# A Genomics Approach to Investigate the Molecular Control of Meiosis in *Triticum aestivum*

by

## Timothy J. Sutton B. Ag. Sc. (Hons.)

*This thesis is submitted in partial fulfilment of the requirements for the degree of* 

Doctor of Philosophy

In

The School of Agriculture and Wine Waite Agricultural Research Institute The University of Adelaide Australia

May 2003

## **TABLE OF CONTENTS**

Abstract	i
Declaration	iii
Acknowledgements	iv
Abbreviations	v
Chapter 1 Literature Review	1
1.1 Introduction	1
1.2 The anther and the stages of meiosis	
1.3 The synaptonemal complex	5
1.3.1 Molecular composition of the synaptonemal complex	9
1.4 Genetic recombination at meiosis	
1.4.1 The Holliday and Meselson-Radding models of recombination	
1.4.2 The double strand break repair model of recombination	
1.4.3 Genes involved in recombination	
1.5 Chromosome pairing.	
1.5.1 Molecular mechanisms for homology recognition	
1.6 Chromosome pairing in <i>Triticum aestivum</i>	
1.6.1 Genetic control of chromosome pairing	
1.6.2 Models that account for the action of <i>Ph</i>	24 20
1.8 Research aims	20 20
Chapter 2 General Materials and Methods	
2.1 Plant genetic stocks	30
2.2 Collection and meiotic staging of wheat anthers	
2.3 Bacterial strains, cloning vectors and electrophoretic size markers	
2.4 Bacterial preparations and plasmid transformation	
2.4.1 <i>E. coli</i> transformation via heat shock	
2.4.2 <i>E. coli</i> transformation via electroporation	
2.5 Electrophoretic separation of DNA samples	
2.5.1 Isolation of fractionated DNA fragments from agarose gel	
2.6 Phenol:chloroform extraction and ethanol precipitation of DNA	
2.7 Indicated preparations	
2.7.1 Plasmid DNA extraction	
2.7.2 Cereal genomic DNA extraction	
2.7.2.1 Sman scale genomic DNA extraction	
2.7.2.2 Internalistate genomic DIVA extraction	
2.7.3 1 Total RNA extraction	
2.7.3.2 Poly(A) RNA purification	

2.8 Subcloning of DNA sequences	37
2.8.1 Dephosphorylation of vectors	37
2.8.2 Ligation of DNA sequences into plasmid vectors	37
2.9 Spectrophotometric quantification of DNA and RNA	38
2.10 PCR conditions and amplification of cloned inserts	38
2.11 Preparation and <sup>32</sup> P labelling of DNA probes	39
2.12 Southern blot analysis	39
2.12.1 DNA digestion and fractionation	39
2.12.2 Transfer of DNA to nylon membranes	40
2.12.3 Hybridisation and autoradiography	40
2.13 Northern blot analysis	41
2.13.1 Formaldehyde gel electrophoresis	41
2.13.2 Transfer of RNA to nylon membranes	42
2.13.3 Hybridisation and autoradiography	42
2.14 Removal of radioactive probe from membranes	42
2.15 cDNA library construction	43
2.16 Sequencing	43
Chapter 3 A Comparative Genetic Study of the Wheat <i>Ph2</i> Region	44
Abstract	44
3.1 Introduction	45
3.2 Materials and Methods	47
3.2.1 Plant Materials	47
3.2.2 DNA isolation and Southern analysis	48
3.2.3 Comparative mapping between wheat, barley and rice	48
3.2.4 Rice PAC contig assembly and consensus sequence generation	50
3.2.5 Identification of wheat ESTs with similarity to rice contig	50
3.2.6 Electronic expression analysis	50
3.3 Results	51
3.3.1 Identification of the rice region syntenous to the $ph2a$ deletion in wheat	51
3.3.2 Rice PAC physical map and consensus sequence construction	51
3.3.3 Identification of wheat ESTs from rice consensus	52
3.3.4 Synteny between the wheat 3DS and rice 1S genomic regions	55
3.3.5 Estimating the size of the $ph2a$ deletion	55
3.3.6 Analysis of rice and barley meiosis related phenotypic traits	57
3.3.7 Genic content of the $Ph2$ region and electronic expression analysis	57
3.4 Discussion	58
Chapter 4 Is <i>Ph2</i> a Meiotic Gene Cluster?	68
4.1 Introduction	68
4.2 Materials and methods	71
4.2.1 Contig assembly and display in CCV	71
4.2.2 Analysis of the meiotic anther cDNA library and <i>Ph2</i> -region ESTs	71
4.2.2.1 Selection of libraries for display	71
4.2.2.2 Contig contributions from ESTs of the meiotic anther cDNA library	73
4.2.2.3 Contigs with similarity to ESTs derived from genes of the Ph2 region	73
4.3 Results and discussion	73

