Mechanism of Bevacinzumab Adsorption with Affinity Ligands And Bioprocess Optimization For Antibody Purification

A THESIS SUBMITTED

BY

Yuzhe Tang

FOR THE DEGREE OF

MASTER OF ENGINEERING SCIENCE

School of Chemical Engineering

The University of Adelaide

Adelaide, Australia

Declaration

I certify that this work contains no material which has been accepted for the award of any other

degree or diploma in any university or other tertiary institution and, to the best of my

knowledge and belief, contains no material previously published or written by another person,

except where due reference has been made in the text. In addition, I certify that no part of this

work will, in future, be used in submission for any other degree or diploma in any university

or tertiary institution without the prior approval of the University of Adelaide and where

applicable, any partner institution responsible for the joint-award of this degree

I give consent to this copy of my thesis, when deposited in the University Library, being made

available for loan and photocopying, subject to the provisions of the Copyright Act 1968

I also give permission for the digital version of my thesis to be made available on the web, via

the University's digital research repository, the Library catalogue and also through web search

engines, unless permission has been granted by the University to restrict access for a period of

time

Signature:

Date:

-1-

Acknowledgments

There are many people I would like to thank, who have helped to make this work possible. First and foremost, I would like to my supervisor Associate Professor Jingxiu Bi and Dr Hu Zhang (School of Chemical Engineering, University of Adelaide) for their support physically and psychotically over the past two years. To Associate Professor Sheng Dai (School of Chemical Engineering, University of Adelaide), his knowledge background has made each of his suggestion becomes my turning point. I would like to thank Sansom Research Institute (University of South Australia) to provide me the lab access of the thermoanalysis equipment. In the end, I would like to thank my family, you always there for me. I would like to give special thanks to my dad, without you I would never have been able to do this. And to my mum, your support is what kept me going

Thank you all

Abstract

Monoclonal antibodies (mAbs) have been found with a wide array of applications as pharmaceutical compounds in the treatment of cancers and diseases such as arthritis, asthma and osteoporosis. In approximate 10 years retrospection, the global market of mAbs experienced a rapid growth, nearly tripling the profit to be approximate US\$16.7 billion in 2014. In order to meet the rising demand for mAbs, it is critical for manufacturers to ensure the production efficiency on the premise of product quality assurance. Especially in downstream purification of mAbs, the affinity chromatography as the major capture stage acts crucially in the removal of contaminates including host cell protein (HCP), DNA, antibody variants, viral particles and endotoxin to obtain rapid isolation and high concentration of the target protein. However, drawbacks associated with this technique are the expense of resins for binding mAbs. To reduce the cost, alternative resins have been explored. However, this raises the significance of understanding the mechanism of ligand-mAb binding in terms of binding sites and binding conformational changes for the optimisation of chromatography performance.

To address the aforementioned binding mechanism, the isothermal titration calorimetry (ITC) method was employed for investigation of the thermal dynamic behaviour during free ligand and mAb binding. Two widely used affinity ligands, native Protein A (nSpA) and MabSelect SuRe (MS) ligand, were selected to bind with Bevacinzumab (BmAb). The binding mechanism was determined based on the isothermal parameters such as binding associated coefficient (ka), binding associated enthalpy changes (Δ H) and entropy changes ($T\Delta$ S).

Further investigations were carried out by applying BmAb into the affinity columns packed with nSpA or MS ligands to evaluate mAb association and disassociation with immobilized ligands at different operational conditions. It was found that the binding breakthrough curves

are related to the mAb association that reveals distinctive dynamic binding capacities and column binding performance.

Based on above studies, it was found that the binding conformation and binding affinity were different between the native Protein A and the recombinant MabSelected SuRe ligand. The formation of ligand-BmAb binding complex was examine d under various conditions such as pH, temperature and solvent ionic strength. In the end, binding mechanism was understood by the analysis of above conditions in both ITC and Binding breakthrough studies.

Table of Contents

Chapter 1 Introduction	9 -
1.1 Introduction	9 -
1.2 Research Scope	11 -
Chapter 2 Literature Review	12 -
2.1 Monoclonal antibody	12 -
2.2 Bevacinzumab	13 -
2.3 Downstream monoclonal antibody purification process	15 -
2.4 Chromatography	16 -
2.4.1 Affinity chromatography	16 -
2.5 Protein-Ligand adsorption of SpA and Immunoglobulin	18 -
2.5.1 Interaction of Immunoglobulin Fab region	19 -
2.5.2 Interaction of immunoglobulin Fc region	21
2.6 Combinatorial SpA domain Z	23
2.7 Effects to the protein-ligand adsorption in chromatography	25
2.7.1 Ionic strength	25
2.7.2 pH	26
2.7.3 Ligand spacer arm	26
2.7.4 Pore size of pack bed	27
2.8 ITC study in Protein-ligand interaction	27
Chapter 3 Isothermal Titration Calorimetry Study on BmAb-ligand Interactions	32
3.1 Introduction	32
3.2 Material and methods	33
3.2.1 Chemicals and reagents	33
3.2.2 Buffer exchange and protein concentration determination	34
3.2.3 ITC analysis	34
3.3 Results and discussion	36
3.3.1 The ITC assay	36
3.3.2 Effect of temperature	38
3.3.3 Effect of ionic strength	44
3.3.4 Effect of pH	48
3.4 Conclusion	53
Chapter 4 Breakthrough study of BmAb dynamic binding to immobilised ligands	54
4.1 Introduction	5.4

