# GENOTOXICITY INVESTIGATION OF ORGANIC N-CHLORAMINES

Somprasong Laingam (MBiotech)

Discipline of Pharmacology

The University of Adelaide

(Faculty of Health Science)

July, 2009

A thesis submitted for the degree of Doctor of Philosophy

## TABLE OF CONTENTS

ABSTRACT	Γ	i
DECLARA	ΓΙΟΝ	iii
ACKNOWL	LEDGEMENTS	iv
PUBLICAT	TIONS IN SUPPORT THIS THESIS	V
ABBREVIA	ATIONS AND SYMBOLS	vi
1. GENE	RAL INTRODUCTION	1
1.1. Wa	ater Disinfection	3
1.1.1.	Chlorine	4
1.1.2.	Chloramine	5
1.1.3.	Chlorine dioxide	6
1.1.4.	Ozone	7
1.2. Dis	sinfection By-products (DBPs)	8
1.2.1.	Trihalomethanes	9
1.2.2.	Haloacetic acids	9
1.2.3.	Bromate	10
1.2.4.	Chlorite	11
1.2.5.	Haloacetonitrile	11
1.2.6.	Halofuranones	12
1.2.7.	Emerging DBPs	15

	1.3.	Health	Risks Associated With DBPs	16
	1.3	.1. D	BPs and Bladder Cancer	16
	1.3	.2. D	BPs and Adverse Reproductive Outcomes	18
	1.4.	Toxico	ological Studies of DBPs	20
	1.5.	Early I	Prioritisation Studies of DBP Research	21
	1.6.	Recent	Prioritisation Study of DBP Research	24
2.	OR	RGANIO	C N-CHLORAMINES AND RESEARCH AIMS	26
	2.1.	Organi	ic Nitrogen in Natural Water	26
	2.2.	Water	Disinfection and Formation of Organic N-Chloramines	28
	2.3.	Toxico	ological Significance of Organic N-chloramines	29
	2.4.	Organi	ic N-Chloramines and Bladder Cancer Risk	34
	2.5.	Signifi	cance of Organic N-Chloramines: Australian Perspectives	34
	2.6.	Projec	t Rationale	36
3.	FL	OW CY	TOMETRY-BASED MICRONUCLEUS ASSAY	38
	3.1.	Introdu	action	38
	3.2.	Materi	als and Methods	41
	3.2	.1. Cl	nemicals and Reagents	41
	3.2	.2. Ce	ell Culture	43
	,	3.2.2.1.	General Maintenance	43
	,	3.2.2.2.	Measurement of Cell Growth	44
	3.2	.3. Fl	ow Cytometry Settings for Measurement of Micronuclei	46
	,	3.2.3.1.	Flow Cytometry Settings	46
	,	3.2.3.2.	Gating Criteria for Measurement of Micronuclei	48
	,	3.2.3.3.	Confirmation of Micronucleus by Flow Cytometry Sorting	49

3.2.4. M	Iicronucleus Assay	50
3.2.4.1.	Chemical Treatment	50
3.2.4.2.	Analysis of MN by Flow Cytometry	52
3.2.4.3.	Analysis of MN by Microscopy	53
3.2.5. M	ITS-Cell Viability Assay	54
3.2.6. A	poptosis Measurement	54
3.2.6.1.	Apoptosis Induction	54
3.2.6.2.	Flow Cytometric NucView Assay	55
3.2.7. S	tatistical Analysis	55
3.3. Result	ss	56
3.3.1. D	oubling Times of WIL2-NS and L5178Y Cell Lines	56
3.3.2. O	ptimisation of the FCMN Assay	57
3.3.2.1.	Flow Cytometry Parameter Settings	57
3.3.2.2.	Flow Cytometric Sorting of Nuclei/Micronuclei	59
3.3.3. F	CMN Assay Validation	61
3.3.3.1.	Spontaneous MN Formation (no-treatment control)	61
3.3.3.2.	Treatment with Mitomycin- C (MMC)	62
3.3.3.3.	Treatment with Methyl Methanesulfonate (MMS)	62
3.3.3.4.	Treatment with Vinblastine (VINB)	63
3.3.3.5.	Treatment with Etoposide (ETOPO)	64
3.3.3.6.	Treatment with Benzo[a]pyrene (BaP)	65
3.3.3.7.	Treatment with Sucrose (SUCRO)	66
3.3.4. C	omparison of Apoptosis Response	69
3.4. Discu	ssion	72

