	Y227
0	7.61	8.18	8.55	8.73	8.22	7.96	8.88	8.48	9.99	8.98	9.23
62.5	7.64	8.22	8.59	8.90	8.18	7.86	8.81	8.35	9.79	8.95	9.31
125	7.67	8.24	8.61	8.99	8.16	7.81	8.76	8.29	9.69	8.91	9.35
187.5	7.68	8.26	8.62	9.04	8.15	7.78	8.73	8.26	9.63	8.89	9.38
250	7.68	8.26	8.62	9.06	8.14	7.76	8.71	8.23	9.60	8.88	9.39
625	7.70	8.28	8.65	9.14	8.12	7.72	8.67	8.18	9.51	8.87	9.43
1250	7.71	8.28	8.65	9.17	8.11	7.71	8.65	8.15	9.47	8.85	9.44

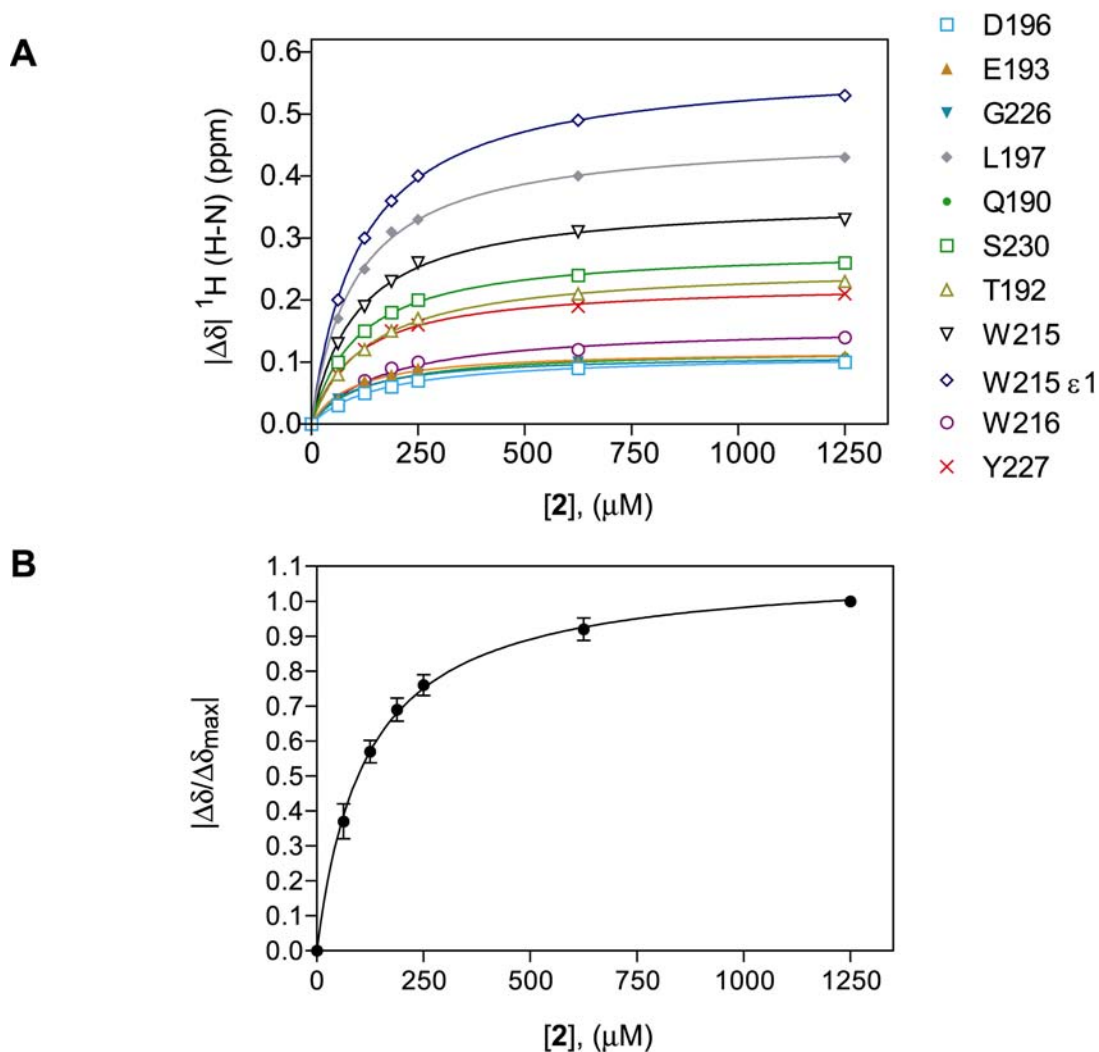
  

Dd <sup>1</sup> H (H-N) (ppm)													
[2] (mM)	D196	E193	G226	L197	Q190	S230	T192	W215	W215e1	W216	Y227		
0	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00		
62.5	0.03	0.04	0.04	0.17	-0.04	-0.10	-0.08	-0.13	-0.20	-0.03	0.08		
125	0.05	0.07	0.06	0.25	-0.06	-0.15	-0.12	-0.19	-0.30	-0.07	0.12		
187.5	0.06	0.08	0.07	0.31	-0.07	-0.18	-0.15	-0.23	-0.36	-0.09	0.15		
250	0.07	0.09	0.08	0.33	-0.08	-0.20	-0.17	-0.26	-0.40	-0.10	0.16		
625	0.09	0.10	0.10	0.40	-0.10	-0.24	-0.21	-0.31	-0.49	-0.12	0.19		
1250	0.10	0.11	0.10	0.43	-0.11	-0.26	-0.23	-0.33	-0.53	-0.14	0.21		
<b>Calculated K<sub>d</sub> (mM)</b>	163.2	97.57	108.9	105.3	129.6	110.4	135.0	108.8	116.4	168.4	108.1	<b>Mean</b>	<b>Std.Dev</b>
												122.88	23.77

Dd/Dd <sub>max</sub>													
[2] (mM)	D196	E193	G226	L197	Q190	S230	T192	W215	W215e1	W216	Y227	Mean	Std.Dev
0	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00
62.5	0.31	0.42	0.39	0.39	0.38	0.40	0.33	0.40	0.38	0.25	0.38	0.37	0.05
125	0.54	0.62	0.61	0.58	0.58	0.59	0.53	0.58	0.58	0.51	0.57	0.57	0.03
