

Biology and ecology of the introduced
snail *Microxeromagna armillata* in
south eastern Australia

Angela L. Lush

B.Ag.Sc. Hons (University of Adelaide)

A thesis submitted for the Degree of Doctor of Philosophy

School of Agriculture, Food and Wine

University of Adelaide

Australia

January 2007





Frontispiece: Active *Microxeromagna armillata* adult (top) and inactive adult and juvenile *M. armillata* (bottom) in leaf litter of a citrus orchard.

Dedicated in loving memory to my dad,

Laurence Cowley Lush

3-4-1927 to 29-5-2006

Table of Contents

List of Figures	6
List of Tables	13
Abstract	16
Declaration	17
Acknowledgements.....	18
Chapter 1 General introduction and thesis outline.....	21
Chapter 2 Life history traits of <i>Microxeromagna armillata</i>	24
2.1 General Introduction.....	24
2.2 Development of a culture technique for <i>Microxeromagna armillata</i>	25
2.3 Reproductive lifespan, number and size of offspring, and the influence of body size on key fecundity characteristics of <i>Microxeromagna armillata</i>	28
2.4 Mode of reproduction	61
2.5 Can <i>Microxeromagna armillata</i> aestivate and does this affect its reproductive characteristics?	90
2.6 Does adult density affect the reproductive characteristics of <i>Microxeromagna armillata</i> ?	104
2.7 General Discussion.....	110
Chapter 3 Phenology and spatial distribution of <i>Microxeromagna armillata</i>	114
3.1 Introduction.....	114
3.2 Phenology of <i>Microxeromagna armillata</i>	115
3.3 Spatial distribution of <i>Microxeromagna armillata</i>	125
3.4 Summary	134

Chapter 4	Movement of <i>Microxeromagna armillata</i> in citrus orchards.....	136
4.1	Introduction.....	136
4.2	Presence of snails in the tree canopy	140
4.3	Development of method to study snail activity	143
4.4	Development of a method to estimate size of active snails.....	153
4.5	Activity and size of snails moving in the orchard	156
4.6	Prevention of snail movement	187
4.7	Discussion	199
Chapter 5	General Discussion	202
Appendix A	: Size-frequency distributions of shells and albumen gland lengths of <i>Microxeromagna armillata</i>	210
Appendix B	: Effect of copper trunk treatments on numbers of <i>Microxeromagna armillata</i> on the trunk and major branches of citrus trees.....	213
References	215

List of Figures

Figure 2-1: Polycarbonate vented jars used to culture <i>Microxeromagna armillata</i>	26
Figure 2-2: Expected relationships between life history traits of <i>Microxeromagna armillata</i> . Filled arrows represent a positive relationship, open arrows represent a negative relationship, and line connector represents no relationship.....	33
Figure 2-3: Mean number of eggs laid by <i>Microxeromagna armillata</i> individuals in 2002 and 2003 breeding seasons standardised for week of egg laying.....	42
Figure 2-4: Mean number of eggs laid per <i>Microxeromagna armillata</i> adult in 2002 as a function of initial shell diameter 2002 (adj $R^2 = 0.164$, $P = 0.014$).....	47
Figure 2-5: Mean number of eggs laid per <i>Microxeromagna armillata</i> adult in 2003 as a function of initial shell diameter in 2003 (adj $R^2 = 0.219$, $P = 0.007$).....	47
Figure 2-6: Onset of egg laying in 2002 as a function of initial experimental <i>Microxeromagna armillata</i> size 2002. (adj $R^2 = 0.389$, $P < 0.001$)	48
Figure 2-7: Relationship between the number of clutches and eggs laid per <i>Microxeromagna armillata</i> adult.....	48
Figure 2-8: Mean <i>Microxeromagna armillata</i> egg size recorded during each week of the experiment. Error bars represent standard error of the mean; note y-axis does not originate at zero.....	52
Figure 2-9: Effect of the number of <i>Microxeromagna armillata</i> adults per container on mean hatchling size in 2003. Bars represent standard error of the mean; note y-axis does not originate at zero.....	55
Figure 2-10: Relationships among life-history traits established for <i>Microxeromagna armillata</i> . Filled arrows represent positive relationships, open arrows represent negative relationships, patterned arrows represent a positive then negative relationship with successive year, line connector represents no relationship.	59
Figure 2-11: Proportion of single eggs to multiple egg clutches laid by paired and isolated <i>Microxeromagna armillata</i>	66