4.2 Materials and Methods	55
4.2.1 Materials	55
4.2.2 Determination of protein concentration	56
4.2.3 BmAbs chromatographic binding breakthrough	56
4.3 Experimental Results of Break-through study of Protein A	58
4.3.1 Effect of Ionic strength in binding solution	60
4.3.2 pH	63
4.3.3 Temperature	67
4.4 Conclusion	71
Chapter 5 Conclusions and Recommendations	72
5.1 Conclusions	72
5.2 Recommendations	73
References	74

List of Figures

Figure 1 Molecular Simulation structure of Bevacinzumab (Wragg and Bicknell, 2013)14 -
Figure 3 Interaction of individual SpA domains to Fab and Fc, residues involved involved in binding
with Fab are highlighted in Cyan, and Fc are highlighted in gray, Fln-32 is in pink (Graille et al., 2000)
Figure 4 Three possible docking conformational clusters between B domain and Fc of IgG, coloured
in magenta, yellow and dark blue respectively (Branco et al., 2012)22
Figure 5 Consensus binding sites to Fc target, diagonal lines indicates the Hydrogen bonding sites,
shaded area is for hydrophobic interaction, and circles are salt bridges (left). Protein A domain B
binding sites to IgG, (2) (5) hydrogen bonding, (3) (4) (6) hydrophobic interaction (right) (DeLano et
al., 2000)
Figure 6 Peptide sequences of natural SpA domains (E, D, A, B, C) and domain Z. A dash (-) means
excact amino acid sequence in comparing with B domain, and Red circle indicates the only change
between B and Z domain (Jansson et al., 1998)
Figure 7 Relative binding activity of six SpA Fc domains (A) and human polyclonal F(ab') (B)
(Jansson et al., 1998)
Figure 8 Thermodynamic parameters for the binding of CytC and mAb 5F8 at temperature gradient
from 270K to 310K (Pierce et al., 1999)
Figure 9 a) The net enthalpy changes of 0.1%, 0.2% and 0.3% BSA at dissociation by the adddition of
NaOH, b) the net enthalpy changes at the dissociation as the function of pH (Kun et al., 2009)30
Figure 10 The adsorption of enthalpy (Δ Hads) of myoglobin with a) butyl-Sepharose b) octyl-
Sepharose at various (NH4)2SO4 concentrations (Tsai et al., 2002)
Figure 11 A typical Isothermal Titration Calorimeter (Pierce et al., 1999)
Figure 12 Thermogram (top) and binding isotherm (bottom) for the interaction between native Protein
A and Bevacinzumab
Figure 13 Effect of binding temperature to thermo-parameters (a) LogKa and (b) ΔG K and ΔG were
derived from the isothermal titration curves of Protein A and BmAb as affinity ligand42
Figure 14 Effect of binding temperature to thermo-parameters (a) ΔH and (b) $T\Delta S$ ΔH and ΔS were
derived from the isothermal titration curves of Protein A and BmAb as affinity ligand
Figure 15 Effect of ionic strength in binding solution to thermo-parameters (a) LogKa and (b) ΔG K
and ΔG were derived from the isothermal titration curves of Protein A and BmAb as affinity ligand46
Figure 16 Effect of ionic strength in binding solution to thermo-parameters (a) ΔH and (b) $T\Delta S$, ΔH
and ΔS were derived from the isothermal titration curves of Protein A and BmAb as affinity ligand 47
Figure 17 Efffect of pH in binding solution to thermo-parameters (a) LogKa and (b) ΔG ,K and ΔG
were derived from the isothermal titration curves of Protein A and BmAb as affinity ligand
Figure 18 Effect of pH in binding solution to thermo-parameters (a) ΔH and (b) TΔS, ΔH and ΔS
were derived from the isothermal titration curves of Protein A and BmAb as affinity ligand
Figure 19 AKTA Pure scheme 57
Figure 20 HiTrap Protein A 1mL breakthrough by loading BmAb at pH 6
Figure 21 Effect of solvent ionic strength on loading BmAb to a) HiTrap Protein A and b) HiTrap
MabSelect SuRe via various NaCl concentration in mobile phase, (Black) 100mM NaCl, (Red)
500mM NaCl and (Blue) 1M NaCl
Figure 22 Effect of pH on loading BmAb to a)HiTrap Protein A and b) MabSelect SuRe via various
pHs in mobile phase, (Black) pH 7, (Red) pH 6, (Blue) pH 5, and (Green) pH 466

Figure 23 Effect of temperature on Loading BmAb to a)HiTrap Protein A and b) MabSelect various temperatures, (Black) 25°C and (Red) 4°C	
List of Tables	
Table 1 Hill slop (H) and EC50 by loading BmAb to HiTrap Protein A and MabSelect SuRe	columns
at various buffer salt concentrations	61
Table 2 Hill slop (H) and EC50 by loading BmAb to HiTrap Protein A and MabSelect SuRe	columns
at various buffer pHs	65
Table 3 Hill slop (H) and EC50 by loading BmAb to HiTrap Protein A and MabSelect SuRe	columns
at various temperatures	69