

# **Genomic approach to understanding variation in bovine fat colour**

**Presented by**

**Rugang Tian**

A thesis submitted to the University of Adelaide in fulfilment  
of the requirement of the degree of  
PhD of Science



**THE UNIVERSITY**  
*of* **ADELAIDE**

**The University of Adelaide**  
**School of Animal and Veterinary Sciences**  
**Discipline of Animal Science**  
**September 2011**

# Table of Contents

Index of Figures .....	vi
Index of Tables.....	ix
Index of Appendices .....	xi
Declaration .....	xiii
Dedications .....	xiv
Acknowledgements.....	xv
Abstract .....	xviii
<b>Chapter 1 Literature Review .....</b>	<b>1</b>
1.1 Introduction.....	2
1.2 Fat colour and market specification .....	2
1.3 Factors affecting fat colour .....	3
1.3.1 Non-genetic factors .....	3
1.3.1.1 Diet .....	3
1.3.1.2 Season.....	6
1.3.1.3 Management strategies .....	6
1.3.1.4 Age.....	8
1.3.1.5 Sex .....	8
1.3.1.6 Growth rate .....	9
1.3.2 Genetic factors .....	10
1.4 Carotenoids and fat colour .....	13
1.4.1 Carotenoid function.....	13
1.4.2 Ruminant digestion .....	15
1.4.3 Carotenoid absorption .....	16
1.4.4 Carotenoid transportation.....	18
1.4.5 Carotenoid deposition .....	19
1.4.6 Enzymes in metabolism of $\beta$ -carotene .....	20
1.4.6.1 $\beta$ -carotene cleavage enzymes .....	20
1.4.6.2 Other enzymes .....	27
1.5 Summary.....	30
<b>Chapter 2 General methods and materials.....</b>	<b>32</b>

2.1	Animal resources.....	33
2.2	Phenotypic data collection .....	35
2.3	DNA preparation .....	37
2.4	Candidate gene selection.....	38
2.5	Single nucleotide polymorphisms experiments .....	38
2.5.1	Primer design and selection .....	38
2.5.2	Optimization of PCR conditions .....	39
2.5.3	Automated cycle sequencing of PCR products .....	41
2.5.4	SNP discovery .....	42
2.5.5	Genotyping.....	43
2.5.5.1	Genotyping by PCR-RFLP .....	43
2.5.5.2	Genotyping by High Resolution Melting (HRM).....	44
2.6	Real-time PCR .....	45
2.6.1	Tissue collection.....	45
2.6.2	RNA isolation.....	45
2.6.3	cDNA production .....	46
2.6.4	Real-time PCR conditions.....	46
2.7	Statistical analysis .....	47
2.7.1	Animal Group 1: .....	47
2.7.2	Groups 2 to 6 :.....	51
2.7.3	Group 7: .....	52
<b>Chapter 3</b>	<b>Candidate gene selection .....</b>	<b>54</b>
3.1	Introduction.....	55
3.2	Methods.....	59
3.3	Results.....	60
3.3.1	Candidate gene selection.....	60
3.3.2	SNP identification and selection for association studies.....	60
3.3.3	Genotype and allele frequencies .....	64
3.4	Discussion: .....	67
<b>Chapter 4</b>	<b>Association between candidate gene SNPs and fat colour traits .....</b>	<b>73</b>
4.1	Introduction.....	74
4.2	Materials and methods .....	74
4.3	Results.....	76
4.3.1	Cohort, breed, and sire effects.....	76
4.3.2	Individual SNP genotype effects .....	79

