

GM-CSF protection of CML CD34⁺ cells from the inhibitory effect of imatinib

Pongtep Viboonjuntra

Melissa White Leukaemia Research Laboratory
Division of Haematology
Institute of Medical &Veterinary Science
And
Hanson Institute

&

Department of Medicine
The University of Adelaide
South Australia

A thesis submitted to the University of Adelaide in candidature of the degree of Doctor of Philosophy February 2006

ABSTRACT

Imatinib mesylate has become the front line therapy for CML. It inhibits the ABL tyrosine kinase activity of the causative oncogenic fusion protein BCR/ABL. The efficacy of imatinib therapy has been impressive, with a majority of patients achieving complete cytogenetic responses. However, to date imatinib therapy has been unable to completely eradicate the leukaemic clone in the majority of cases. This suggests that leukaemic stem cells may not be totally dependent on the activity of BCR/ABL, or that there may be other factors supporting the survival of leukaemic cells. Cytokines of the IL-3 family are involved in the survival and growth of myeloid haemopoietic cells. A number of the signals generated are in common with the pathways used by BCR/ABL and previous published studies showed that the IL-3 family member GM-CSF was found in elevated levels in CML patient serum. Therefore, the central aim was to test whether GM-CSF could modulate the response of CML progenitors to imatinib. In addition, a further aim was to examine CML cell production of GM-CSF. When CML CD34+ cells were exposed to imatinib, spontaneous BCR/ABL driven cell division was strongly inhibited, and cells became apoptotic. However, the addition of GM-CSF reversed this effect, returning cell division and survival back to spontaneous levels. GM-CSF specificity was confirmed using the antagonist E21R. The cell division and survival of leukaemic CD34+ cells in cultures of total CML mononuclear cells were less sensitive to imatinib inhibition. However, inclusion of E21R in such cultures resulted in a similar pattern to CD34+ CML cells cultured alone. This suggested that non-CD34+ cells may be a source of GM-CSF. This was confirmed by sorting CD34+ and non-CD34+ CML cells and examining GM-CSF production, which was markedly more apparent in the non-CD34+ compartment. Finally, the effects of GM-CSF on the survival and proliferation pathways induced by BCR/ABL were examined, showing it may exert its effects through p-Erk and p-Akt mediated mechanisms. Our data suggest that GM-CSF can signal via pathways which are normally suppressed due to alternative activation of major substrates by BCR/ABL, as well as others which are not activated by BCR/ABL, leading to cell survival when kinase activity is blocked by imatinib.

TABLE OF CONTENTS

DECL	ARATION	I
ACKN	NOWLEDGEMENTS	.II
ABST	RACT	VI
ABBR	EVIATIONSV	'II
PRES	ENTATIONS AND PUBLICATIONS	II
СНАР	TER 1: GENERAL INTRODUCTION	
1.1. P	PREFACE	1
1.2. H	HISTORY OF CHRONIC MYELOID LEUKEMIA	1
1.3	EUKAE MIA 3.1. Epidemiology 3.2. History of CML treatment 1.3.2.1. Busulfan and Hydroxyurea 1.3.2.2. Interferon-α 1.3.2.3. Chemotherapy and bone marrow transplantation 1.3.2.4. The tyrosine kinase inhibitor imatinib	2334
1.4. N	Molecular pathophysiology of CML	6
1.4 1.4	4.1. Normal p145 protein 4.2. Normal BCR protein 4.3. More studies on the function of normal BCR and ABL 4.4. The BCR/ABL fusion protein or p210 ^{BCR/ABL} protein	.10
1.5.	Cellular signaling pathways relating to the BCR/ABL protein	15
1.: 1.: 1.:	5.1. Ras pathways 5.2. PI3-K pathway 5.3. C-Cbl and Crkl pathways 5.4. Myc pathway 5.5. Jak/STAT	17 19. 20.
	Haemopoietic cytokines of the IL-3 family involved in signaling through BCR/ABL	21
1.0 1.0	6.1. IL-3 receptor super family	.22