187.5	0.66	0.76	0.71	0.70	0.70	0.70	0.63	0.68	0.68	0.66	0.70	0.69	0.03
250	0.70	0.81	0.76	0.76	0.76	0.79	0.72	0.77	0.75	0.74	0.75	0.76	0.03
625	0.90	0.93	0.98	0.93	0.90	0.94	0.90	0.92	0.93	0.85	0.92	0.92	0.03
1250	1.00	1.00	1.00	1.00	1.00	1.00	1.00	1.00	1.00	1.00	1.00	1.00	0.00

Table A-1 above contains a list of all amino acid residues whose  $^1\text{H}$  (H-N) chemical shifts were altered by at least 0.1 ppm at or near saturation binding of 2-aminoquinoline **2**. The absolute changes in  $^1\text{H}$  (H-N) chemical shift  $|\Delta\delta|$  were then plotted against **[2]** (as illustrated in Figure A-1A below), and non-linear regression analysis performed using GraphPad Prism.<sup>55</sup> The calculated  $K_d$  for each residue was then obtained from the Prism results table (Table A-2). The mean  $\pm$  standard deviation of the obtained  $K_d$  values was then calculated (Table A-1), to provide the  $K_d$  value that has been quoted in the main text of this thesis (given in  $\mu\text{M}$ , following rounding to the nearest whole number). For ease of comparison between different ligands,  $|\Delta\delta/\Delta\delta_{\text{max}}|$  terms were calculated for each residue (Table A-1). The mean  $\pm$  standard deviation of these  $|\Delta\delta/\Delta\delta_{\text{max}}|$  terms was then calculated (Table A-1) and this standardised data is plotted as in Figure A-1B below. Unless otherwise specified, this is the method by which equilibrium binding isotherms have been illustrated in the main text of this thesis.