4.3.3	SNP additive and dominance effects .....	83
4.3.4	Non-candidate gene SNP associations .....	85
4.3.5	<i>BCO2</i> W80X effects on significance of other SNPs.....	87
4.3.6	QTL analysis .....	89
4.3.6.1	<i>BCO2</i> W80X effects on QTL .....	91
4.3.6.2	<i>ALDH8A1</i> SNP16 effects on QTL.....	93
4.3.6.3	<i>PPARGC1A</i> SNP12 effects on QTL.....	95
4.3.6.4	<i>RDHE2</i> SNP2 effects on QTL.....	98
4.4	Discussion .....	101
4.4.1	Breed effects.....	101
4.4.2	Multiple and individual SNP testing .....	104
4.4.3	Non-coding polymorphisms.....	104
4.4.4	Functional significance of candidate genes.....	105
4.4.4.1	<i>ALDH8A1</i> .....	105
4.4.4.2	<i>APOM</i> .....	106
4.4.4.3	<i>RARA</i> .....	107
4.4.4.4	<i>RDHE2</i> .....	107
4.4.4.5	<i>PPARGC1A</i> .....	108
4.4.4.6	<i>SCARB1</i> .....	109
4.4.4.7	<i>BCMO1</i> .....	110
4.4.4.8	<i>BCO2</i> .....	111
4.4.4.9	Other non-candidate genes .....	112
4.4.5	Additional SNP effects on QTL .....	113
<b>Chapter 5</b>	<b>Interactions between candidate gene SNPs .....</b>	<b>115</b>
5.1	Introduction.....	116
5.2	Materials and methods .....	116
5.3	Results.....	117
5.3.1	Interaction effects between SNPs within genes .....	117
5.3.2	Haplotype associations.....	119
5.3.3	Epistatic interactions of major SNPs.....	122
5.4	Discussion .....	125
5.4.1	Haplotype effects.....	125
5.4.2	Epistatic effects .....	126
<b>Chapter 6</b>	<b><i>BCO2</i> and <i>RDHE2</i> gene validation studies.....</b>	<b>128</b>
6.1	Introduction.....	129

6.2	Materials and methods .....	129
6.2.1	Animal resources.....	129
6.2.2	Statistical analysis .....	130
6.3	Results.....	130
6.3.1	<i>BCO2</i> validation.....	130
6.3.1.1	Australian Jersey dams .....	130
6.3.1.2	Validation data from New Zealand .....	131
6.3.2	<i>RDHE2</i> validation .....	133
6.3.3	Interaction effects between <i>RDHE2</i> and <i>BCO2</i> .....	133
6.4	Discussion .....	135
6.4.1	<i>BCO2</i> effects .....	135
6.4.2	<i>RDHE2</i> effects.....	136
<b>Chapter 7</b>	<b>Gene expression studies.....</b>	<b>139</b>
7.1	Introduction.....	140
7.2	Materials and methods .....	141
7.2.1	Animal resources and methods .....	141
7.2.2	Statistical analysis .....	141
7.3	Results.....	142
7.3.1	RNA extraction.....	142
7.3.2	Optimization of PCR.....	142
7.3.3	Evaluation of reference genes .....	143
7.3.4	Gene expression analysis .....	144
7.3.4.1	Gene expression and fat colour.....	144
7.3.4.2	Genotype effects on gene expression.....	146
7.3.4.3	Gene expression effects on $\beta$ -carotene concentration.....	149
7.3.4.4	Relationships between relative gene expression levels .....	149
7.4	Discussion .....	151
7.4.1	<i>BCMO1</i> gene expression analysis .....	151
7.4.2	<i>BCO2</i> gene expression analysis .....	152
7.4.3	<i>RDHE2</i> gene expression analysis.....	157
7.5	Conclusion .....	158
<b>Chapter 8</b>	<b>General Discussion.....</b>	<b>159</b>
8.1	Introduction.....	160
8.2	SNP association studies.....	160
8.3	Non-synonymous SNP effects .....	163

8.4	Candidate pathways .....	168
8.4.1	Vitamin A synthesis via BCMO1 pathway.....	170
8.4.1.1	BCMO1 direct effects .....	170
8.4.1.2	Regulation of BCMO1 by transcription factors.....	171
8.4.1.3	Regulation of BCMO1 by negative feedback mechanisms .....	173
8.4.2	Vitamin A synthesis via BCO2 pathway .....	176
8.4.2.1	Asymmetric cleavage of $\beta$ -carotene .....	176
8.4.2.2	Asymmetric cleavage of lutein .....	177
8.4.3	Absorption and transportation of carotenoids .....	178
8.4.4	Dilution of carotenoids in the fat .....	182
8.5	Epistasis .....	187
8.6	Future studies .....	189
8.7	Conclusion .....	191
<b>Appendices</b>	.....	<b>195</b>
<b>References</b>	.....	<b>225</b>

