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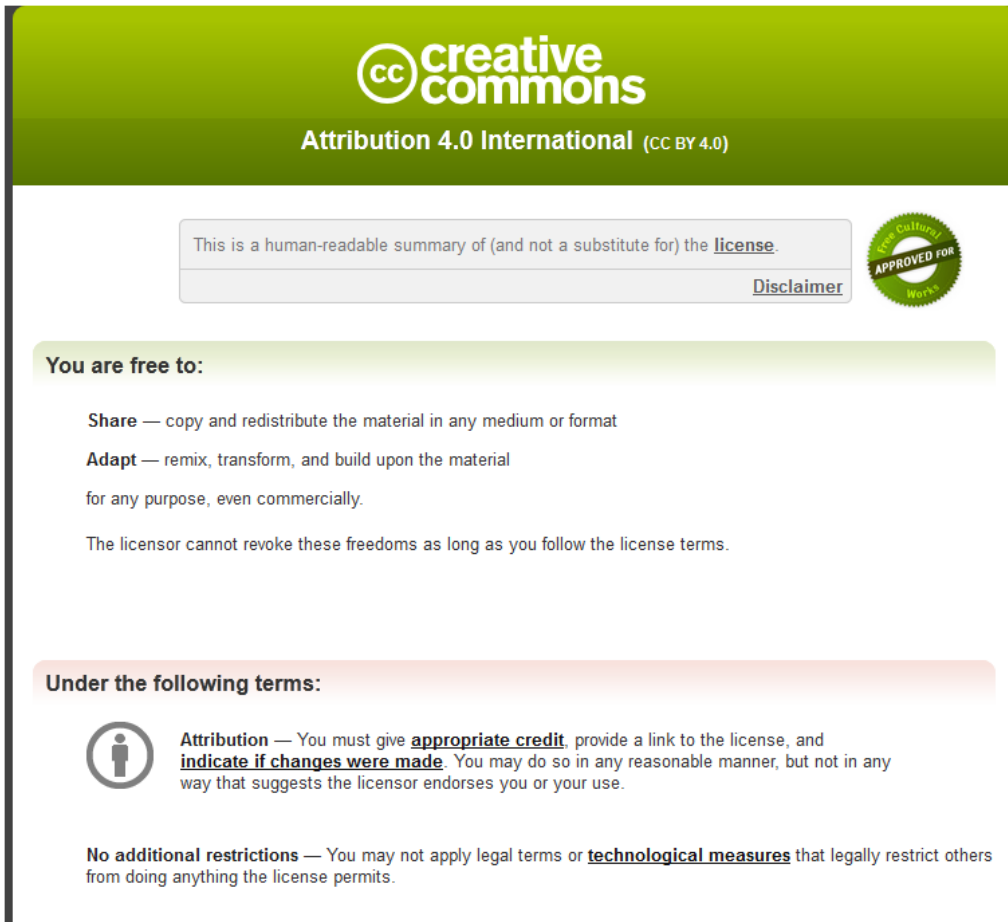
**A dual sensor for pH and hydrogen peroxide using polymer-coated optical fibre tips**  
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Article

# A Dual Sensor for pH and Hydrogen Peroxide Using Polymer-Coated Optical Fibre Tips

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**Abstract:** This paper demonstrates the first single optical fibre tip probe for concurrent detection of both hydrogen peroxide (H<sub>2</sub>O<sub>2</sub>) concentration and pH of a solution. The sensor is constructed by embedding two fluorophores: carboxyperoxyfluor-1 (CPF1) and seminaphtharhodafluor-2 (SNARF2) within a polymer matrix located on the tip of the optical fibre. The functionalised fibre probe reproducibly measures pH, and is able to accurately detect H<sub>2</sub>O<sub>2</sub> over a biologically relevant concentration range. This sensor offers potential for non-invasive detection of pH and H<sub>2</sub>O<sub>2</sub> in biological environments using a single optical fibre.