4.3.1.1 The distribution of contigs contributed by ESTs of other large	
chromosomal regions	74
4.3.2 CCV analysis of the wheat meiotic anther cDNA library	
4.4 Conclusions	80
Chapter 5 Transcript Profiling During Meiotic Development	82
5.1 Introduction	82
5.1.1 Microarray background	82
5.1.2 Prospects for plant meiotic gene discovery	85
5.1.3 Experimental design	
5.1.4 General considerations	87
5.2 Materials and methods	89
5.2.1 Preparation of amplified meiotic targets for microarray hybridisation	89
5.2.1.1 Meiotic anther collection	89
5.2.1.2 Total RNA and poly(A) RNA isolation	89
5.2.1.3 First and second strand cDNA synthesis	
5.2.1.4 <i>In vitro</i> transcription (amplification)	
5.2.1.5 Target labelling	
5.2.2 Preparation of microarray slides	
5.2.2.1 Amplification of probe sequences	
5.2.2.2 Robotic printing and slide blocking	
5.2.3 Hybridisation, washing and scanning	
5.2.4 Data analysis and quality control	
5.2.4.1 Signal intensity acquisition and pseudocolour inspection	
5.2.4.2 Data transformation and presentation	
5.2.4.3 Hierarchical clustering	99
5.3 Results and discussion	99
5.3.1 Microarray design	99
5.3.2 Data normalisation	103
5.3.2.1 Within slide lowess normalisation	103
5.3.2.2 Between slide normalisation	106
5.3.3 Verification and quality control of T7 amplification	108
5.3.3.1 A verification experiment	108
5.3.3.2 Keeping track of amplification	110
5.3.3.3 Expected yields from amplification, and aRNA size spread	112
5.3.4 Wild-type vs. <i>Ph</i> mutant differential expression	114
5.3.5 Temporal analysis of gene expression during meiosis	118
5.3.5.1 Consistency and verification of expression profiles	121
5.3.5.2 Analysis of the differentially expressed genes during meiosis	123
5.3.5.2.1 Hierarchical clustering	123
5.3.5.2.2 A comment on genes involved in cellular metabolism	126
5.3.5.2.3 Early expressed genes	128
5.3.5.2.3.1 Cluster group IV	128
5.3.5.2.3.2 Cluster group V	138
5.3.5.2.3.3 Cluster group II	141
5.3.5.2.4 Mid-late expressed genes	141
5.3.5.2.4.1 Cluster group III	141
5.3.5.2.5 Late expressed genes.	146
5.3.5.2.5.1 Cluster group VI	146

5.3.5.2.5.2 Cluster group I	
5.4 General discussion and conclusions	
5.4.1 <i>Ph</i> mutant vs. wild-type microarray experiments	
5.4.2 Temporal microarray experiments	
5.4.3 Experimental design	
Chapter 6 General Discussion	
Chapter 6 General Discussion	158 166