## Index of Figures

Figure 1-1 $\beta$ -carotene and lutein content of forages, cereals and root crops used in cattle diets .....	5
Figure 1-2 Three forms of vitamin A .....	14
Figure 1-3 Schematic overview of $\beta$ -carotene metabolism.....	22
Figure 2-1 Backcross design of the Davies Gene Mapping Herd.....	34
Figure 2-2 AUS-MEAT standard colour chips .....	37
Figure 4-2 Effects of cohort, breed and sire on biopsy sample $\beta$ -carotene concentration .....	78
Figure 4-1 Effects of cohort, breed and sire on carcass samples fat colour score..	78
Figure 4-3 Effects of cohort, breed and sire on biopsy samples fat colour score ..	78
Figure 4-4 <i>BCO2</i> W80X effects on BTA 15 QTL across families.....	92
Figure 4-5 <i>BCO2</i> W80X effects on BTA 15 QTL for sire 398 family .....	92
Figure 4-6 <i>ALDH8A1</i> SNP16 effect on BTA 9 QTL across families.....	94
Figure 4-7 <i>ALDH8A1</i> SNP16 effect on BTA 9 QTL for sire 368 family .....	94
Figure 4-8 BTA 6 QTL across families without <i>PPARGCIA</i> SNP12 in the model	97
Figure 4-9 BTA 6 QTL across families with <i>PPARGCIA</i> SNP12 in the model ....	97
Figure 4-10 <i>PPARGCIA</i> SNP12 effect on BTA 6 QTL in sire 361 family.....	97
Figure 4-11 <i>RDHE2</i> SNP2 effect on BTA 14 QTL across families .....	99
Figure 4-12 <i>RDHE2</i> SNP2 effect on BTA 14 QTL in sire 361 family.....	99
Figure 4-13 <i>RDHE2</i> SNP3 effect on BTA 14 QTL across families.....	100
Figure 4-14 <i>RDHE2</i> SNP3 effect on BTA 14 QTL in sire 361 family .....	100
Figure 5-1 Interaction between <i>PPARGCIA</i> SNP 12 and <i>BCO2</i> W80X on $\beta$ -carotene concentration .....	124
Figure 5-2 Interaction between <i>PPARGCIA</i> SNP 12 and <i>SCARB</i> SNP1 on biopsy fat colour .....	124
Figure 5-3 Interaction between <i>PPARGCIA</i> SNP 12 and <i>RDHE2</i> SNP3 on carcass fat colour .....	124
Figure 7-1 Average expression stability values (M) of the reference genes with stepwise exclusion of the least stable expressed reference genes ....	144
Figure 7-2 <i>BCMO1</i> mRNA expression in the two fat colour groups .....	145
Figure 7-3 <i>BCO2</i> mRNA expression in the two fat colour groups .....	145

Figure 7-4 <i>RDHE2</i> mRNA expression in the two fat colour groups.....	145
Figure 7-5 Relative normalised <i>BCMO1</i> gene mRNA levels for different gene genotypes.....	147
Figure 7-6 Relative normalised <i>BCO2</i> gene mRNA levels for different gene genotypes.....	147
Figure 7-7 Relative normalised <i>RDHE2</i> gene mRNA levels for different gene genotypes.....	148
Figure 7-8 Correlation between <i>BCMO1</i> mRNA expression and <i>BCO2</i> mRNA expression.....	150
Figure 7-9 Correlation between <i>BCMO1</i> mRNA expression and <i>RDHE2</i> mRNA expression .....	150
Figure 7-10 Correlation between <i>BCO2</i> mRNA expression and <i>RDHE2</i> mRNA expression .....	150
Figure 7-11 Regression of <i>BCO2</i> mRNA level and beta carotene concentration	155
Figure 7-12 Regression of <i>BCO2</i> mRNA level and log beta carotene concentration .....	155
Figure 8-1 ALDH8A1 protein tertiary structure as predicted by the (PS) <sup>2</sup> program .....	165
Figure 8-2 ALDH8A1 protein secondary structure as predicted by the (PS) <sup>2</sup> program .....	165
Figure 8-3 PPARGC1A protein tertiary structure as predicted by the (PS) <sup>2</sup> program .....	166
Figure 8-4 PPARGC1A protein secondary structure as predicted by the (PS) <sup>2</sup> program .....	166
Figure 8-5 RDHE2 protein tertiary structure as predicted by the (PS) <sup>2</sup> program.	167
Figure 8-6 RDHE2 protein secondary structure as predicted by the (PS) <sup>2</sup> program .....	167
Figure 8-7 Three forms of vitamin A .....	168
Figure 8-8 Retinoic acid formation from $\beta$ -carotene and retinol .....	169
Figure 8-9 Putative transcription factor binding sites in the promoter region of <i>BCMO1</i> .....	173
Figure 8-10 Crosstalk between RAR and ISX signalling controls $\beta$ -carotene absorption.....	175