**Keywords:** optical fibre; hydrogen peroxide probe; pH sensor; dual sensor; fibre tip sensor

## 1. Introduction

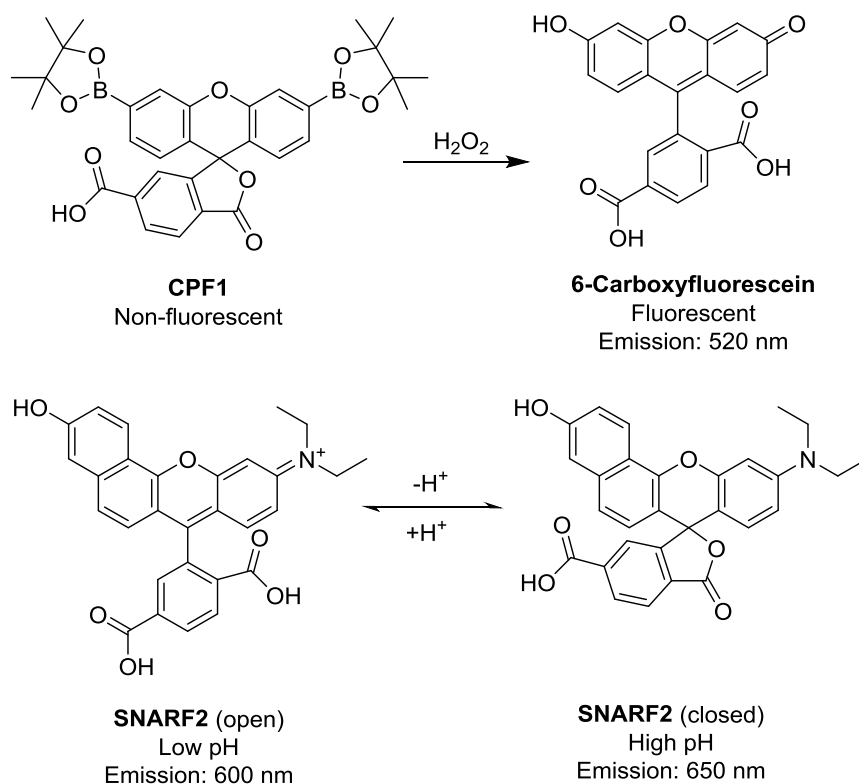
Hydrogen peroxide (H<sub>2</sub>O<sub>2</sub>) and pH play vital combined roles in cellular signalling [1–3], tumour development [4–7] and reproductive health science [8–11]. For example, the unregulated production of H<sub>2</sub>O<sub>2</sub> by an embryo is a hallmark of embryonic stress [12], while pH fluctuations during embryo culture can negatively affect embryonic development [13]. The simultaneous detection of pH and H<sub>2</sub>O<sub>2</sub> would therefore provide significant benefit in monitoring the associated cellular processes. H<sub>2</sub>O<sub>2</sub> and pH can be detected in cells by specific fluorophores, measuring either an increase in fluorescence intensity [14,15], or a change in emission spectra respectively [16,17]. However the use of these fluorescent probes in applications such as *in vitro* fertilisation (IVF) poses significant scientific and ethical questions, as their effect on the development of embryos is unknown. As such, direct contact of fluorophores with an embryo is ethically unsound and not allowable in most regulatory jurisdictions.

Optical fibre-based probes offer an attractive and non-invasive approach. Here a fluorophore of interest can be attached to the fibre surface for localised measurement without being released into the solution [18–20]. Various configurations of optical fibres have been examined for development of such fluorescent sensors, specifically; functionalized end-faces (tip sensors) [21,22], exposed core [23] and microstructured fibres [24,25]. Although microstructured fibre based sensors can be more

sensitive than tip sensors [26], filling of the air holes with analyte is required in order to perform a measurement. This typically restricts microstructured fibres to single temporal measurements, unless microfluidics or external flushing systems are employed, and these may impede on the cell culture environment. Exposed-core fibre sensors are ideal for environmental sensing and do not require microfluidics or external flushing and offer advantages in distributed sensing. However, tip-based sensors offer potential for temporal measurements in a single location rather than distributed along the length, or can be repositioned to obtain a spatial map of the sample as desired. Tip sensors often have reduced sensitivity compared to microstructured fibres [26], especially as conventional attachment of a single layer of fluorophore to a fibre tip results in a low signal intensity [27]. However, the signal intensity can be improved by increasing the density of fluorophore on the fibre tip.

$H_2O_2$  can be detected by aryl boronate-based fluorophores such as peroxyfluor-1 (PF1) [28] and carboxyPF1 (CPF1, Figure 1) [29]. These aryl boronates have been shown as particularly effective fluorescent probes for detection of  $H_2O_2$  in human spermatozoa and bovine oocytes [29,30]. pH can be detected using a range of fluorophores, with seminaphthorhodofluor-2 (SNARF2, Figure 1) offering some advantages over alternative probes, as the ratiometric emission from this probes changes its spectral features over the physiological pH range, with a  $pK_a$  of 7.5 [31]. This minimises potential errors which could arise from using a solely intensity-based probe. Additionally, its emission spectrum overlaps minimally with the emission of CPF1 [32], allowing the separate interrogation of each fluorophore.

This paper reports the first dual probe for sensing pH and the detection of  $H_2O_2$  by immobilising two separate fluorophores (CPF1 and SNARF-2) onto a single optical fibre tip in a polyacrylamide matrix. The two fluorophores are attached to a multi-mode fibre tip by a light-catalysed polymer coating [33], to allow for greater control of fluorophore surface density and thus subsequent signal intensity. This then allows detection of both  $H_2O_2$  and pH within a single system.