Figure 2-12: Egg hatching in single egg clutches laid by paired and isolated *Microxeromagna armillata*. 71

Figure 2-13: Initial shell diameter of *Microxeromagna armillata* across treatment groups (15/5/2003). SFS = self-fertilised progeny, single snail, SFP = self-fertilised progeny, snails in a pair, CFS = Cross-fertilised progeny, single snail, CFP = cross-fertilised progeny, snails in a pair. Error bars represent standard deviations: n = 20 for all groups, mean shell diameter used for snails within a pair. 75

Figure 2-14: Survival function for *Microxeromagna armillata*, self-fertilised progeny, isolated (SFS), self-fertilised progeny, paired (SFP), cross fertilised progeny, single (CFS) and cross fertilised progeny, paired (CFP) snails (n =20 for single snails and n=40 for paired snails). 76

Figure 2-15: Changes in mean shell diameter of *Microxeromagna armillata* in SFP (self fertilised progeny, pair) and CFP (cross fertilised progeny, pair) during the experiment. Error bars represent standard deviation; numbers next to points represent number of samples. * symbol between points indicates growth rates (slope of the lines) are significantly different between treatments. 78

Figure 2-16: Mean shell diameter of *Microxeromagna armillata* in SFS (single self fertilised snail) and CFS (single cross fertilised snail) during the experiment. Error bars represent standard deviations; numbers alongside points are number of samples. * symbol between points indicates growth rates (slope of lines) are significantly different between treatments. 79

Figure 2-17: Mean number of eggs laid per *Microxeromagna armillata* across treatment groups. SFS = self-fertilised progeny, single snail, SFP = self-fertilised progeny, snails in a pair, CFS = cross-fertilised progeny, single snail, CFP = cross-fertilised progeny, snails in a pair. Error bars represent standard deviations; the number above the bars represents sample size, while treatment groups with differing letters are significantly different at $p < 0.05$ 80

Figure 2-18: Mean number of clutches laid per *Microxeromagna armillata* across treatment groups. SFS = self-fertilised single snail, SFP = self-fertilised snails in a pair, CFS =

Cross-fertilised single snail, CFP = cross-fertilised snails in a pair. Error bars represent standard deviations; numbers above bars represents number of replicates.... 81

Figure 2-19: Mean number of eggs per clutch laid by *Microxeromagna armillata* across treatment groups. SFS = self-fertilised progeny, single snail, SFP = self-fertilised progeny, snails in a pair, CFS = Cross-fertilised progeny, single snail, CFP = cross-fertilised progeny, snails in a pair. Error bars represent standard deviation, numbers above bars represent sample size. Bars containing different letters are significantly different from each other..... 83

Figure 2-20: Mean egg and hatchling size of *Microxeromagna armillata* across treatment groups. SFS = self-fertilised progeny, single snail, SFP = self-fertilised progeny, snails in a pair, CFS = Cross-fertilised progeny, single snail, CFP = cross-fertilised progeny, snails in a pair. Error bars represent standard deviations; sample size is given above the bars. Different letters represent significantly different groups. 84

Figure 2-21: Percent egg hatching of *Microxeromagna armillata* across treatments. SFS = self-fertilised progeny, single snail, SFP = self-fertilised progeny, snails in a pair, CFS = Cross-fertilised progeny, single snail, CFP = cross-fertilised progeny, snails in a pair. Error bars represent standard deviations; sample size is given above the bars..... 85

Figure 2-22: Mean clutch hatch time of *Microxeromagna armillata* across treatment groups. SFS = self-fertilised progeny, single snail, SFP = self-fertilised progeny, snails in a pair, CFS = Cross-fertilised progeny, single snail, CFP = cross-fertilised progeny, snails in a pair. Error bars represent standard deviations; sample size is given above the bars. 86

Figure 2-23: Survival function for *Microxeromagna armillata* aestivated isolated (AS), aestivated paired (AP), cross fertilised single (CFS) and cross fertilised paired (CFP) snails (n=20 for single snails and n=40 for paired snails)..... 93