Figure 8-11 Correlation between beta-carotene concentration and fat depth.....	184
Figure 8-12 Correlation between biopsy fat colour and fat depth.....	184
Figure 8-13 Correlation between carcass fat colour and fat depth.....	184
Figure 8-14 Interactions between genes .....	188

## Index of Tables

Table 2-1 PCR reagent concentrations for <i>AmpliTaq Gold</i> DNA polymerase .....	39
Table 2-2 PCR reagent concentrations for <i>Kapa</i> DNA polymerase .....	40
Table 3-1 QTL detected by interval mapping for fat colour traits .....	57
Table 3-2 QTL detected by multiple-trait multiple marker for fat colour traits .....	58
Table 3-3 QTL on BTA and orthologous regions in human genomes and putative candidate genes .....	62
Table 3-4 DNA variants of the candidate genes .....	63
Table 3-5 Genotyped DNA variants .....	63
Table 3-6 Genotypic and allelic frequencies of SNPs .....	65
Table 4-1 Least squares means with standard errors for cohort, breed, and sire .....	77
Table 4-2 Association of candidate gene SNPs with fat colour traits .....	80
Table 4-3 SNP size of effect on fat colour traits as percentage of total SNP variation (%) .....	82
Table 4-4 Additive and dominance effects for significant trait association .....	84
Table 4-5 Additional gene SNP variance as percentage of total SNP variance (%) .....	86
Table 4-6 Comparison of individual SNP significance P values when <i>BCO2</i> W80X was excluded or included in the model for fat colour traits in Group 1 animals .....	88
Table 4-7 Major SNP effects on QTL across sire families .....	90
Table 4-8 Major SNP effects on QTL for individual sire families .....	90
Table 4-9 SNP allele frequency in XL and XJ backcross .....	103
Table 5-1 Interaction effects between SNPs within genes .....	118
Table 5-2 Diplotype association with significance and trait means .....	120
Table 5-3 Size of interaction effects between SNPs as percentage of total SNP variation (%) .....	123
Table 6-1 Means (SE) for year, <i>BCO2</i> W80X genotype, additive and dominant effects in Australian Jersey dams .....	131
Table 6-2 Substitution effects of <i>BCO2</i> W80X alleles for fat colour traits in the New Zealand cattle .....	132
Table 6-3 Effects of <i>RDHE2</i> SNPs on fat colour traits .....	134
Table 6-4 <i>RDHE2</i> allele frequencies in experimental cattle populations .....	137
Table 7-1 Optimal annealing temperature for primer pairs .....	142

Table 7-2 M values of reference genes.....	143
Table 7-3 Relative normalised gene mRNA expression <sup>a</sup> .....	144
Table 7-4 Relative normalised gene mRNA levels among <i>RDHE2</i> SNP2 genotypes and SNP <i>BCO2</i> W80X genotypes .....	146
Table 7-5 Regression co-efficients (slope of linear regression analysis) for the relationship between $\beta$ -carotene concentration and relative gene expression level.....	149
Table 8-1 Proportion of the sum of squares accounted for all SNP additive effects.	162

## Index of Appendices

Appendix 1: Protocols .....	196
Appendix 1.1: Extraction and quantification of carotenoids .....	196
Appendix 1.2: DNA Purification .....	197
Appendix 1.3: Purification of PCR products for sequencing .....	198
Appendix 1.4: Ambion TURBO DNA- <i>free</i> <sup>TM</sup> DNase Treatment .....	198
Appendix 1.5: TRI Reagent <sup>®</sup> DNA Isolation .....	199
Appendix 2: Chemicals and solutions .....	200
Appendix2.1: TE Buffer .....	200
Appendix 2.2: TAE buffer .....	200
Appendix 2.3: Formamide Loading Buffer .....	200
Appendix 2.4: 1M Tris-HCl.....	200
Appendix 3: PCR conditions .....	201
Appendix 3.1: PCR conditions for the candidate genes .....	202
Appendix 3.2: PCR conditions for the genotyped SNPs .....	206
Appendix 3.3: Real-time PCR conditions .....	208
Appendix 4: Identified DNA variants.....	209
Appendix 5: Candidate gene haplotypes .....	212
Appendix 6: Effects of candidate gene SNPs on fat colour traits with covariates ..	213
Appendix 7 QTL.....	214
Appendix 7.1: Most likely position and F-statistic values of putative QTL detected by individual family analysis (Sire 361 family) .....	214
Appendix 7.2: Most likely position and F-statistic values of putative QTL detected by individual family analysis (Sire 368 family) .....	216

