

**AN INVESTIGATION INTO THE DEVELOPMENT  
AND PATTERNING OF DORSAL LONGITUDINAL  
ASCENDING INTERNEURONS IN *DANIO RERIO***

**Simon J. Wells, B.Sc. (Hons)**



**THE UNIVERSITY**  

---

***of* ADELAIDE**

**Discipline of Genetics**  
**School of Molecular and Biomedical Science**  
**University of Adelaide**  
**AUSTRALIA**

**February 2011**

## **Acknowledgements**

I would like to express my thanks to the following people

My primary supervisor requires special mention. Thanks to Michael for his patience and his persistently positive encouragement. You had belief when often I had little.

Lardelli lab members past and present for support and comradeship over the years.

The SMBS and especially the Genetics Department have provided a wonderful environment to develop as a scientist and educator. Thank you to those who I have had contact with throughout my Ph.D.

Velta for your support in numerous ways.

Clare for the many valuable discussions and constant gentle motivation.

My family for understanding and trying to understand.

Authors and journals for reprint permissions.

Lastly, thanks to all the fish. The silent partner in all of this science; at various times a source of joy or consternation, but always a source and for that I am thankful.

Without you all I would not have made it

Dedicated to  
Terry and Casey

You left before your time was up

# Contents

<b>Abstract</b> .....	<b>1</b>
<b>Declaration</b> .....	<b>3</b>
<b>Introduction</b> .....	<b>4</b>
1. Zebrafish as a model system .....	5
2. The <i>spadetail</i> mutation.....	6
3. The <i>tbx16</i> gene is mutated in <i>spadetail</i> embryos .....	7
4. The T-box gene family .....	8
5. T-box genes are important during development.....	9
6. Expression of <i>tbx16</i> in the central nervous system .....	11
7. How to build a spinal cord .....	12
7a. Gross morphogenetic movements as the tail extends .....	12
<a href="#">Figure 1. Schema of vertebrate spinal cord structure</a> .....	13
7b. Patterning the dorsoventral axis .....	14
7c. Patterning the rostrocaudal axis .....	14
8. Zebrafish spinal cord neurons .....	16
<a href="#">Figure 2. Identified interneurons in the embryonic zebrafish spinal cord</a> .....	18
9. DoLA neurons .....	19
10. Spatial analysis of geometric distributions .....	20
11. Generating transgenic animals for investigation of DoLA interneurons .....	21
<b>Summary of papers and continuity</b> .....	<b>23</b>
<b>Paper I</b> .....	<b>26</b>
<b>Paper II</b> .....	<b>42</b>
Supplementary material.....	57
<b>Paper III</b> .....	<b>68</b>
Supplementary material.....	84
<b>Paper IV</b> .....	<b>86</b>
<b>Discussion</b> .....	<b>106</b>
1. Mechanism 1.....	107
2. Mechanism 2.....	109
3. Mechanism 3.....	110
4. A model for the mechanism underlying DoLA cell contralateral correlation .....	111
<a href="#">Figure 3. A model for the mechanism underlying DoLA cell contralateral correlation</a> .....	112
5. Future directions.....	113
6. The creation of a new <i>tbx16:GFP</i> transgenic line .....	114
<b>Bibliography</b> .....	<b>116</b>

## Abstract

The dorsal longitudinal ascending (DoLA) interneurons are an uncommon, seemingly irregularly distributed interneuron type of the developing embryonic zebrafish spinal cord. For reasons not yet understood DoLA interneurons express *tbx16*, a T-box transcription factor originally recognised for its important role in mesodermal development. This is the only cell type expressing *tbx16* in the developing spinal cord, making DoLA neurons one of the few neuronal types that can be identified by expression of a unique molecular marker.

Throughout the natural world regularity in pattern formation is frequent; mechanisms that direct the production of regular patterns have been studied and many are well understood. The creation of irregular "patterns", especially in embryo development has been subjected to far less analysis. This is largely because studies in developmental biology frequently involve methods that disrupt regular patterning while the disruption of an irregular pattern is likely to result in similarly irregular pattern. The DoLA interneurons with their unique genetic marker offer a rare opportunity to investigate the mechanisms behind irregular patterns in development. This is of particular importance in the development of the spinal cord, as most of the known vertebrate spinal interneurons appear to have irregular distributions.

The main focus of the research presented in this thesis has been to try to understand how the distribution pattern of DoLA interneurons is generated. This knowledge may then be extended to other spinal neurons and possibly to other irregular developmental patterns.

In the work described in this thesis the distribution of DoLA interneurons has been extensively examined statistically. It was found that there is an underlying cryptic organisation to their peculiar distribution. This led to the surprising discovery that these neurons migrate rostrally a significant distance along the spinal cord. These neurons were also found in larval zebrafish at much older times than has previously been described, suggesting that they may play a role in post-embryonic stages.

Notch signalling appears to have an influence on DoLA interneuron distribution since perturbing Presenilin (Psen) function affects the number of these cells. Interestingly, DoLA cell number is not affected when Psen1 function is inhibited but increases when Psen2 function is inhibited. Furthermore the wild type level of DoLA interneuron number can be partially rescued by inhibiting Psen1 function in combination with inhibition of Psen2 function.

The creation of transgenic zebrafish lines where GFP is transcribed from *tbx16* promoter sequence is described. These animals were produced to attempt to discover more about the patterning of DoLA interneurons and the function of *tbx16* during development. Serendipitously, one of these transgenic lines expresses GFP in the commissural primary ascending (CoPA) interneurons. This led to the discovery that the CoPA interneurons are marked by *mafba/valentino*, revealing a new unique spinal neuron molecular marker.

## Declaration

This work contains no material which has been accepted for the award of any other degree or diploma in any university or other tertiary institution and, to the best of my knowledge and belief, contains no material previously published or written by another person, except where due reference has been made in the text.

I give consent to this copy of my thesis when deposited in the University Library, being made available for loan and photocopying, subject to the provisions of the Copyright Act 1968.

The author acknowledges that copyright of published works contained within this thesis (as listed below\*) resides with the copyright holder(s) of those works.

I also give permission for the digital version of my thesis to be made available on the web, via the University's digital research repository, the Library catalogue, the Australasian Digital Theses Program (ADTP) and also through web search engines, unless permission has been granted by the University to restrict access for a period of time.

\* Publications (copyright holder/s):

The identity and distribution of neural cells expressing the mesodermal determinant *spadetail* (Authors: Tamme, R., Wells, S., Conran, J.G., Lardelli, M.)

Independent and cooperative action of Psen2 with Psen1 in zebrafish embryos (Elsevier B.V., Amsterdam)

Cryptic organisation within an apparently irregular rostrocaudal distribution of interneurons in the embryonic zebrafish spinal cord (Elsevier B.V., Amsterdam)

Transgenic zebrafish recapitulating *tbx16* gene early developmental expression (Submitted manuscript, under review)

Signed

.....Date.....