Figure 2-24: Mean initial shell diameter of *Microxeromagna armillata* across treatment groups. Error bars represent standard deviations; n = 20 for each group, mean shell diameter of snails within a pair used..... 94

Figure 2-25: Mean shell diameter of *Microxeromagna armillata* AS (aestivated, isolated) and CFS (cross fertilised, isolated) hatchlings over time. Error bars represent standard deviations; numbers next to points represent sample number; * symbol between points indicates growth rates (slopes of the lines) are significantly different between treatments. 95

Figure 2-26: Mean shell diameter of *Microxeromagna armillata* AP (aestivated, paired) and CFP (cross fertilised progeny, paired) snails over time. Error bars represent standard deviations; numbers near points represent sample number. * symbol between points indicates growth rates (slopes of the lines) are significantly different between treatments. 96

Figure 2-27: Percent hatching of *Microxeromagna armillata* clutches (all clutches and clutches where % hatch >0) across treatment groups. AS = aestivated isolated, AP = aestivated pair, CFS = cross-fertilised single, CFP cross-fertilised pair. Error bars represent standard error of the mean; numbers above bars indicate sample size. 98

Figure 2-28: Relationships between fecundity characteristics in control (CFP, top) and aestivated (AP, bottom) snails. Filled arrows represents a positive relationship, open arrows indicate a negative relationship..... 101

Figure 2-29: Containers used to investigate the effect of density on the fecundity characteristics of *Microxeromagna armillata*. The white powder visible in containers Y and R is the food source provided to *M. armillata*. 106

Figure 3-1: Density of live *Microxeromagna armillata* in leaf litter collected from a Riverland citrus orchard. Four 30cm² quadrats of leaf litter were sampled per tree for 10 trees on each sampling date. Error bars represent 95% confidence intervals. 119

Figure 3-2: Size frequency histograms of live *Microxeromagna armillata* from the Riverland orchard, June 1999 to August 2000. Note change of scale on 6/7/00 and 30/8/00..... 120

Figure 3-3: Length of albumen gland as a function of shell diameter for *Microxeromagna armillata* collected from a Riverland orchard..... 121

Figure 3-4: Density of live <i>Microxeromagna armillata</i> in leaf litter in a citrus orchard in Sunraysia. N = number of 30cm ² quadrats of leaf litter sampled on each sampling date. Error bars represent 95% confidence intervals; note scale on x-y axis differs from Figure 3-1.....	122
Figure 3-5: Size frequency histograms for live <i>Microxeromagna armillata</i> collected from the Sunraysia orchard, June 1999 to August 2000.....	123
Figure 3-6: Mean number of live <i>Microxeromagna armillata</i> per square metre in leaf litter and grassy sod. Error bars represent standard deviations.....	128
Figure 3-7: Length of albumen gland as a function of shell diameter for <i>Microxeromagna armillata</i> collected from a Sunraysia orchard in autumn and spring 2003.....	129
Figure 3-8: Size frequency histograms of <i>Microxeromagna armillata</i> collected in leaf litter from a Sunraysia orchard in autumn, winter and spring 2003.....	130
Figure 3-9: Size frequency histograms of <i>Microxeromagna armillata</i> collected in grass from a Sunraysia orchard in autumn, winter and spring 2003.....	131
Figure 3-10: Mean number of <i>Microxeromagna armillata</i> found on the trunk and major branches of Navel trees during autumn, winter and spring. Error bars represent standard deviation.	132
Figure 3-11: Mean number of <i>Microxeromagna armillata</i> found per canopy quadrat (n = 10 quadrats from 20 trees). Error bars represent standard deviations.	133
Figure 3-12: Mean number of <i>Microxeromagna armillata</i> found per piece of fallen fruit underneath the tree canopy (n=20 trees). Error bars represent standard deviations. ...	133
Figure 4-1: Mean number of <i>Microxeromagna armillata</i> on the trunk and major branches of Navel orange trees (block 1) in Nangiloc, Victoria. Error bars represent standard deviations, n = 30 trees.....	141
Figure 4-2: Mean number of <i>Microxeromagna armillata</i> on trunk and major branches of Navel orange trees (block 2) in Nangiloc, Victoria. Error bars represent standard deviations, n = 10 trees.	142