4.	PR	RODU	JCTION OF ORGANIC N-CHLORAMINES	76
	4.1.	Intr	oduction	76
	4.2.	Mat	terials and Methods	81
	4.2	2.1.	Chemicals and Reagents	81
	4.2	2.2.	Production of Organic N-chloramines	82
	4.2	2.3.	Measurement of Chlorine Residual	83
	4.2	2.4.	Measurement of Chloramines	83
	4.3.	Res	ults	84
	4.3	3.1.	Optimal Chlorine Contact Time	84
	4.3	3.2.	Optimal Precursor Concentration	85
	4.4.	Dis	cussion	88
5.	TO	XIC	ITY AND GENOTOXICITY OF ORGANIC N-CHLORAMINES	90
	5.1.	Intr	oduction	90
	5.2.	Mat	terials and Methods	92
	5.2	2.1.	Chemicals	92
	5.2	2.2.	Cell Culture and Seeding Preparation	93
	5.2	2.3.	Cell Treatment – Optimisation for Short Term Exposure	93
		5.2.3	.1. Selection of Treatment Medium	93
		5.2.3	.2. Comparison between Short Term Exposure vs. Continuous Exposure	94
	5.2	2.4.	Cell Treatment for Genoxicity Studies of N-chloramines	95
	5.2	2.5.	Measurement of Cytotoxicity by MTS assay	97
	5.2	2.6.	Measurement of MN Formation by Flow Cytometry	97
	5.2	2.7.	Measurement of MN Formation by Microscopy	97
	5.2	2.8.	Statistical Analysis	98
	5.3.	Res	ults	99

5.3.1.	Short-term Exposure to N-chloramines	99
5.3.	1.1. Selection of Treatment Medium	99
5.3.	1.2. Comparison of Continuous vs. Short-term Exposure Treatme	ent 101
5.3.2.	Cytotoxicity of Organic N-chloramines	102
5.3.3.	Genotoxicity of Organic N-chloramines	105
5.4. Di	scussion	109
	NIC N-CHLORAMINES AND THE ROLE OF OXIDATIVE	
	roduction	
	aterials and Methods	
6.2.1.	Chemicals and Materials	
6.2.2.		
	Classification Asserts	
6.2.3.	Glutathione Assay	
6.2.	3.1. Chemical Exposure	
6.2.	3.2. Sample Preparation and Measurement of Total Glutathione	118
6.2.4.	Malondialdehyde Assay	120
6.2.	4.1. Chemical Exposure	120
6.2.	4.2. Sample Preparation and Measurement of MDA	120
6.2.5.	Statistical Analysis	121
6.3. Re	esults	122
6.3.1.	Glutathione Depletion	122
6.3.	1.1. Glutathione Standard Curve	122
6.3.	1.2. Reduction of Cellular Glutathione Levels Following FeSO <sub>4</sub>	Γreatment124
6.3.	1.3. Effect of Organic N-chloramines on Cellular Glutathione Le	vels 125
6.3.2.	Analysis of Lipid Peroxidation Products	127

6.3.2.1. MDA Standard Curve	27
6.3.2.2. Effect of FeSO <sub>4</sub> on MDA Production	29
6.3.2.3. Effects of Organic N-chloramines on Lipid Peroxidation	30
6.4. Discussion	32
APPLICATION OF CELL BASED ASSAY FOR IDENTIFICATION OF	F
PRECURSORS OF TOXIC DBPs IN AUSTRALIAN WATER 1	35
7.1. Introduction	35
7.2. Materials and Methods	37
7.2.1. Water Sample and Preparation of MW Fractions	37
7.2.2. Chlorination of MW Fractions	38
7.2.2.1. Modified Colorimetric DPD Method	38
7.2.2.2. Determination of Chlorine Dose for Chlorination of MW Fractions 1	39
7.2.2.3. Chlorination of MW Fractions for Cytoxicity and Genotoxicity Assays1	40
7.2.3. Cytotoxicity and Genotoxicity of Chlorinated MW Fractions	40
7.2.3.1. Cell Culture and Seeding Preparation	40
7.2.3.2. Cell Treatment	41
7.2.3.3. Cytotoxicity Measurement	42
7.2.3.4. Measurement of MN Formation by Flow Cytometry	42
7.2.3.5. Measurement of MN Formation by Microscopy	42
7.2.4. Data Analysis	43
7.3. Results	44
7.3.1. Preparation of MW Fractions1	44
7.3.2. Chlorination of MW Fractions	45
7.3.2.1. Modified Colorimetric Microplate DPD Assay	45
7.3.2.2. Determination of Chlorine Dose for Chlorination of MW Fractions 1	46