Appendix 7.3: Most likely position and F-statistic values of putative QTL detected by individual family analysis (Sire 398 family) .....	218
Appendix 7.4: Position and F-statistic values of putative QTL detected by across family analysis .....	220
Appendix 8: Sequence alignment .....	222
Appendix 8.1: <i>RDHE2</i> and <i>RDHE2</i> similar genes from bovine and human .....	222
Appendix 8.2: Pairwise alignments score for bovine, human <i>RDHE2</i> and <i>RDHE2</i> <i>similar</i> genes .....	222
Appendix 8.3: Bovine, human <i>RDHE2</i> and <i>RDHE2 similar</i> genes amino acid sequence alignments .....	223
Appendix 9: RNA quality .....	224

## **Declaration**

I declare that this thesis is a record of original work and contains no material that has been accepted for the award of any other degree or diploma in any university or other tertiary institution to Rugang Tian except where as stated in chapter 3, some sequencing (*BCO2* gene) was completed as part of a research project for a Masters by Coursework degree at the University of Adelaide in 2006. To the best of my knowledge and belief, this thesis contains no material previously published or written by any other person, except where due reference is made in the text.

I give consent to this copy of my thesis, when deposited in the University Library, being made available for loan and photocopying, subject to the provisions of the Copyright Act 1968.

I also give permission for the digital version of my thesis to be made available on the web, via the University's digital research repository, the Library catalogue, the Australasian Digital Theses Program (ADTP) and also through web search engines, unless permission has been granted by the University to restrict access for a period of time.

Rugang Tian

September, 2011

## **Dedications**

I dedicate this work to my parents and my wife, Yuan Li, for their great support with love during the period of this study.

## **Acknowledgements**

During my PhD study, I have learnt a lot from many people. It is with great pleasure that I acknowledge the assistance provided by people who supported and helped me to complete this thesis.

I would sincerely like to thank my supervisors, Dr. Cynthia D.K. Bottema and Assoc. Prof. Wayne Pitchford for their excellent supervision, guidance, trust and criticism over the course of my years of study. In particular, I would like to thank Dr. Bottema for encouraging me to continue the research journey. Dr Bottema is an excellent model as a multi-skilled, very knowledgeable and kind scientist, who led me to become a researcher and encouraged me to always strive to improve the quality of my project. Without her help, my life would be in a different path.

I am very grateful for the help from Assoc. Prof. Wayne. He was always willing to understand, discuss, listen, and encourage. The discussions on the analytical models were very beneficial in my understanding the complicated statistics. From his critical attitude, I gradually learned to criticise the research statistically.

To the laboratory members and my friends, Andrew Egarr, Irida Novianti, Nadiatur Akmar Zulkifli, David Lines, Leiyao Chang, Ruidong Xiang and Dr Graham Webb. Thank you all for the wonderful time we shared. My time in the laboratory would be boring without all your support, friendship and sharing the stories. I will always value your friendship and will remember the great and joyful times we have had forever.



Many thanks are extended to our collaborators, Chris Morris and Neil Cullen, for their useful comments on manuscripts and for their kind assistance with the animal sampling in New Zealand abattoirs. Thanks to Dr. Madan Naik for his assistance in learning various techniques in the molecular laboratory and his valuable advice during the course of my study. I would also like to acknowledge Dr. Zbigniew Kruk for his excellent pioneering work. The experience in the abattoir with Dr. Kruk was very valuable to help me to do the animal sampling in the abattoir by myself. My gratitude goes to Dr. Brian Siebert who dedicated much time and effort teaching me the techniques of  $\beta$ -carotene extraction.

Great thanks must go to the J.S. Davies Bequest for funding the cattle gene mapping project in Australia. My sincere thanks also extend to AgResearch in New Zealand for access to their data. They made a great contribution by providing the phenotype and genotype data for Groups 2, 4, 5 and 6. I also extend my sincere gratitude to the University of Adelaide for providing the PhD scholarship, without which this work was not possible.

My deepest gratitude goes to my mum and dad for being generous supporters over the past years. It was your love, support and strength that got me through the toughest time. Thank you for always being proud of my achievements. The moment when I finished typing the last word for my thesis, I can hear your voice, I can see your smile, and I can feel your love from thousands of miles away. This long distance cannot keep my love from you.