Figure 4-3: Relationship between <i>Microxeromagna armillata</i> shell diameter and trail width. Error bars represent standard deviation of measurements within a trail.	154
Figure 4-4: Inverse prediction of shell diameter based on trail width (90% CI).....	155
Figure 4-5: Navel tree banded with flagging tape to assess snail movement in the canopy (left) and tile banded with flagging tape to assess snail activity in the leaf litter.	160
Figure 4-6: Mean estimated shell diameter of <i>Microxeromagna armillata</i> active on different orchard surfaces pre-harvest. Error bars represent standard error; numbers above bars represent the sample size.	164
Figure 4-7: Size distribution of <i>Microxeromagna armillata</i> active on different orchard surfaces during pre-harvest	166
Figure 4-8: Mean number of <i>Microxeromagna armillata</i> trails per tape on different orchard surfaces during harvest. Error bars represent standard deviation; n = 30 trees.....	170
Figure 4-9: Mean estimated shell diameter of <i>Microxeromagna armillata</i> active on different orchard surfaces during harvest. Error bars represent standard error; numbers above columns represent sample sizes.	173
Figure 4-10: Size distribution of <i>Microxeromagna armillata</i> active on different orchard surfaces during harvest.....	174
Figure 4-11: Mean number of <i>Microxeromagna armillata</i> trails on different tree surfaces during one week in Spring. Error bars represent standard error, n = 20 for all surfaces, differing letters above columns indicate significant differences.	180
Figure 4-12: Mean estimated shell diameter of active <i>Microxeromagna armillata</i> on different orchard surfaces in spring. Error bars represent standard error; numbers in columns are the number of snail trails used to estimate snail size.	182
Figure 4-13: Size distribution of <i>Microxeromagna armillata</i> active on different orchard surfaces in spring	183

Figure 4-14: Mean number of <i>Microxeromagna armillata</i> trails found per day across orchard surfaces and seasons. Error bars represent standard error; numbers above columns represent sample sizes.	185
Figure 4-15: Estimated shell diameter of <i>Microxeromagna armillata</i> in pre-harvest (autumn), harvest (winter) and post harvest (spring) active on different orchard surfaces. Error bars represent standard error; numbers in columns represent the number of trails sampled to estimate snail size.	186
Figure 4-16: Copper band applied to the trunk of a Navel orange tree	189
Figure 4-17: Mean number of <i>Microxeromagna armillata</i> snails counted on the trunk and major branches of Navel orange trees after application of copper bands and Socusil. Stars indicate sampling dates where treatments are significantly different, n = 30 at all dates. Standard deviations and test statistics are presented in Appendix B.	190
Figure 4-18: Copper adhesive tape (a) and copper foil bands (b) applied to Navel orange tree trunks in winter. White flagging tape was placed below and in between the copper bands to assess snail activity.....	192
Figure 5-1: Desiccated eggs of <i>Microxeromagna armillata</i>	208
Figure A-1: Size frequency histograms of dead <i>Microxeromagna armillata</i> collected from the Riverland orchard, June 1999 to August 2000.....	210
Figure A-2: Size frequency histograms of dead <i>Microxeromagna armillata</i> collected from the Sunraysia orchard, June 1999 to August 2000.....	211
Figure A-3: Length of albumen gland as a function of shell diameter for <i>Microxeromagna armillata</i> collected from a Sunraysia orchard.....	212