	7.	3.3.	Cy	totoxicity and Genotoxicity of Pre-chlorinated MW Fractions	147
	7.	3.4.	Cyt	totoxicity and Genotoxicity of Chlorinated MW Fractions	148
		7.3.4	.1.	Cytotoxicity of Chlorinated MW Fractions	148
		7.3.4	.2.	Genotoxicity of Chlorinated MW Fractions	150
7	'.4.	Disc	cuss	ion	152
8.	G	ENER	RAL	DISCUSSION	156
8	3.1.	Esta	ablis	shment of the FCMN Assay	156
8	3.2.	Gen	oto	xicity of Organic N-chloramines	159
8	3.3.	Sign	nific	cance of This Study to Water Research in Australia	164
8	3.4.	Con	clus	sion	165
8	3.5.	Futi	ıre l	Research Direction	166
API	PEN	DICE	S		168
BIB	LIC	<b>OGRA</b>	PH	Y	179

## LIST OF TABLES

Table 1-1. Summary of DPBs and their occurrence in disinfected water	13
Table 2-1. Levels of free amino acids in natural water source	27
Table 3-1. Genotoxic and non-genoxic test chemicals used in assay validation	42
Table 4-1. List of the organic N-chloramine candidates and their precursors	77
Table 5-1. Cytotoxicity of organic N-chloramines	104
Table 5-2. Determination of cytotoxicity of the pre-chlorinated amines	104
Table 5-3. Genotoxicity of organic N-chloramines	107
Table 5-4. Determination of genotoxicity of the pre-chlorinated amines	108
Table 7-1. MW Fractions, DOC and predicted chlorine doses	146

### LIST OF FIGURES

Figure 1.1. Halogenated DBPs as proportionate to total organic halogen	15
Figure 2.1. Formation of organic N-chloramines in water and <i>in vivo</i> significance	33
Figure 3.1. Hemocytometer (Improved Neubauer) used for cell count.	45
Figure 3.2. Optical layout of the FACSCalibur™ flow cytometer	47
Figure 3.3. Summary of protocols used for <i>in vitro</i> genotoxicity assessment	51
Figure 3.4. Growth rate of WIL2-NS and L5178Y cell lines.	56
Figure 3.5 Gating criteria for flow cytometric measurement of micronuclei	58
Figure 3.6. Flow cytometry sorting of PI-stained nuclei (N) and micronuclei (MN)	60
Figure 3.7. Cytotoxicity and micronuclei formation responses of WIL2-NS	67
Figure 3.8. Cytotoxicity and micronuclei formation responses of L5178Y	68
Figure 3.9. Apoptosis measurement by the flow cytometric NucView Assay	70
Figure 3.10. Comparisons of apoptosis responses between WIL2-NS and L5178Y	71
Figure 4.1. Molecular structure of amino acids and amine precursors	78
Figure 4.2. Amines and rate of chlorination.	84
Figure 4.3. Production of organic N-chloramines (I)	86
Figure 4.4. Production of organic N-chloramines (II)	87
Figure 5.1. Experimental plan of chemical treatment in a 24-well tissue culture plate	96
Figure 5.2. Comparison of cytotoxicity of chlorine in serum-free treatment media	100
Figure 5.3. Comparative results between 24 hr and 3 hr exposure protocols	101
Figure 5.4. Cytotoxicity of organic N-chloramines.	103
Figure 5.5. Genotoxicity of organic N-chloramines	106
Figure 6.1. Formation of organic N-chloramine and predicted radical formation	115
Figure 6.2. Reaction scheme for the Glutathione assay	117
Figure 6.3. Representative total glutathione standard curve	. 123

Figure 6.4. Effect of ferrous sulphate (FeSO <sub>4</sub> ) on intracellular glutathione levels
Figure 6.5. Levels of intracellular glutathione after treatment with organic N-chloramines.126
Figure 6.6. Representative MDA standard curve
Figure 6.7. MDA induction in WIL2-NS cells following treatment with FeSO <sub>4</sub> 129
Figure 6.8. MDA levels in WIL2-NS cells following N-chloamine exposure
Figure 7.1. SEC-UV chromatogram of highly coloured surface water
Figure 7.2. Representative chlorine standard curve
Figure 7.3. Cytotoxicity of chlorinated MW fractions
Figure 7.4. Genotoxicity of chlorinated MW fractions
LIST OF APPENDICES
Appendix 1: Media, Buffers and Solutions
Appendix 2: Additional Results