### **LIST OF FIGURES**

Figure 1.1:	Meiosis
Figure 1.2:	The synaptonemal complex7
Figure 1.3:	The double strand break repair model of meiotic recombination18
Figure 3.1:	Wheat ESTs identified from the rice chromosome 1 region syntenous to the $ph2a$ deletion in wheat
Figure 3.2:	Southern analysis locating clones to the region deleted in the wheat <i>ph2a</i> mutant
Figure 3.3:	Predicted polypeptide sequence and similarity alignment of two wheat ESTs identified as candidates for <i>Ph2</i> 63
Figure 4.1:	Weighted distribution of contig membership showing contigs with similarity to ESTs derived from the region deleted in the <i>ph2a</i> mutant
Figure 4.2:	Weighted distribution of contig membership contributed by ESTs derived from two chromosomally mapped EST bins
Figure 4.3:	Weighted distribution of contig membership contributed by ESTs from the wheat meiotic anther cDNA library
Figure 5.1:	A generalised scheme for the construction and screening of a cDNA microarray
Figure 5.2:	Procedure for linear T7 RNA amplification of microarray targets90
Figure 5.3:	The design of temporal series and <i>Ph</i> mutant vs. wild-type microarray experiments
Figure 5.4:	Pseudocolour image of Cy3/Cy5 signal intensities from microarray hybridisation
Figure 5.5:	<i>MA</i> -plots showing systematic sources of variation in microarray expression data and the effects of normalisation
Figure 5.6:	Side-by-side box plots showing the effect of between slide scale normalisation
Figure 5.7:	Experimental design to evaluate T7 RNA amplification

Figure 5.8:	Box-plot of normalised <i>M</i> -values obtained in the microarray experiment to evaluate T7 RNA amplification
Figure 5.9:	Quality control during T7 RNA amplification and a comparison of cDNA size distribution synthesised from poly(A) RNA and aRNA templates
Figure 5.10:	Side-by-side box plot showing the range of normalised <i>M</i> -values from <i>Ph</i> mutant genotypes <i>ph1b</i> , <i>ph2a</i> , and <i>ph2b</i> compared to wild-type Chinese Spring wheat
Figure 5.11:	The overlap in gene expression from <i>Ph</i> mutant vs. wild-type microarray experiments
Figure 5.12:	Expression profiles of all microarray probes from pre-meiosis to the tetrad stage of meiosis
Figure 5.13:	Consistency in expression profiles of microarray probes of predicted identical function
Figure 5.14:	A comparison of expression profiles derived from microarray and Northern hybridisation
Figure 5.15:	Hierarchical cluster analysis of 128 microarray probes showing greater than a two-fold change in at least one meiotic stage compared to the immature pollen reference tissue
Figure 5.16:	Temporal expression profiles of genes encoding metabolic enzymes
Figure 5.17:	Temporal expression profiles of genes from cluster group IV 130
Figure 5.18:	Temporal expression profiles of genes from cluster group V139
Figure 5.19:	Temporal expression profiles of genes from cluster group II142
Figure 5.20:	Temporal expression profiles of genes from cluster group III 143
Figure 5.21:	Temporal expression profiles of genes from cluster group VI 147
Figure 5.22:	Temporal expression profiles of genes from cluster group I 149

### LIST OF TABLES

Table 3.1:	Oligonucleotide primer sequences designed for amplification of selected wheat ESTs for Southern analysis
<b>Table 3.2</b> :	Details of wheat ESTs identified from the rice chromosome 1 region syntenous to the $ph2a$ deletion in wheat
<b>Table 3.3</b> :	Classification of identified wheat ESTs based on molecular function
Table 4.1:	Details of Triticeae cDNA libraries selected for CCV analysis 72
Table 5.1:	Wheat sequences selected for microarray printing