List of Tables

Table 2-1: Mean number of <i>Microxeromagna armillata</i> eggs laid per snail pair on different substrates.....	27
Table 2-2: Number of <i>Microxeromagna armillata</i> eggs measured per egg clutch.....	35
Table 2-3: Reproductive traits of <i>Microxeromagna armillata</i> recorded over 75 weeks.....	38
Table 2-4: Summary of reproductive characters of small sized snail species living in leaf litter. Reproduced from Baur [38].....	40
Table 2-5: Reproductive characteristics of <i>Microxeromagna armillata</i> in 2002 and 2003. Mean values significantly different between years are followed by different letters and highlighted in bold type.....	43
Table 2-6: Linear regression analysis of the impact of <i>Microxeromagna armillata</i> size and successive reproductive events on number of eggs per clutch in 2002 and 2003.....	49
Table 2-7: Linear regression analysis of fecundity characteristics which significantly predicted mean egg size of <i>Microxeromagna armillata</i> in both 2002 and 2003.....	50
Table 2-8: Linear regression of fecundity characteristics with mean hatchling size of <i>Microxeromagna armillata</i> in 2003.....	53
Table 2-9: Reproductive characteristics of isolated and paired <i>Microxeromagna armillata</i> . Mean values that are significantly different are followed by different letters and highlighted in bold type.....	65
Table 2-10: Linear regression analysis of characteristics which can significantly predict mean egg size in isolated and paired <i>Microxeromagna armillata</i>	68
Table 2-11: Linear regression analysis of characteristics which can significantly predict mean hatchling size in isolated and paired <i>Microxeromagna armillata</i>	69
Table 2-12: Reproductive characteristics of aestivated and non –aestivated <i>Microxeromagna armillata</i> , which have been isolated or paired. AS = aestivated isolated, AP = aestivated pair, CFS = cross-fertilised single (non-aestivated), CFP cross-fertilised	

pair (non-aestivated). Mean values are significantly different when followed by different letters and are highlighted in bold type.	97
Table 2-13: Multiple linear regression analysis of the interrelatedness of fecundity characteristics of aestivated, paired <i>Microxeromagna armillata</i> (AP).	100
Table 2-14: Life history traits of <i>Microxeromagna armillata</i> at different adult densities.	109
Table 3-1: Orchard substrates sampled, and sampling methods used, to examine the distribution of <i>Microxeromagna armillata</i> in a citrus orchard	126
Table 4-1: Treatment schedule of tape strips stained on day 0 to 32.	146
Table 4-2: Number of mucus trails stained on flagging tape with Alcian blue at different concentrations and immersion times. (Number of strips per treatment combination =5)	148
Table 4-3: Number of tape strips showing stained mucus trails after exposure to sunlight, irrigation and dust treatments. C – clear trails, F – faint trails, NT – no trails	149
Table 4-4: Proportion of mucus trails stained on flagging tape after exposure to field conditions on a tree-surface (T), upper- tile surface (UL) and lower- tile surface (LL). n = number of tape strips examined, s = number of stained trails, p = number of pre-marked trails.	150
Table 4-5: Summary of analysis using a log-linear model on the number of stained and non-stained trails recorded after differing lengths of field exposure and application surfaces	150
Table 4-6: Mean number of <i>Microxeromagna armillata</i> mucus trails (\pm sd) stained on flagging tape applied to different orchard surfaces and after varying field exposure times. n=5 for all combinations.	161
Table 4-7: ANOVA of number of stained trails per day for trunk and lower branches (tree surface) at day 13 and 26 (time of exposure)	163
Table 4-8: Analysis of variance of estimated snail size between orchard surfaces pre-harvest.	165

Table 4-9: Pearson correlations between numbers of stained trails per orchard surface during harvest (17/6/2003 – 24/6/2003). Significant correlations are highlighted in bold, n = 30.....	171
Table 4-10: Pearson correlation of the numbers of <i>Microxeromagna armillata</i> counted on the trunk and major branches at band application and number of snail trails per day across orchard surfaces during harvest. Significant correlations are highlighted in bold, n = 30	172
Table 4-11: Analysis of variance of estimated <i>Microxeromagna armillata</i> size between orchard surfaces during harvest (log transformed data).	173
Table 4-12: Pearson’s correlations of activity (number of trails) on different orchard surfaces. Significant correlations are highlighted in bold.....	179
Table 4-13: Analysis of variance for number of <i>Microxeromagna armillata</i> trails stained per tree surface	180
Table 4-14: Correlation between number of <i>Microxeromagna armillata</i> on the trunk and major branches and <i>Microxeromagna armillata</i> activity (stained trails) on different orchard surfaces in Spring. Significant relationships are highlighted in bold type.....	181
Table 4-15: Mean number of <i>Microxeromagna armillata</i> trails on flagging tape placed below, in between, or above copper bands on Navel tree trunks. Means ± standard deviation followed by differing letters are significantly different across rows, n=5 for all samples except those marked with *, where n = 30.....	193
Table B-1: Mean number of <i>Microxeromagna armillata</i> on the trunk and major branches of Navel trees after application of copper bands and Socusil. Comparison of treatments within each date was made using analysis of variance (Tukey’s post-hoc testing) or Kruskal-Wallis non-parametric analysis of variance as appropriate. Means (± standard deviation) followed by differing letters are significantly different and are highlighted in bold type, total n = 30 (n=10 each treatment) and df =2 on all dates.....	213