**Figure 1.** Chemical structures of fluorescent probes used in this study. Carboxyperoxyfluor-1 (CPF1) reacts with  $H_2O_2$  to form the fluorescent 6-carboxyfluorescein. Seminaphthorhodofluor-2 (SNARF2) is found in the protonated (open) form and lactone (closed) at low and high pH respectively.

## 2. Experimental Section

### 2.1. Materials

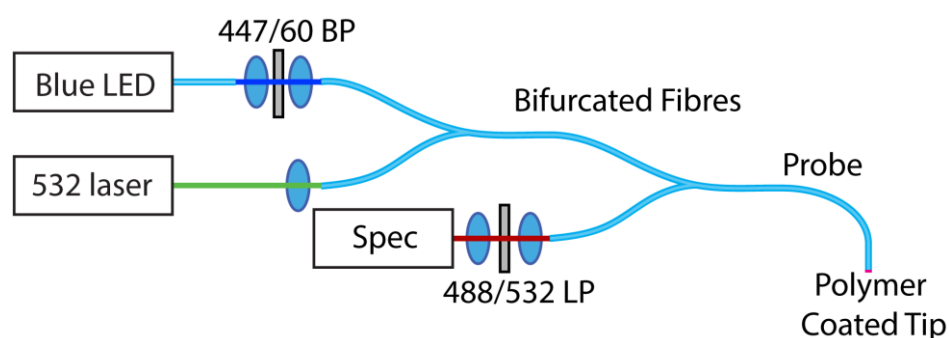
All chemicals were purchased from Sigma-Aldrich unless otherwise stated. Bis(acrylamide) was purchased from Polysciences (Warrington, PA, USA). HPLC grade acetonitrile was purchased from Scharlau. 100 mM Phosphate buffer solutions were prepared from monosodium phosphate and disodium phosphate in Milli-Q water. Multimode fibre (200  $\mu\text{m}$  core diameter, FG200UCC) was purchased from Thorlabs (Newton, NJ, USA), with one end connectorised for attachment to the optical setup.

### 2.2. Polyacrylamide Photo-Polymerisation on Optical Fibre Tips

A solution of 3-(trimethoxysilyl)propyl methacrylate (5  $\mu\text{L}$ ) in 10% acetic acid solution (30  $\mu\text{L}$ ) and ethanol (1 mL) was mixed and sonicated until clear. Multi-mode fibre was cleaved and each segment immersed in the methacrylate solution for 1 h. The fibre tip was then dried under  $\text{N}_2$ , rinsed with Milli-Q water and re-dried under  $\text{N}_2$ . The distal end of the fibre was coupled into a 405 nm source (Crystalaser 405 nm) using a  $10\times$  microscope objective. A monomeric stock solution comprising of 3% bisacrylamide, 27% acrylamide and 70% pH 6.5 phosphate buffer solution was dissolved under sonication. CPF1-NHS (0.2 mg), and SNARF2-NHS (0.2 mg) were added to this solution (400  $\mu\text{L}$ ) and 200  $\mu\text{L}$  of the resulting solution was pipetted into a small Eppendorf tube. Triethylamine (10  $\mu\text{L}/\text{mL}$ ) was added to the mixture, and the fibre tip was immersed in this solution exactly 60 s after addition of the triethylamine and immediately irradiated under 405 nm light for 10 s at 13.4 mW, to form a polymeric coating on the fibre tip.

### 2.3. Optical Measurements

The experimental configuration used for optical measurements is shown in Figure 2 below.



**Figure 2.** Experimental configuration for optical measurements of the combined pH/peroxide sensor. The blue LED source was used to illuminate the peroxide sensitive carboxyperoxyfluor-1 (CPF1) fluorophore, while the green excites the pH sensitive seminaphtharhodafuor-2 (SNARF).

A 470 nm blue LED source (Thorlabs M470F1, Newton, NJ, USA) with an appropriate bandpass filter (Semrock Brightline 447/60, Rochester, NY, USA) was coupled into one input of a bifurcated fibre (Ocean Optics 200  $\mu\text{m}$ , UV/VIS). Attached CPF1 was then excited with light from a 532 nm green laser (Crystalaser 25 mW, Reno, NV, USA) coupled into the other input for excitation of the SNARF. An additional bifurcated fibre was used to connect the excitation sources to the sensing fibre, with the remaining input connected to the input of the spectrometer (Horiba iHR550, Synapse detector, Kyoto, Japan). Long-pass filters were inserted directly into the spectrometer input cage, with 488 nm (Semrock 488 nm Edgebasic) and 532 nm (Semrock 532 nm Razoredge) used for peroxide and pH respectively.