#### **ABSTRACT**

Organic N-chloramines have long been recognised as disinfection by-products (DBPs) found in both chlorinated and chloraminated water, but have gained little attention from water authorities in the past. However, in recent years studies have shown that organic N-chloramines are molecules involved in inflammation and several chronic diseases including cancers. A recent study (Bull *et al.*, 2006) has suggested that organic N-chloramines can be potential health risks but due to a lack of available toxicological information toxicity studies of compounds in this group have been recommended as a priority in DBPs research.

The aim of this study was to investigate genotoxicity of individual organic N-chloramines utilising a mammalian cell-based genotoxicity assay to help determine which compound(s) should be subject to further *in vivo* studies. The flow cytometry-based micronucleus (FCMN) assay was optimised and validated for use as a rapid screening for genotoxicity of organic N-chloramine candidates. A number of assay validations were conducted on two mammalian cell lines (WIL2-NS and L5178Y) using model genotoxicants with various modes of action. Comparative studies on these two cell lines showed that WIL2-NS cells were suitable for the FCMN assay and therefore selected for use in all studies described in this thesis.

For the genotoxicity investigation of organic N-chloramines, 16 compounds were synthesised by chlorination of amine precursors. At least 3 concentrations (in µM range) were subjected to screening for genotoxicity using the validated FCMN assay and confirmed by microscopic counting of micronuclei. This study found that of the 16 compounds, 4 were genotoxic to WIL2-NS cells by both FCMN and microscopy based MN

assay. Oxidative stress was hypothesised as a possible genotoxic mechanism of these compounds and also was investigated in this study. Following exposure to the 4 genotoxic organic N-chloramines, it was found that although there was a small reduction of cellular glutathione the change in lipid peroxidation was not observed. This suggested that oxidative stress is unlikely to be a mechanism involved in genotoxicity of these organic N-chloramines.

The final part of this research demonstrated an application of using the optimised FCMN assay to identify genotoxic DBP precursors in Australian water. We collaborated with Curtin University, Western Australia on this aspect. Highly coloured surface water was collected, concentrated, and fractionated based on molecular weight (MW) of the organic contents by researchers at Curtin University. Eight MW fractions (pre- and post chlorination) were tested for genotoxicity using the FCMN assay. No genotoxicity was observed in all pre-chlorinated MW fractions while significant genotoxicity was seen in chlorinated products of several fractions of medium to high MW. This result indicated that these fractions contain materials that are precursors to genotoxic DBPs and may lead to future studies such as characterisation of the genotoxic DBP precursors for their removal prior to the disinfection process.

**DECLARATION** 

This work contains no material which has been accepted for the award of any other degree

or diploma in any university or other tertiary institution and, to the best of my knowledge

and belief, contains no material previously published or written by another person, except

where due reference has been made in the text.

I give consent to this copy of my thesis, when deposited in the University Library, being

made available for loan and photocopying, subject to the provisions of the Copyright Act

1968. The author acknowledges that copyright of published works contained within this

thesis (as listed on page IV) resides with the copyright holder(s) of those works.

I also give permission for the digital version of my thesis to be made available on the web,

via the University's digital research repository, the Library catalogue, the Australasian

Digital Theses Program (ADTP) and also through web search engines, unless permission

has been granted by the University to restrict access for a period of time.

Somprasong Laingam

Singed .....

Date .....

iii

#### **ACKNOWLEDGEMENTS**

I would like to thank my supervisors, Dr Andrew Humpage and Dr Suzanne Froscio from the Australian Water Quality Centre for their guidance and support throughout my candidature. Their knowledge and experience in the field of toxicology as well as their constructive criticisms have made the completion of this thesis possible. I would also like to thank my co-supervisor, Dr Ian Musgrave from the discipline of Pharmacology, who has always ensured that my candidature outside the university ran smoothly.

Thanks to all researchers and staff at the Australian Water Quality Centre for their help and support during my study. Special thanks go to Stella and Melody, who have helped with the smooth running of the cell culture laboratory and Rebecca for all her encouragement and sharing her PhD journey, particularly during late hours and weekend work. Similar thanks go to all researchers and staff in the Applied Chemistry Research and Water Treatment who shared their knowledge and frequently help me with the chlorination work.