Most importantly, my special thanks must be given to my dear wife Yuan, who has

constantly motivated me and encouraged me in every way. I will never forget that you sacrificed your sleeping time to prepare breakfast for me every morning with your sleepy eyes. Thank for all your support even during the most emotionally challenging time. It is your understanding and your love that enabled me to complete this work.

## Abstract

Subcutaneous fat is important not only in the live animal but also in the carcass, as it prevents the rapid chilling of the underlying muscle tissues, thereby reducing weight loss during chilling. However, beef with yellow fat is considered undesirable by consumers in most European and Asian markets. Beta-carotene is the major carotenoid deposited in adipose tissue, which results in the yellowness. Genes involved in the metabolism of  $\beta$ -carotene in the cattle are likely to regulate beef fat colour. Therefore, DNA variants in candidate genes related to  $\beta$ -carotene metabolism were examined for association with beef fat colour.

Based on their location in fat colour quantitative trait loci (QTL) and function in the metabolism of  $\beta$ -carotene, *ALDH8A1*, *APOM*, *BCMO1*, *BCO2*, *RARA*, *RDHE2*, *PPARGC1A* and *SCARB1* were chosen as candidate genes. One hundred eleven (111) DNA variants were identified from the direct sequencing of 3 F1 sires for these eight genes, of which, 27 DNA single nucleotide polymorphisms (SNPs) were selected for association studies (3-5 SNPs per gene). Most of these genotyped SNPs and their interactions were associated with fat colour related traits (biopsy fat colour (Fc-bio), carcass fat colour (Fc-car), beta-carotene concentration (Bc-bio)), although the size of the effects was relatively low for many of the variants. However, among the DNA variants, a nonsense mutation in the *BCO2* gene (*BCO2* W80X) accounted for a large proportion (12-16%) of the total SNP variation in fat colour related traits in Jersey-Limousin backcross progeny. Validation of this SNP in other independent herds (Group 2-7) confirmed the *BCO2* W80X genotype has a large effect on beef fat colour and milk colour. The individual genotypic effects of *RDHE2* SNP2 and SNP3 were also large.

However, these effects were greater in the New Zealand abattoir samples than from pedigreed Jersey-Limousin backcross progeny, amounting to 8-17% of the variance in one population. There was a significant interaction between the *BCO2* W80X and the *RDHE2* SNP2, which accounted for 1.8% of the total SNP variance in milk fat colour in a New Zealand Holstein cow population, and 4.0% of the total SNP variance in carcass fat colour in New Zealand Jersey-Limousin backcross progeny.

In addition to the individual SNP effects, the effects of the haplotypes formed for each gene were also investigated. Only haplotypes of *BCMO1*, *PPARGC1A*, *RDHE2* and *SCARB1* genes had effects on beef fat colour.

The most likely pathways involved in the beef fat colour were clarified. The association studies showed that the *BCMO1*, *BCO2*, *RARA*, *RDHE2*, *PPARGC1A* and *ALDH8A1* genes and their interactions account for a large proportion of the variation in beef fat colour. These genes have roles involving retinol or retinoic acid synthesis. Therefore, the retinol/retinoic acid synthesis pathway appears to be the most important in terms of the contribution to the  $\beta$ -carotene concentration in adipose tissue. The effects of *APOM* and *SCARB1* indicate that transportation of  $\beta$ -carotene is also important in the regulation of the  $\beta$ -carotene concentration in fat.

Differences in the expression of *BCMO1*, *BCO2* and *RDHE2* genes were investigated. The *RDHE2* gene mRNA transcript level was significantly different between yellow fat and white fat samples. The gene expression of *BCO2* was highly correlated with  $\beta$ -carotene concentration. The results further support the role of *BCO2* in cleaving  $\beta$ -carotene eccentrically and the association of *RDHE2* with  $\beta$ -carotene concentration.

The results also indicate that the control of the retinol/retinoic acid pathway at the gene expression level is important for the  $\beta$ -carotene concentration in subcutaneous adipose tissue and consequently, for beef fat colour.

The study conducted herein contributes to the understanding of the metabolism of carotenoids and their numerous derivatives. *BCO2*-mediated conversion of  $\beta$ -carotene to vitamin A is confirmed in cattle. Epistatic effects accounted for much of the beef fat colour and  $\beta$ -carotene concentration variation. DNA variants that have a large influence on fat colour, such as the *BCO2* W80X, can be used in marker selection systems to rapidly reduce the incidence of yellow fat colour in beef.