The two excitation channels were controlled independently, with only one excitation wavelength used at any particular time to excite either the peroxide or pH channel. The corresponding emission

filter was used with each excitation source to attenuate residual pump light from the fibres. The use of connectorised fibres and multi-mode fibres greatly simplifies the measurement procedure, as no adjustments or realignment are required when swapping between pH and peroxide measurements.

### 3. Results and Discussion

#### 3.1. Hydrogen Peroxide Detection

##### 3.1.1. Detection of Biologically Relevant H<sub>2</sub>O<sub>2</sub> Concentrations

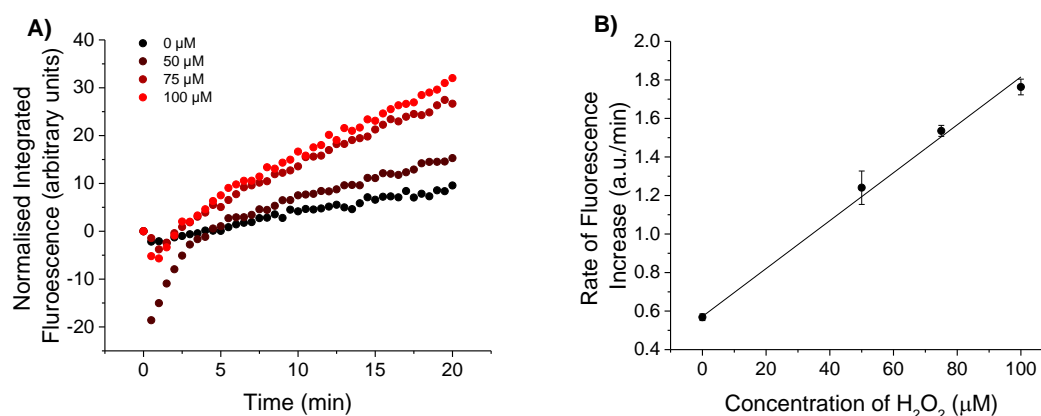
CPF1 and SNARF2 immobilised on fibre tips were tested in solutions containing H<sub>2</sub>O<sub>2</sub> to establish the sensitivity of this surface configuration. Fibre tips were first functionalised with 3-(trimethoxysilyl)propyl methacrylate, then dipped into a solution of acrylamide/bisacrylamide with N-hydroxysuccinimide esters of CPF1 and SNARF2. The N-succinimide esters of CPF1 and SNARF2 increase solubility in the acrylamide solution to provide a more reproducible density of fluorophores embedded in the polymer matrix. Excitation light (405 nm) was coupled into the distal end of the fibre and the tip irradiated for 10 s to form a polymer layer on the tip containing the fluorophores. The functionalised fibres were dipped into a range of concentrations of H<sub>2</sub>O<sub>2</sub> (0, 50, 75 and 100 µM) in pH 7.5 phosphate buffer, and the emission peaks from CPF1 at 520 nm and SNARF at 600 and 660 nm were observed under 473 nm excitation. A low excitation power was used (27 µW) for these trials to minimise any potential effects of photobleaching. The entire spectrum was then integrated and normalised to the initial peak of CPF1 at 520 nm. This was necessary because each probe has slightly different initial fluorescence values and hence raw intensity values cannot be directly compared. Furthermore, each fibre probe was only used once for detection of H<sub>2</sub>O<sub>2</sub> to ensure maximum consistency between trials. Figure 3 shows an increase in normalised integrated fluorescence due to CPF1 over a 20 min exposure to H<sub>2</sub>O<sub>2</sub>. This time interval was dictated by the reaction rate of aryl boronates such as CPF1 with H<sub>2</sub>O<sub>2</sub> [34].

Normalised fluorescence of CPF1 in the presence of 100 µM H<sub>2</sub>O<sub>2</sub> is greater than for the control, which lacked H<sub>2</sub>O<sub>2</sub> (Figure 3A). This increase in fluorescence is consistent with CPF1 reacting with H<sub>2</sub>O<sub>2</sub> on the fibre tip. Furthermore, a plot of the rate of increase in fluorescence vs concentration of H<sub>2</sub>O<sub>2</sub> (Figure 3B) clearly shows this rate increasing as the concentration of H<sub>2</sub>O<sub>2</sub> increases from 0 µM to 50 µM, 75 µM and 100 µM of H<sub>2</sub>O<sub>2</sub>. A similar increase was observed in our previous studies on the detection of H<sub>2</sub>O<sub>2</sub> with CPF1 in solution [27], and we have also shown that CPF1 is able to detect relevant H<sub>2</sub>O<sub>2</sub> concentrations in reproductive biology [29]. Hence, this probe exhibits sufficient sensitivity for this biological environment.