A sincere thank-you to the CRC for Water and Quality Treatment and the Discipline of Pharmacology, Faculty of Health Science who provided my scholarship and the funding for this project. It has been an honour being a CRC PhD candidate. Finally, I would like to thank my family and friends for their encouragement and support during my study. You have had a big influence on the completion of this thesis for which I am truely thankful.

#### **PUBLICATIONS IN SUPPORT THIS THESIS**

Laingam, S., Froscio, S.M. and Humpage, A.R. (2008). Flow-cytometric Analysis of *in vitro* Micronucleus Formation: Comparative Studies with WIL2-NS Human Lymphoblastoid and L5178Y Mouse Lymphoma Cell Lines. *Mutatation Research*, 656 (1-2): 19-26.

Laingam, S., Froscio, S.M. and Humpage, A.R. (2009). Toxicity and Genotoxicity of Disinfection By-products, Organic N-chloramines. *In preparation*.

Laingam, S., Froscio, S.M. and Humpage, A.R. (2009). Use of the Flow Cytometry-based Micronucleus Assay to Determine Genotoxic DBP Precursors from Australian Water. *In preparation*.

#### ABBREVIATIONS AND SYMBOLS

ANOVA Analysis of variance

ATCC American type culture collection

ADWG Australian drinking water guidelines

AWQC Australian water quality centre

BA Bromoacetic acid

BaP Benzo[a]pyrene

BrO<sub>3</sub> Bromate

CHCl<sub>3</sub> Chloroform

CHO Chinese hamster ovary

CI Confident interval

Cl<sub>2</sub> Chlorine

ClO<sub>2</sub> Chlorine dioxide

ClO<sub>2</sub> Chlorite

DBP(s) Disinfection by-product(s)

DNA Deoxyribonucleic acid

DOC Dissolved organic carbon

DON Dissolved organic nitrogen

DPD N,N-diethyl-p-phenyl diamine

dsDNA Double stranded deoxyribonucleic acid

DTNB 5, 5'- dithiolbis-2-nitrobenzoic acid

EC30 Effective concentration at 30% of the untreated control

EC50 Effective concentration at 50% of the untreated control

ECL Enterochromaffin-like

EDTA Ethylenediaminetetraacetic acid

EPA Environmental protection agency

ETOPO Etoposide

FACS Fluorescence-activated cell sorting

FAS Ferrous ammonium sulphate

FBS Foetal bovine serum

FCMN Flow cytometry based micronucleus

FeSO<sub>4</sub> Ferrous sulphate

FSC Forward scatter

GSH Glutathione (Reduced form)

GSSG Glutathione (Oxidised form)

H<sub>2</sub>O<sub>2</sub> Hydrogenperoxide

HAAs Haloacetic acids

HANs Haloacetonitriles

HBSS Hank balanced salt solution

HCl Hydrochloric acid

HEPES 4-(2-hydroxyethyl)-1-piperazineethanesulfonic acid

HOCl Hypochlorous acid

HPLC High pressure liquid chromatography

HPSEC High performance size exclusion chromatography

IARC International agency for research on cancer

LOAELs Lowest observed adverse effect levels

MDA Malondialdehyde

MMC Mitomycin C

MMS Methyl methanesulfonate

MN Micronucleus

MW Molecular weight

MX 3-chloro-4(dichloromethyl)-5-hydroxy-2(5H) furanone

N Nucleus

NCl<sub>3</sub> Trichloramine

NCP N-chloropiperidine

NDMA N-nitrosodimethylamine

NH<sub>2</sub>Cl Monochloramine

NH<sub>3</sub> Ammonia

NHCl<sub>2</sub> Dichloramine

NOM Natural organic matter

O<sub>3</sub> Ozone

OCl Hypochlorite ion

OECD Organization for economic co-operation and development

OR Odds ratio

PBS Phosphate buffered saline

PI Propidium iodide

QSTR Quantitative structure toxicity relationship

RO Reverse osmosis

RPMI Roswell park memorial institute medium

r Spearman's coefficient

SD Standard deviation

SDS Sodium dodecyl sulphate

SEM Standard error of the mean

SSC Side scatter

SUCRO Sucrose

TB Trypan blue

TBARS Thiobarbituric acid reactive substances

TDN Total dissolved nitrogen

THMs Trihalomethanes

US The United States

VINB Vinblastin

WHO World health organization