**Figure A-1:** Equilibrium binding of **2** to Tec SH3 domain as represented by change in  $^1\text{H}$  (H-N) chemical shift from  $[^1\text{H}, ^{15}\text{N}]$ -HSQC NMR experiments. (A) Binding isotherms for amino-acid residues where  $\Delta\delta = 0.1$  ppm at close to saturation binding of **2**. (B) Binding isotherm represented by averaging the normalized chemical shift changes  $|\Delta\delta/\Delta\delta_{\text{max}}|$  calculated for all residues where  $|\Delta\delta_{\text{max}}| = 0.1$  ppm. (See Table A-1).

**Table A-2:** Equilibrium binding of **2** to Tec SH3 domain using NMR. Summary of GraphPad Prism output following non-linear regression analysis of chemical shift changes for amino-acids residues involved in binding of **2** (Obtained from  $\Delta\delta$  values presented in Table A-1 above.)

	<b>Residue</b>	<b>D196</b>	<b>E193</b>	<b>G226</b>	<b>L197</b>	<b>Q190</b>	<b>S230</b>
Equation 1							
Best-fit values							
	BMAX	0.1135	0.1192	0.1126	0.4684	0.121	0.2838
	KD	163.2	97.57	108.9	105.3	129.6	110.4
Std. Error							
	BMAX	0.001262	0.004195	0.002824	0.005674	0.0009455	0.002202
	KD	5.461	12.54	9.539	4.515	3.309	2.975
95% Confidence Intervals							
	BMAX	0.1103 to 0.1167	0.1084 to 0.1300	0.1053 to 0.1199	0.4538 to 0.4830	0.1185 to 0.1234	0.2781 to 0.2895
	KD	149.1 to 177.2	65.33 to 129.8	84.35 to 133.4	93.65 to 116.9	121.1 to 138.1	102.7 to 118.0
Goodness of Fit							
	Degrees of Freedom	5	5	5	5	5	5
	R <sup>2</sup>	0.9993	0.9906	0.9954	0.9989	0.9996	0.9996
	Absolute Sum of Squares	5.03E-06	8.28E-05	3.48E-05	0.0001438	3.42E-06	2.09E-05
	Sy.x	0.001003	0.00407	0.002638	0.005363	0.0008273	0.002046
Data							
	Number of X values	7	7	7	7	7	7
	Number of Y replicates	1	1	1	1	1	1
	Total number of values	7	7	7	7	7	7
	Number of missing values	0	0	0	0	0	0
	<b>Residue</b>	<b>T192</b>	<b>W215</b>	<b>W215e1</b>	<b>W216</b>	<b>Y227</b>	
Equation 1							
Best-fit values							
	BMAX	0.256	0.3627	0.5813	0.1588	0.2273	
	KD	135	108.8	116.4	168.4	108.1	
Std. Error							
	BMAX	0.002719	0.004506	0.002543	0.00932	0.003846	
	KD	4.616	4.723	1.733	29.45	6.409	
95% Confidence Intervals							
	BMAX	0.2490 to 0.2630	0.3511 to 0.3743	0.5748 to 0.5878	0.1348 to 0.1827	0.2174 to 0.2371	
	KD	123.1 to 146.9	96.64 to 120.9	112.0 to 120.9	92.70 to 244.1	91.60 to 124.6	
Goodness of Fit							
	Degrees of Freedom	5	5	5	5	5	
	R <sup>2</sup>	0.9993	0.9989	0.9999	0.9819	0.9979	
	Absolute Sum of Squares	2.74E-05	8.86E-05	2.69E-05	0.0002664	6.49E-05	
	Sy.x	0.002341	0.004209	0.002317	0.007299	0.003601	
Data							
	Number of X values	7	7	7	7	7	
	Number of Y replicates	1	1	1	1	1	
	Total number of values	7	7	7	7	7	
	Number of missing values	0	0	0	0	0	