An initial drop in fluorescence was observed for some probes (see Figure 3A), particularly the probe immersed in a 50 µM H<sub>2</sub>O<sub>2</sub> solution. This is likely due to a change in emission properties of the fluorophores as the probe is moved from air into the solution. A more rapid increase in fluorescence was observed in the first 5 min, suggesting the probe is equilibrating in the new medium. After 5 min the probes show a near linear increase in fluorescence intensity. Thus, an incubation time of greater than 5 min is required to give an accurate indication of the rate of increase in fluorescence due to H<sub>2</sub>O<sub>2</sub>. A plateau was not observed over the time course of the experiment, suggesting quantitative data should be obtained from the rate of increase in fluorescence rather than the overall increase in fluorescence.

It is also important to note that an increase in the integrated fluorescent signal was evident, even in the absence of H<sub>2</sub>O<sub>2</sub> (Figure 3A). A decrease in fluorescence would be expected if photobleaching or leaching of the fluorophore from the polymer occurred over the course of the experiment. The effect of photobleaching on CPF1 was of particular interest, since CPF1 is oxidised by H<sub>2</sub>O<sub>2</sub> to give 5-carboxyfluorescein. 5-carboxyfluorescein is known to photobleach [35], a process which occurs more rapidly in the presence of reactive oxygen species such as H<sub>2</sub>O<sub>2</sub> [36]. In order to accurately sense H<sub>2</sub>O<sub>2</sub> with CPF1, low rates of photobleaching must be achieved. An increase in fluorescence in the absence of H<sub>2</sub>O<sub>2</sub> suggests that photobleaching was not occurring on the fibre during the experiment.

This is an important observation for the practical use of the probe since the fibre's ability to sense accurately would be reduced by photobleaching of the fluorophores on the fibre tip.

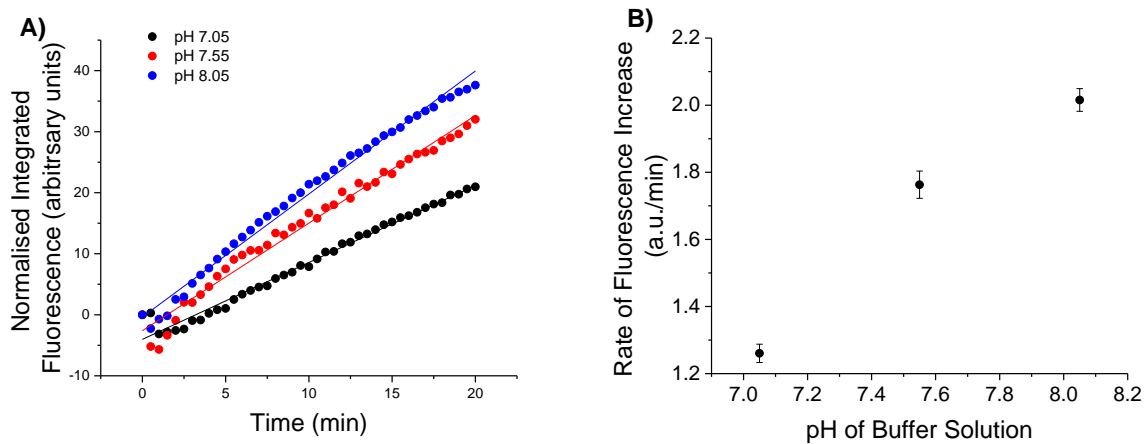


**Figure 3.** (A) Integrated fluorescence intensity from CPF1 using blue excitation with varied peroxide concentration in pH 7.5 buffer; 100 μM H<sub>2</sub>O<sub>2</sub> shows an increased fluorescent response from the fibre without H<sub>2</sub>O<sub>2</sub>; (B) Slope of integrated fluorescence for increasing concentrations of H<sub>2</sub>O<sub>2</sub> (0, 50, 75 and 100 μM). Error bars indicate the standard error of the calculated slope.

### 3.1.2. Effect of Change in pH on Detection of H<sub>2</sub>O<sub>2</sub>

The effect of pH on the detection of H<sub>2</sub>O<sub>2</sub> in fibre was next investigated by immersing functionalised fibre tips in solutions of H<sub>2</sub>O<sub>2</sub> of differing pH. Fibre tips were functionalised with CPF1 and SNARF2 as before and separately immersed in 100 μM separate solutions of H<sub>2</sub>O<sub>2</sub> at a pH of 7.05, 7.55 or 8.05. A 470 nm light source was coupled into the fibre for excitation and the increase in fluorescence was recorded over 20 min. All spectra were integrated and normalised as before, with the results shown in Figure 4A. The rate of increase in fluorescence was calculated as depicted in Figure 4B. The observed rate at pH 8.05 was 1.5 times greater than the rate observed at pH 7.05 (Figure 4B).