#### ABSTRACT

Meiosis is a cell division process central to the life cycle of all sexual eukaryotic organisms. Chromosome pairing, genetic recombination and subsequent nuclear division during meiosis produces four genetically distinct haploid gametes from a single diploid cell. Allohexaploid wheat (*Triticum aestivum*) behaves meiotically as a diploid, despite the existence in the genome of three closely related (homoeologous) genomes, A, B and D. Chromosome pairing during prophase I of meiosis in wheat is restricted to true homologous chromosomes, the result being the formation of 21 bivalents at meiotic metaphase I. The genetic control of chromosome pairing in wheat is under the control of several pairing <u>homoeologous</u> (*Ph*) genes, located predominantly on chromosome groups 3 and 5. The major suppressors of homoeologous pairing are *Ph1* and *Ph2*. Their cytogenetic effect has been intensively studied but at the molecular level little is known about their function. The isolation and characterisation of *Ph* genes from wheat would lead to greater understanding of chromosome pairing mechanisms in complex allopolyploids, and may enable development of effective strategies for alien gene introgression from related species to modern wheat cultivars.

In this study, several genomics-based approaches were adopted to explore the expressed portion of the wheat genome in order to identify and characterise genes that could function in the molecular processes regulating meiosis.

The first approach used comparative genetics to characterise the region deleted in the ph2a mutant (a deletion mutant at Ph2). The rice genomic region syntenous to that deleted in the ph2a mutant was identified through comparative mapping and used in searches of wheat databases to identify ESTs with significant similarity. Southern analysis confirmed a syntenous relationship in the wheat and rice genomic regions and defined precisely the position of the breakpoint in ph2a. What seems to be a terminal deletion on 3DS is estimated to be approximately 80 Mb in length. We can tentatively predict the identification of approximately 220 genes from the region deleted in ph2a. The putative role of identified candidate Ph2 genes is discussed.

The second approach explored the validity of recent proposals suggesting the presence of a meiotic gene cluster in the region of Ph2. The transcriptional characteristics of genes linked to Ph2 were investigated using data from wheat EST databases in combination with recently developed analysis software. The tissue-distribution of mRNAs derived from genes linked to Ph2 is shown to resemble that of other large chromosomal regions in the wheat genome. It is concluded that the apparently high number of genes from the Ph2 region expressed in wheat meiotic tissue is not indicative of a meiotic gene cluster in this region, but rather highlights the transcriptional complexity of meiotic anther tissue.

Finally, the meiotic expression pattern of approximately 1800 wheat genes was examined using cDNA microarrays. Two approaches were taken. Firstly, the applicability of microarrays to identify differentially expressed genes between wild-type anthers and anthers of three Ph mutant genotypes was investigated. These experiments failed to reveal significant down-regulation of genes in Ph mutant anthers compared to wild-type. Possible explanations are discussed. Secondly, the expression of all microarray clones was examined from pre-meiotic interphase through to the tetrad stage of meiosis. A number of candidate wheat genes involved in meiotic and anther developmental processes have been identified and are discussed.

Prior to this study, the methods available to identify wheat meiotic genes, in particular as candidates for Ph2, were limited. The recent development of genomics in plant biology provided an opportunity for a new approach towards gene discovery and genome structural analysis in relation to meiosis. This research illustrates the need for, and the effectiveness of a new approach to study meiosis, contributing to our knowledge of the structural and functional characteristics of genes linked to Ph2, and establishing a strong basis for further wheat meiotic gene characterisation.

#### DECLARATION

This work contains no material which has been accepted for the award of any other degree or diploma in any university or other tertiary institution and, to the best of my knowledge and belief, contains no material previously published or written by another person, except where due reference has been made in the text.

I give consent to this copy of my thesis, when deposited in the University Library, being available for loan and photocopying.

T. J. Sutton May 2003

#### ACKNOWLEDGEMENTS

A number of people and institutions have provided support and assistance throughout the course of this study. I wish to express my appreciation to the following people for their contribution.

My supervisors Professor Peter Langridge and Dr Andreas Houben for their patient supervision, guidance throughout the course of this study and critical reading of this thesis. I particularly thank Professor Peter Langridge for his support of my research ideas and goals and I am extremely grateful for the exciting opportunities available to me throughout the course of this research.