# Appendix 3

## Data analysis process for Fluorescence Polarisation peptide displacement assay

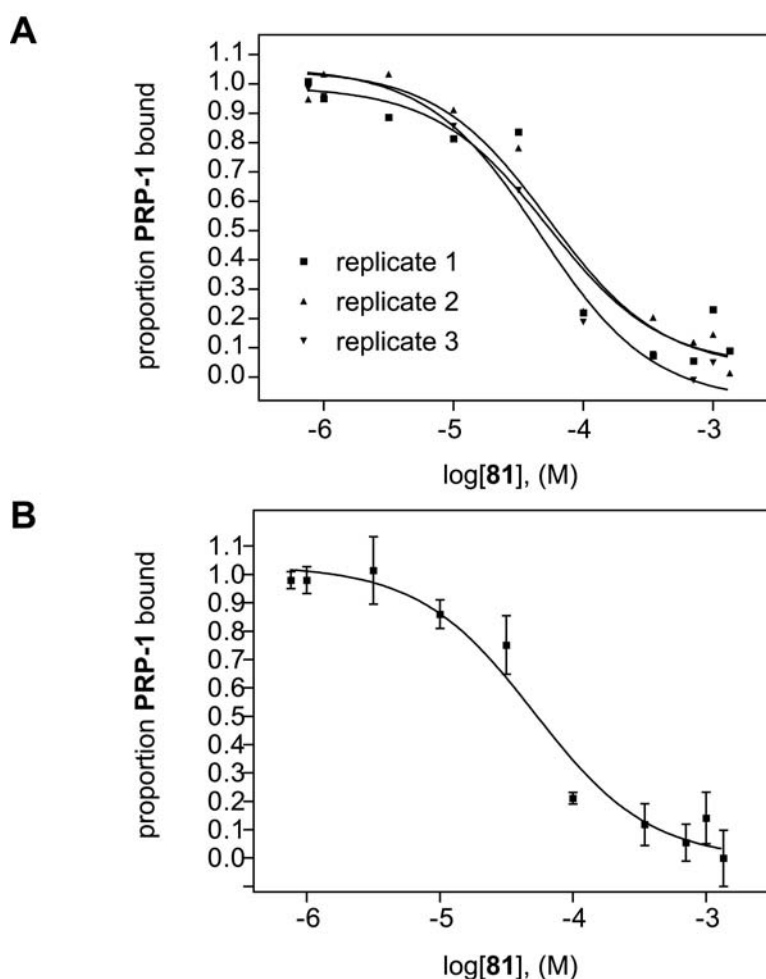
**Table A-3:** FP peptide competition assay with **PRP-1** and **81** for Tec GST-SH3 protein.

PRP-1 alone mP		replicate 1			replicate 2			replicate 3		
[L] M	log[L]	mP	DmP	Proportion bound	mP	DmP	Proportion bound	mP	DmP	Proportion bound
0.00E+00		132.75	58.19	1.00	130.87	59.49	1.00	129.00	51.26	1.00
7.50E-07	-6.12	133.22	58.66	1.01	127.78	56.40	0.95	128.11	50.37	0.98
1.00E-06	-6.00	129.83	55.27	0.95	132.88	61.50	1.03	126.78	49.04	0.96
3.16E-06	-5.50	126.11	51.56	0.89	132.92	61.53	1.03	135.18	57.44	1.12
1.00E-05	-5.00	121.86	47.30	0.81	125.67	54.28	0.91	121.61	43.88	0.86
3.16E-05	-4.50	123.19	48.63	0.84	117.92	46.54	0.78	110.36	32.63	0.64
1.00E-04	-4.00	87.33	12.77	0.22	84.76	13.38	0.22	87.36	9.63	0.19
3.50E-04	-3.46	78.81	4.25	0.07	83.53	12.14	0.20	81.72	3.99	0.08
7.00E-04	-3.15	77.74	3.18	0.05	78.47	7.09	0.12	77.17	-0.57	-0.01
1.00E-03	-3.00	87.92	13.36	0.23	80.05	8.67	0.15	80.23	2.49	0.05
1.35E-03	-2.87	79.77	5.21	0.09	72.19	0.80	0.01	72.22	-5.51	-0.11