Abstract

Microxeromagna armillata (Lowe, 1852) is a snail introduced snail to Australia which has established populations in the Riverland and Sunraysia citrus growing regions. Citrus exported from these regions to the USA has been rejected due to contamination with *M. armillata*, causing significant economic losses. The life history, phenology and activity of *Microxeromagna armillata* has not been studied in Australia: this forms the basis of this thesis.

Microxeromagna armillata employs an iteroparous egg laying strategy in semi-field conditions and lays approximately 500 eggs per year. Field populations can reach high densities (~4000 snails/m²), particularly during the winter months when juvenile recruitment occurs. Snails reach sexual maturity at ~ 6mm in shell diameter and can grow to this size from a juvenile stage (2mm) within six weeks. *Microxeromagna armillata* can reproduce successfully by self-fertilisation, and juveniles are able to aestivate with little reduction in subsequent fecundity. These traits make control of this pest a significant challenge. Leaf litter is the preferred habitat of *M. armillata*, but snails do move in the tree canopy. Cues for snail activity in the leaf litter and canopy appear to differ, as does the size of active snails in these areas. *Microxeromagna armillata* activity was low in the tree canopy during harvest compared to post harvest, intimating that fruit contamination is either occurring infrequently or post-harvest. Copper trunk bands were shown to minimise snail movement into the canopy and may be an important preventative measure.

These findings have changed the recommendations for *M. armillata* management in citrus groves of south eastern Australia.

Declaration

This work contains no material which has been accepted for the award of any other degree or diploma in any university or other tertiary institution and, to the best of my knowledge and belief, contains no material previously published or written by another person, except where due reference has been made in the text.

I give consent to this copy of my thesis, when deposited in the University Library, being made available for loan and photocopying, subject to the provisions of the Copyright Act 1968.

Portions of this research have been published by the author in:

Lush, A.L. (2001). The biology, ecology and control of the small brown snail *Microxeromagna armillata* in citrus orchards of south eastern Australia. Final Report for the Citrus Market Development Group; Agriculture, Fisheries and Forestry Australia. South Australian Research and Development Institute (SARDI) pp 39.

Lush, A.L. and Baker, G.H. (2003) A novel method for the field study of mollusc movement using mucus trails. *In* Slugs and snails: Agricultural, Veterinary and Environmental Perspectives. BCPC Symposium Proceedings No. 80

Lush, A.L. (2004). Field and postharvest control of snails in citrus. Final report for Horticulture Australia Ltd project CT00040. South Australian Research and Development Institute (SARDI) pp 69

This work was funded by a grant to the author from Horticulture Australia Ltd, administered through the South Australian Research and Development Institute (SARDI).

Angela L. Lush

January, 2007

Acknowledgements

The research in this thesis was funded by a Horticulture Australia Ltd grant (CT00040) to the author whilst in the employ of the South Australian Research and Development Institute (SARDI). A lesser portion of the research was funded by the Citrus Market Development Group, Agriculture Fisheries and Forestry Australia, also whilst employed at SARDI. Thank you in particular to Gerard McEvelly from Horticulture Australia Ltd for suggesting I use this grant to gain a PhD whilst fulfilling the project outcomes for industry, and Dennis Hopkins (SARDI) for approving my request.

Thanks go to my supervisors Assoc. Prof. Michael Keller (University of Adelaide) and Dr. Geoff Baker (CSIRO) for their advice, time, encouragement and support. Mike, thanks for being so encouraging when I decided to leave SARDI, not to mention providing me with office space in order for me to finish writing up. Thanks to Geoff in particular for letting me 'run away' to his lab in Canberra when I needed to nut out a problem and gain some perspective. I always returned from these visits with a clearer focus and renewed enthusiasm. Thanks also go to the Entales lab group, in particular Katja Hoogendorn for her support and friendship.