Research fellow Dr Ute Baumann, for the countless times that I have sought opinion and advice over the past years, and for valuable suggestion and solution to the bioinformatic challenges of my research.

All members of the Langridge laboratory and the Department of Plant Science who have helped me during the course of this research. In particular I thank the help of Dr Ryan Whitford as both a colleague and good friend, and Dr Jason Able for his critical reading of this thesis. From DuPont Ag Biotech I thank Nathan Uhlmann, Dr Antoni Rafalski, Dr Scott Tingey and Dr Petra Wolters for excellent assistance with microarrays and the practical aspects of my research visits to the US. From the US Department of Agriculture I thank Dr Gerard Lazo and Dr Olin Anderson for supplying cDNA clones, extensive sequencing of my cDNA libraries and associated bioinformatic analysis.

Finally, I extend my profound gratitude to my wife Kristen, and family John, Elaine and Craig for their understanding, support and encouragement.

This research was financially supported through an Australian Postgraduate Award provided through the University of Adelaide, a scholarship provided by the Cooperative Research Centre for Molecular Plant Breeding, and assistance from DuPont Ag Biotech.

#### **ABBREVIATIONS**

aa	amino acid
aRNA	antisense ribonucleic acid
ATP	adenosine 5'-triphosphate
BAC	bacterial artificial chromosome
bp	base pair/s
BLAST	Basic Logical Alignment Search Tool
BSA	bovine serum albumin
°C	degrees Celsius
CCV	Contig Constellation Viewer
cDNA	complementary deoxyribonucleic acid
cm	centimetre/s
CV	cultivar
Cy3	cyanine 3 dUTP
Cy5	cyanine 5 dUTP
dATP	2'-deoxyadenosine 5'-triphosphate
dCTP	2'-deoxycytidine 5'-triphosphate
dGTP	2'-deoxyguanosine 5'-triphosphate
dTTP	2'-deoxythymidine 5'-triphosphate
dUTP	2'-deoxyuridine 5'-triphosphate
DMSO	dimethyl sulfoxide
DNA	deoxyribonucleic acid
dNTP	deoxynucleoside triphosphate
DTT	diothiothreitol
EDTA	ethylenediaminetetraacetic acid
EMS	ethyl methane sulphonate
EST	expressed sequence tag
FISH	fluorescent in situ hybridisation
g	gram/s
x g	9.81 m/s <sup>2</sup>
h	hour/s
IPTG	isopropyl-1-thio-b-D-galactosidase

IQR	interquartile range
ITEC	International Triticeae EST Cooperative
Kb	kilobase/s
kDa	kilodalton/s
L	litre/s
LB	Luria-Bertaini
М	molar
mA	milliampere
Mb	megabase/s
min	minute/s
mg	milligram/s
mL	millilitre/s
mM	millimolar
ng	nanogram/s
nm	nanometre/s
MOPS	3-(N-morpholino)propane-sulfonic acid
mRNA	messenger ribonucleic acid
OD <sub>260</sub>	optical density at 260 nm
PAC	P1 artificial chromosome
PCR	polymerase chain reaction
poly(A)	polyadenylated
PVP	polyvinyl pyrollidone
RFLP	restriction-fragment length polymorphism
RNA	ribonucleic acid
RNase	ribonuclease
rpm	revolutions per minute
RT	room temperature
RT-PCR	reverse transcription-polymerase chain reaction
sarkosyl	N-lauroylsarcosine
SDS	sodium dodecyl sulphate
SSC	sodium chloride/sodium citrate
TAE	Tris/acetate/EDTA
Taq	Thermus aquaticus

TE	Tris/EDTA
Tris-HCl	Tris(hydroxymethyl)aminomethane hydrochloride
U	units
μFD	microfarad/s
μg	microgram/s
μL	microlitre/s
UV	ultraviolet
V	volt/s
$\mathbf{v}/\mathbf{v}$	volume/volume
W	watt/s
w/v	weight/volume
x-gal	5-bromo-4-chloro-3-indolyl-b-D-galactosidase