Summary	log[L]	Proportion bound			Average of	St Dev of		
		replicate 1	replicate 2	replicate 3	Replicates	Replicates		
	-6.12	1.01	0.95	0.98	0.98	0.03		
	-6.00	0.95	1.03	0.96	0.98	0.05		
	-5.50	0.89	1.03	1.12	1.01	0.12		
	-5.00	0.81	0.91	0.86	0.86	0.05		
	-4.50	0.84	0.78	0.64	0.75	0.10		
	-4.00	0.22	0.23	0.19	0.21	0.02		
	-3.46	0.07	0.20	0.08	0.12	0.07		
	-3.15	0.06	0.12	-0.01	0.05	0.07		
	-3.00	0.23	0.15	0.05	0.14	0.09		
	-2.87	0.09	0.01	-0.11	0.00	0.10		
<b>CALCULATED EC<sub>50</sub> (M)</b>		5.35E-05	5.40E-05	4.65E-05				

EC <sub>50</sub>	
Average (mM)	51.34
StDev (mM)	4.17

Table A-3 above contains a list of the millipolarisation (mP) values for three replicate experiments involving the competition between the binding of **PRP-1** and ligand **81** for the GST-SH3 protein, at different concentrations of **81** (refer to Section 7.7.2 for definition of mP, and other equations relevant to this discussion). For each replicate,  $\Delta$ mP values were calculated, followed by proportion bound terms (Table A-3). The proportion bound terms were then plotted against  $\log[\mathbf{81}]$  for each replicate (Figure A-2A), and non-linear regression analysis using GraphPad Prism used to determine  $EC_{50}$  values (Table A-4). The mean  $\pm$  standard deviation of each  $EC_{50}$  value provided the  $EC_{50}$  value as quoted in the main text of this thesis (given in  $\mu$ M, following rounding to the nearest whole number). For ease of comparison between different ligands, the mean  $\pm$  standard deviation of the proportion bound terms (produced for the individual replicates) was calculated (Table A-3), and this data plotted against  $\log[\mathbf{81}]$  as illustrated in Figure A-2B. This is the method by which the competition binding isotherms have been illustrated in the main text of this thesis.



**Figure A-2:** FP peptide competition assay for **PRP-1** and **81** for binding to Tec GST-SH3 protein. (A) Competition isotherm for individual replicate data sets produced from data presented in Table A-3 above. (B) Competition isotherm for average of replicates, produced from data presented in Table A-3 above.



**Table A-4:** Summary of FP peptide competition assay with **PRP-1** and **81** for binding to Tec SH3 domain. [Output obtained from non-linear regression analysis of data in Table A-3 and curves in Figure A-2 (A) above.]

	replicate 1	replicate 2	replicate 3
Equation 1			
Best-fit values			
BOTTOM	0.03615	0.02892	-0.08215
TOP	0.991	1.048	1.057
LOGEC50	-4.271	-4.268	-4.332
EC50	5.36E-05	5.40E-05	4.65E-05
Std. Error			
BOTTOM	0.0798	0.06049	0.05375
TOP	0.07344	0.05548	0.0526
LOGEC50	0.2238	0.1587	0.1301
95% Confidence Intervals			
BOTTOM	-0.1526 to 0.2249	-0.1141 to 0.1720	-0.2093 to 0.04497
TOP	0.8174 to 1.165	0.9169 to 1.179	0.9323 to 1.181
LOGEC50	-4.800 to -3.742	-4.643 to -3.893	-4.640 to -4.025
EC50	1.5830e-005 to 0.0001811	2.2740e-005 to 0.0001280	2.2910e-005 to 9.4500e-005
Goodness of Fit			
Degrees of Freedom	7	7	7
R <sup>2</sup>	0.9346	0.966	0.9766
Absolute Sum of Squares	0.09929	0.0568	0.04865
Sy.x	0.1191	0.09008	0.08337
Data			
Number of X values	10	10	10
Number of Y replicates	1	1	1
Total number of values	10	10	10
Number of missing values	0	0	0

## Appendix 4

Published article:

Inglis et al., (2004) 'Identification and specificity studies of small-molecule ligands for SH3 protein domains'. *J. Med. Chem.*, 47 (22), pp. 5405-5417

This publication is included in the print copy of the thesis in the Barr Smith Library.

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