Effect of growth factors on the osteoinductive potential of Hydroxyapatite β -Tricalcium Phosphate (HA-TCP)

A report submitted to the University of Adelaide in partial fulfilment of the requirements of the Degree of Doctor of Clinical Dentistry

(Periodontology)

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3. Appendices

3.1 Abbreviations

ALP Alkaline Phosphatase

BCP Biphasic Calcium Phosphate
BMP- Bone Morphogenetic ProteinBMP Bone Morphogenetic Proteins

CaP Calcium Phosphate

Ca/P ratio Calcium/Phosphate ratio
CdA Calcium deficient Apatite

DBBM Deproteinised Bovine Bone Matrix

DFDBA Demineralised Freeze Dried Bone Allograft

EMD Enamel Matrix Protein Derivative

EMP Enamel Matrix Protein

ePTFE Expanded Polytetrafluoroethylene

FDBA Freeze Dried Bone Allograft
FGF Fibroblast Growth Factor

GBR Guided Bone Regeneration

HA Hydroxyapatite

HA-TCP Hydroxyapatite-Tricalcium Phosphate

ICTP Carboxyterminal Telopeptide of Type I collagen

IGF- Insulin-like Growth Factor-MSCs Mesenchymal Stem Cells

PDGF- Platelet Derived Growth Factor-

PDL Periodontal Ligament

PGA Propylene Glycol Alginate

rhPDGF- Recombinant human Platelet Derived Growth Factor-

 β -TCP β -Tricalcium Phosphate

TGF- Transforming Growth Factor-

VEGF Vascular Endothelial Growth Factor

vSMC Vascular Smooth Muscle Cell

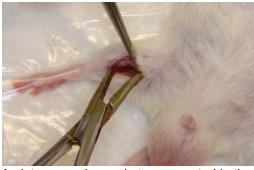
3.2 Appendix One: Surgical Protocol



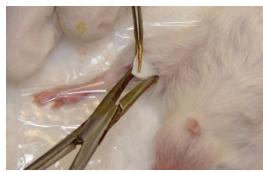
1. Inhalation anaesthesia of the animal was maintained using a nose cone fitted to a rodent stereotactic frame.



2. After disinfection, a skin incision was made on the medial aspect of both hindlegs.



3. An intramuscular pocket was created in the quadriceps muscle using blunt dissection.



4. A half gelatine capsule containing the graft was placed within the intramuscular pocket.



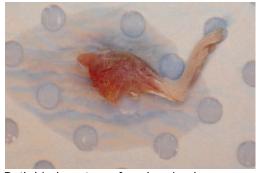
placement within the intramuscular closed with metal staples. proper pocket.



5. The implanted graft was checked to ensure 6. After implantation of the graft, the incision was

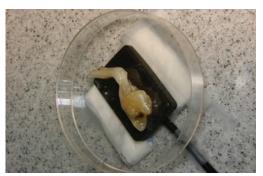
3.3 Appendix Two: Retrieval Protocol





1. At the end of the implantation period, the animal was sacrificed by means of CO₂ removed followed by removal of skin and fur. asphyxiation.

3.4 Appendix Three: Radiographic Protocol

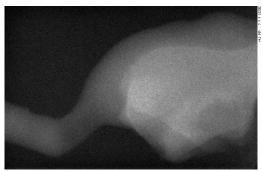


1. The specimen was placed on a device incorporating the dental radiography sensor.



2. The dental radiographic tube was positioned directly above the sensor at a standardised distance for all samples.

3.5 Appendix Four: Histological Preparation Protocol



1. After decalcification, radiographs were taken of the hindlimbs to ensure complete decalcification of the specimen



2. Prior to embedding, the paw and superior portion of the specimen were excised and discarded. The remaining specimen was sectioned in half according to the location of the graft material as determined by the post retrieval radiographs.



3. The sectioned specimens were oriented with the adjacent cut surfaces facing superiorly.



4. The specimens were paraffin embedded in this orientation.

3.6 Appendix Five: Semi-quantitative analysis of experimental groups at 4 weeks

	Category	HA-TCP	HA-TCP + PDGF	HA-TCP + EMD
	0	5	5	10
Acute Inflammation	1	7	10	5
	2	0	0	0
	3	0	0	0
	4	0	0	0
Chronic Inflammation	0	0	0	0
	1	2	9	3
	2	10	6	11
	3	0	0	1
	4	0	0	0
	0	2	3	4
Resorption/Foreign Body Reaction	1	10	12	11
	2	0	0	0
	3	0	0	0
	4	0	0	0
	0	0	0	0
	1	0	2	0
Fibrosis - Distribution	2	0	2	0
	3	6	4	2
	4	6	7	13
	0	0	0	0
	1	0	2	0
Fibrosis - Density	2	2	8	1
	3	8	5	4
	4	2	0	9
	0	0	0	0
	1	2	7	
Vascularity - Distribution		7		6
	2		4	5
	3	3	2	4
	4	0	2	0
	0	0	0	0
Vascularity - Area	1	5	5	9
	2	3	4	5
	3	4	1	0
	4	0	5	1
Adipose - Distribution	0	6	5	13
	1	4	3	1
	2	2	1	1
	3	0	6	0
	4	0	0	0
Adipose - Area	0	6	5	13
	1	4	2	1
	2	1	1	0
	3	<u> </u>	2	1
	4	0	5	0
		0	0	
	0			0
I factor and the first	1	3	10	5
Lining cell thickness	2	7	5	8
	3	2	0	2
	4	0	0	0
	0	12	15	15
Osteoinduction	1	0	0	0
	2	0	0	0
	3	0	0	0
1	4	0	0	0

Numbers represent the number of analysed sections included in each category at 4 weeks

3.7 Appendix Six: Semi-quantitative analysis of experimental groups at 8 weeks

8 WEEK	Category	HA-TCP	HA-TCP + PDGF	HA-TCP + EMD
Acute Inflammation	0	14	12	13
	1	1	1	2
	2	0	0	0
	3	0	0	0
	4	0	0	0
Chronic Inflammation	0	10	3	6
	1	5	10	9
	2	0	0	0
	3	0	0	0
	4	0	0	0
Resorption/Foreign Body Reaction	0	13	10	13
	1	2	3	2
	2	0	0	0
	3	0	0	0
	4	0	0	0
Fibrosis - Distribution	0	0	0	0
	1	0	1	0
	2	0	1	0
	3	4	7	11
	4	11	4	4
	0	0	0	0
	1	3	1	1
Fibrosis - Density	2	3	3	3
	3	7	8	8
	4	2	1	3
	0	3	0	1
	1	2	2	6
Vascularity - Distribution	2	6	3	6
	3	4	8	2
	4	0	0	0
	0	3	0	1
Vascularity - Area	1	5	6	12
	2	7	3	1
	3	0	4	1
	4	0	0	0
Adipose - Distribution	0	11	9	3
	1	3	3	1
	2	0	0	5
	3	1	1	5
	4	0	0	1
Adipose - Area	0	11	9	3
	1	3	3	2
	2	0	0	1
	3	1	0	4
	4	0	1	5
Lining cell thickness	0	0	0	0
	1	12	6	11
	2	1	7	3
	3	2	0	1
	4	0	0	0
Osteoinduction	0	15	13	15
	1	0	0	0
	2	0	0	0
	3	0	0	0
	4	0	0	0

Numbers represent the number of analysed sections included in each category at 8 weeks