The reaction rate of aryl boronates (such as CPF1) with H<sub>2</sub>O<sub>2</sub> is higher in more basic solutions [37]. CPF1 reacts with the conjugate base of H<sub>2</sub>O<sub>2</sub> (hydroperoxide ion HOO<sup>−</sup>) [15] to give fluorescent 5-carboxyfluorescein. As the pH increases, more H<sub>2</sub>O<sub>2</sub> dissociates into its conjugate base, HOO<sup>−</sup>. The concentration of HOO<sup>−</sup> available to react with CPF1 will therefore be higher in more basic solutions, accounting for the observed upward trend in rates due to increasing pH (Figure 4B). Furthermore, the product of CPF1 with H<sub>2</sub>O<sub>2</sub> is a carboxyfluorescein, and fluorescein exhibits different quantum yields of fluorescence at differing pH [38]. Thus, the pH of the solution is highly pertinent to the accurate detection of H<sub>2</sub>O<sub>2</sub>. This is further highlighted by a comparison of rates of increase in fluorescence at different pH and H<sub>2</sub>O<sub>2</sub> concentration, shown in Figures 3B and 4B. A rate of approximately 1.2 a.u./min was calculated for a 100 μM solution of H<sub>2</sub>O<sub>2</sub> in pH 7.05 (Figure 4B). However, a similar rate was calculated for a 50 μM solution of H<sub>2</sub>O<sub>2</sub> at a pH of 7.55 (Figure 3B). The observed rate in these experiments is clearly dependent on the pH, in addition to the concentration of H<sub>2</sub>O<sub>2</sub>. It is hence necessary that the pH of a solution must be known in order to calculate an unknown concentration of H<sub>2</sub>O<sub>2</sub> resulting from an increase in fluorescence. Therefore, it is critical that the probe also incorporates a pH sensitive component to accurately determine the H<sub>2</sub>O<sub>2</sub> concentration. This improves upon many systems for detection of H<sub>2</sub>O<sub>2</sub> that do not simultaneously measure pH, despite the reported effect on H<sub>2</sub>O<sub>2</sub> detection [15].

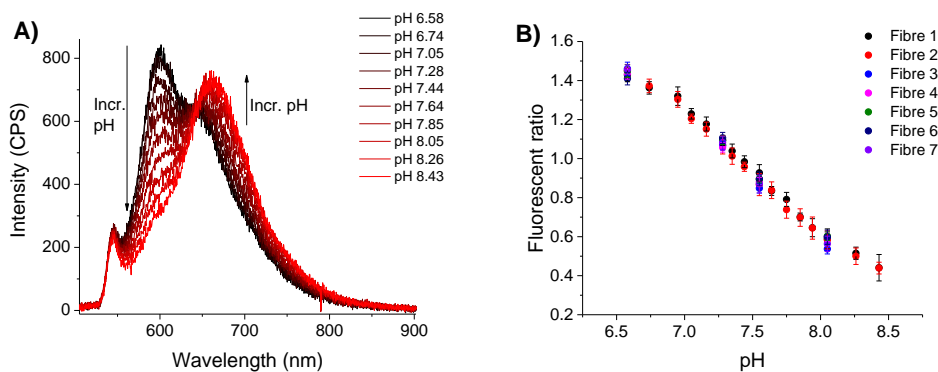


**Figure 4.** Response of CPF1 to 100  $\mu\text{M}$   $\text{H}_2\text{O}_2$  in solutions that varied in pH. (A) Integrated fluorescent responses of probes to 100  $\mu\text{M}$   $\text{H}_2\text{O}_2$  in pH 7.05, 7.55 and 8.05 over 20 min using blue excitation; (B) Rate of increase in fluorescence of each probe with  $\text{H}_2\text{O}_2$  in each of the 3 pH solutions. Error bars indicate the standard error of the calculated slope.

### 3.2. pH Sensing

#### 3.2.1. Initial pH Sensing

The sensitivity of these functionalised fibre tips was defined across a series of solutions of differing pH, ranging from 6.5 to 8.5. Fibres functionalised with CPF1 and SNARF2 as before were dipped into phosphate buffer solutions of each pH. 532 nm light attenuated to 13  $\mu\text{W}$  was coupled into the fibre for excitation, and the fluorescent signal from immobilised SNARF2 was collected after 1 min equilibration time. The fibre was removed from solution, dried, and immersed in a subsequent buffer solution. Two fibre probes were calibrated in this way using sixteen buffer solutions ranging from pH 6.5 to 8.5 as shown in Figure 5. This broad pH range (6.4–8.5) was chosen in order to demonstrate the potential of the probe in biological applications beyond the narrower constraints of an embryo. Five additional probes were then calibrated in selected solutions across this range (see fibres 3–7 in Figure 5B). The fluorescence spectra were recorded as shown in Figure 5A. The probe exhibits a decrease in intensity of fluorescence at 600 nm as the pH increases, with an increase in intensity at 660 nm.