I received a great deal of support from my SARDI colleagues throughout my PhD and would like to thank them very much for their assistance and encouragement. Particular thanks go to Peter Taverner for giving me the inside info on the citrus industry, helping me to write the grant (not to mention introducing me to the joys of the steering committee), offering good advice and great conversation on shared field trips. My long suffering former office mates Gabriella Caon and Dijana Jevremov deserve medals for tolerating my foibles, 'quick questions', and interruptions, and Nancy 'the go to girl' Cunningham was invaluable in helping me unravel the mysteries of SARDI. Throughout my PhD, I was fortunate to have technical assistance along the way and thanks go to all the casual staff members who helped me complete my research. Special thanks to Bill Kimber, David Bigham and the snail girls – Julie Lindner, Susan Ivory and Nola 'the warbler' Lucas. Who knew counting eggs could be so

much fun! Former colleague and now friend Suzanne Charwat helped immeasurably in discussions on snail behaviour and the world snail scene, as well as being a great travelling companion.

Nancy Schellhorn has had a huge influence on me during my PhD and I would like to thank Nancy for her encouragement and support. She helped me to see what real science and collaboration is all about, and was extremely generous with her time and advice, both on a professional and personal level. Nancy on more than one occasion helped me with a problem well into the night, and often saved me from certain statistical disaster, always with patience and encouragement. Her approach to problem solving, formulating focused research questions, experimental design, and all things statistical has taught me an immeasurable amount. No-one is more surprised than I am when suddenly I start advising others on which statistical methods they should be using and why - this is in no small part due to Nancy.

Cate Paull and Judy Bellati have also been an integral part of my PhD. Without their support I am doubtful that I would have made it this far. Cate and Judy were always encouraging and ready with a huge smile or a shoulder to cry on depending on the situation. For everything from providing a taxi service when I was incapacitated, letting me 'vent', provision of chocolates, to helping me answer the 'oh my god, what am I going to do' questions – THANK YOU! I can't imagine what it would have been like without you two and you have helped me more than I can say.

I would also like to say a big thank you to my family for all their support. Mum and Dad have always been encouraging and I'm very grateful for the opportunities they have given me and their unerring belief in me. I'm very lucky to have such great parents. My sisters in Adelaide, Sonya and Roslyn, have helped me an enormous amount in many different ways. From calling ambulances to climbing through windows to rescue me and doing my washing, they have gone above and beyond the call! Thanks guys.

Thanks to Renee and Kaylee for coming out on the town with me when I needed to let loose, reminding me of 'the normal life', and understanding when I couldn't participate. I couldn't ask for better friends, Cheers.

My sincere thanks also go to the following people:

Rodney Hand, for letting me experiment on his property, his friendly welcome (and more importantly the friendly dogs), the good conversations and advice on snail management from the grower's perspective, and the exceptional pistachio nuts. Peter Morrish, for showing me around the Sunraysia district, introducing me to growers, directing me over the phone when I got lost, chairing my steering committee, excellent organisation of grower meetings, and for his overall support of me and my project. Kym Theil, for also directing me over the phone when I got lost on the way to grower meetings, participating in my steering committee and helping me hunt for small brown snails. Megan Leyson, for her company on the great UK snail adventure, snail related conversations, and for helping me realise that doing a PhD and having a social life were not altogether mutually exclusive. Sheridan Purvis, for her company on field days and CITTgroup extravaganzas. Vanessa Carne-Cavagnaro, for introducing me to Entales and her support at the beginning of my journey. Steve and Violet Csuturos, for welcoming me with a shot of ouzo on arrival (and the occasional schnapps with the early breakfast??) at their Redcliffs motel: after a six hour drive it ensured I slept very well. The Redcliffs pub, for the best (and cheapest) steaks ever. Kerrie and Valerie, for listening over coffee to my results and progress. Ian Riley for his support as post-graduate co-ordinator. Derek, for doing my dishes, mowing my lawn and baking the most spectacular chocolate brownies. Nathan and Kaye, for helping me over the last hurdles and supporting me during my dad's illness.

Please note: in the print copies the only pages coloured are the ones with photographs.