	disease and neoplasia	25
1.7.	Proposed studies in this thesis	27
	1.7.1. Cell growth driven by BCR/ABL is cytokine dependent	27
	1.7.2. Imatinib therapy of CML is not curative	
	1.7.3. GM-CSF may maintain survival of CML progenitors and help	CML
	progenitors to escape eradication by imatinib	
1.8.		
1 0	Summary	22
1.9.	Summary	32
CH.	APTER 2: MATERIALS AND METHODS	
2.1.	Suppliers of commonly used reagents	33
2.2.	Buffers and Solutions used un cell culture	36
2.3.	Cell Culture Techniques	
	2.3.1. Maintenance of cell lines	
	2.3.1.1. Cryopreservation of cell lines	
	2.3.1.2. Thawing cell lines	
	2.3.1.3. Performing the cell culture	
	2.3.2. Maintenance of primary CML cells	
	2.3.2.1. Ficolling technique for primary CML cells	
	2.3.2.2. Cryopreservation of primary CML cells	
	2.3.2.3. Thawing the primary CML cells	
	2.3.2.4. Performing CML cell culture	47
2.4.	General cell washing	47
2.5.	. General cell number and viability assessment	48
2.6.	. Immunofluorescence and flow cytometry	48
2.7.	. Haematopoietic colony assays	50
2.8.	. Protein analysis	53
	2.8.1. Molecular Biology reagents	
	2.8.2. Method of Western immunoblot	
2.9.	. Statistical methods	59
CH	IAPTER 3: THE RESPONSE OF CML CD34 ⁺ CELLS TO GM-CSF	
3.1.	. Introduction	60
3.2.	. Aims of the study	63
3.3.	. Special methods	63
	3.3.1. Dye dilution technique (CFSE)	
	3.3.1.1. Non-division related decrease in CFSE intensity	
	3.3.1.2. Calculation of proliferation index	
3.4.	. Results	

3.4 3.4	4.1. 4.2. 4.3. 4.4.	Titration of GM-CSF study	67 .68
3.5.	Discussi	ion	
		MODULATION OF IMATINIB EFFECT ON CML CD34 ⁺ CELLS	1
	M-CSF		7 2
4.1.		uction	
4.2.		f the study	
4.3.	•	methods	
4.3	.1.	Colorimetric MTS assays	.76
4.4.	Statisti	cal methods	.77
4.5.		S	
	5.2.	A time-course experiment to evaluate cell proliferation by CFSE	
4.:	5.3.	cells with imatinib	
	5.4. 5.5.	cells	80
	5.6.	GM-CSF and E21R	
4	5.8.	could be replaced by added GM-CSF in the presence of imatinib	
4.6.	Discu	ıssion	84
СНАН	PTER 5:	EFFECT OF BYSTANDER CELLS ON CML CD34 ⁺ CELL	
BEHA	VIOUF	₹	
5.1.	Introd	uction	88
5.2.		of the study	
5.	Specia 3.1. 3.2. 3.3.	I methods MTS assay to compare between the survival of CML CD34 ⁺ cells and mixed mononuclear cells in the conditioned culture Evaluation of the bystander effect	89 90 92
	5.3.3	.1. Preparation of ELISpot plate	.92

	5.3.3. 5.3.3.	1	
5.4.		5	
	5.4.1.	Comparison of cell survival between CML CD34 ⁺ cells in CML CD34	4 ⁺
		cell only and in mixed mononuclear culture	
		CML CD34 ⁺ cells underwent more proliferation in mixed mononuclea	
		culture	
		of CML CD34 ⁺ cells	
		The effect of bystander helps to maintain cell survival rather than cell	
		cell proliferation	96
	5.4.5.	GM-CSF is predominantly produced by non-CD34 ⁺ CML cells	97
5.5.	Discuss	sion	97
		MECHANISM OF PROTECTION FROM IMATINIB BY GM-C	
6.1.		overview from the previous Chapter	
		Role of normal c-ABL	
		Mechanism of c-ABL shuttling between nucleus and cytoplasm	
		Regulation of cell apoptosis	
		Role of Bim	
6.2.	Aims o	f the study	106
6.3.	Special	l methods	107
		MTS assay to evaluate bio-kinetic activity of cell lines	
		Western immunoblot method.	
		2.1. Western immunoblot with total cell lysate	
	6.3.2	2.2. Western immunoblot with immunoprecipitated sample	108
6.4.	Results		109
		Wt-BCR-ABL/ CTLEN cells in culture with titrated GM-CSF by MTS assay	100
		MTS assay to evaluate different cell lines.	
		MTS assay to evaluate the wt-BCR-ABL / CTLEN with the titrated	110
		imatinib	110
	6.4.4.	Mechanism of GM-CSF to protect cells from imatinib	
	6.4.4		
	6.4.4	1	
	6.4.4		
	6.4.4 6.4.4	F -	
	6.4.4		
	6.4.4	1	114
	3.11	Culture	115
6.5.	Disaus	sion	
v.J.	Discus	51UII	110

7.2.	Aims of the	study	12
7.3.	Special meth	10ds	12
		ination of active caspase expression by FLICA method	
7.4.	Results		12
	7.4.1. Study one p	y of the proliferation of CML CD34 ⁺ cells and bystander effect patient	in 120
		parison of p-Akt in both primary CML CD34 ⁺ cells and cell line BCR-ABL / CTLEN	
	7.4.3. Evalu	uation of p-Erk	12
		gulation of the apoptotic regulatory protein Bim by imatinib	
		total caspase activation in CML CD34 ⁺ cells in CML CD34 ⁺ on are and in mixed mononuclear culture	•
	7.4.6. Apop	ptotic index	13
7.5.	Discussion		1