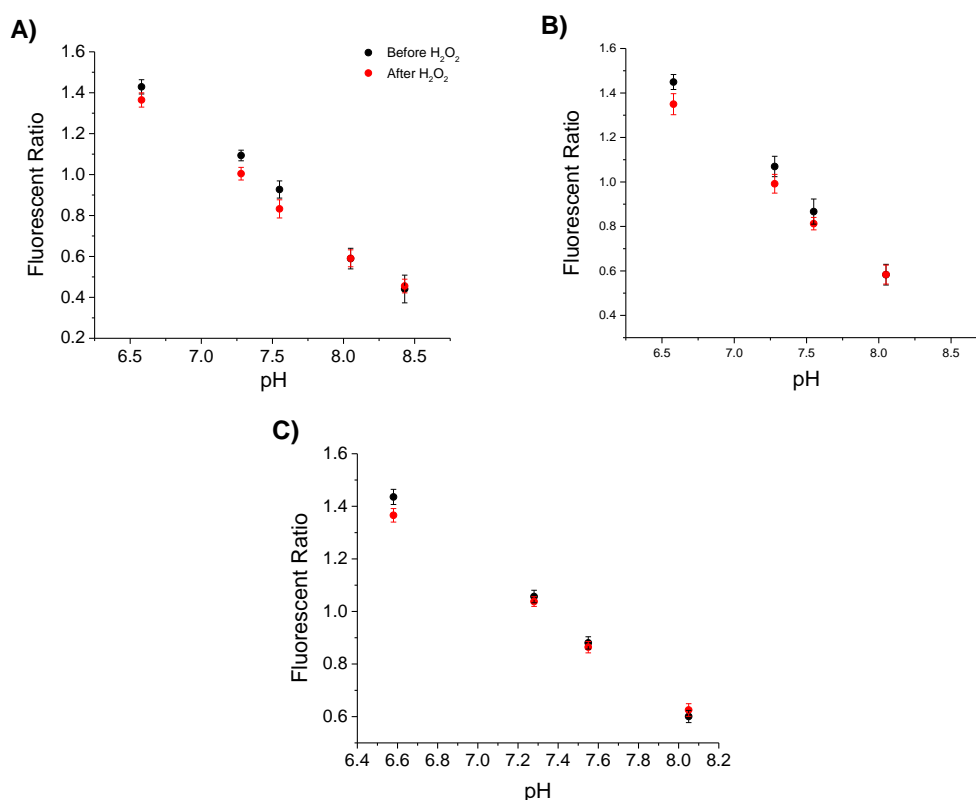


**Figure 5.** pH response of SNARF-2 embedded in polyacrylamide on fibre tip to varied pH. (A) Emission spectra of SNARF in various pH buffers; (B) Ratio of emission peak intensities 600/660nm shown with over multiple trials. The effect of noise was reduced by taking the mean of eight values between 598–602 nm and 558–662 nm. Error bars represent the standard deviation of these values.

The pH of the buffer was correlated with the observed fluorescent signal by calculating the ratio of intensities at 600 nm and 660 nm for each spectrum (Figure 5B). This analysis revealed an inverse correlation between this fluorescent ratio and pH of the solution. The plot also shows a linear trend over the pH range 7.0 to 8.0 and indicates that the sensor should be of use for determining the pH of a biological sample near physiological pH. Moreover, multiple fibre trials produced similar pH calibration curves (Figure 5B) showing good reproducibility between fibres. This data also indicates that SNARF2 bound to a fibre tip behaves as in solution [32]. Therefore, SNARF2 effectively senses the pH of a buffer solution when embedded in polyacrylamide on a fibre tip.

### 3.2.2. pH Sensing before and after Detection of Hydrogen Peroxide

The functionalised fibre probes were used to sense pH before and after immersion in  $H_2O_2$ , in order to determine if this affected the pH sensing capability. Each fibre was functionalised with CPF1 and SNARF2 and calibrated in phosphate buffer solutions of known pH as described in Section 3.2.1. The fibre tips were then immersed for 20 min in one of three  $H_2O_2$  solutions: 100  $\mu M$  solution of  $H_2O_2$  in pH 7.55 buffer, 50  $\mu M$   $H_2O_2$  in pH 7.55 buffer or 100  $\mu M$   $H_2O_2$  in pH 7.05 buffer. These conditions represent the range of conditions that the probe may experience in unknown samples. Each probe was calibrated again in phosphate buffer solutions as above. The resulting pH calibration curves were plotted for each probe both before and after immersion in  $H_2O_2$  (Figure 6).



**Figure 6.** Sensing of pH before and after immersion in  $H_2O_2$ . Each graph plots the ratio of emission peaks of SNARF2 at 600/660 nm with the pH of the buffer solution tested, before and after solutions: (A) 100  $\mu M$  solution of  $H_2O_2$  in pH 7.55 buffer; (B) 50  $\mu M$   $H_2O_2$  in pH 7.55 buffer; (C) 100  $\mu M$   $H_2O_2$  in pH 7.05 buffer. Three different samples were trialled before and after  $H_2O_2$  solutions to demonstrate the independence of the result to concentration of  $H_2O_2$  or the pH of the  $H_2O_2$  solution. To reduce any effect of noise, the mean of eight values between 598–602 nm and 558–662 nm is given. Error bars represent the standard deviation of these values.



The fluorescent ratios shown in Figure 6A show minimal changes before and after immersion in 100  $\mu\text{M}$   $\text{H}_2\text{O}_2$  in pH 7.55. This indicates that reaction of  $\text{H}_2\text{O}_2$  with CPF1 does not affect the sensing of pH by SNARF2. Figure 6B reveals similar fluorescent ratios before and after immersion in 50  $\mu\text{M}$   $\text{H}_2\text{O}_2$  in pH 7.55 buffer.  $\text{H}_2\text{O}_2$  again does not affect the sensing of pH over the tested concentration range, 50  $\mu\text{M}$  to 100  $\mu\text{M}$   $\text{H}_2\text{O}_2$ . Figure 6C shows the fluorescent ratios of SNARF on fibre tips before and after immersing in 100  $\mu\text{M}$   $\text{H}_2\text{O}_2$  in pH 7.05 buffer. As per the previous results, the fluorescent ratio curves did not change significantly after immersion in  $\text{H}_2\text{O}_2$ . Importantly, immersing the probe into a solution containing  $\text{H}_2\text{O}_2$  does not affect the pH sensing capability of the probe. This demonstrates that SNARF2 can be used on optical fibre tips for sensing pH independent of the detection of  $\text{H}_2\text{O}_2$  by CPF1.

#### 4. Conclusions/Outlook

The tip of an optical fibre has been functionalised with two separate fluorophores, CPF1 and SNARF2 embedded in polyacrylamide, in order to allow measurement of the  $\text{H}_2\text{O}_2$  concentration and pH respectively. The probe is demonstrated to effectively detect  $\text{H}_2\text{O}_2$  over a physiological pH range. The probe shows a minimum detectable concentration of 50  $\mu\text{M}$   $\text{H}_2\text{O}_2$  at pH 7.5, and pH was measured repeatedly over the range 6.5–8.5 with resolution of 0.1 pH units. Each fluorophore was used in tandem by alternating excitation sources, *i.e.*, blue excitation to interrogate CPF1 for  $\text{H}_2\text{O}_2$  detection, and green excitation for SNARF2 to sense pH, with minimal cross-talk. The combination of pH and  $\text{H}_2\text{O}_2$  detection also addressed the crucial issue of accurate measurement of  $\text{H}_2\text{O}_2$  in solutions with varying or unknown pH, where the pH of the solution alters the apparent  $\text{H}_2\text{O}_2$  concentration. This is the first example of a dual pH and  $\text{H}_2\text{O}_2$  probe and is an important proof of concept for the detection of pH and  $\text{H}_2\text{O}_2$  in ethically complex biological environments such as found in an IVF laboratory.

This probe could find potential application if placed near the cumulus cells of an oocyte for monitoring of extracellular pH and  $\text{H}_2\text{O}_2$  fluxes during fertilisation and during early embryonic development. Tapering of the fibre tip could also increase the resolution from 200  $\mu\text{m}$  to a few microns if required [39]. This fibre probe may offer potential not only in embryology, but a range of biological applications whereby the system must remain isolated from any external agents such as organic fluorophores.

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**Author Contributions:** M.S.P., E.P.S., T.M.M. and A.D.A. conceived and designed the experiments; J.G.T. provided biological specifications for the probe; M.S.P. performed the experiments and analysed the data; E.P.S. and A.D.A. assisted with analysis; M.S.P. wrote the paper with contributions from E.P.S., A.D.A., J.G.T. and T.M.M.

**Conflicts of Interest:** The authors declare no conflict of interest.

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